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SENNIGER, POWERS, LEAVITT & ROEDEL
ATTORNEYS AT LAW

ONE METROPOLITAN SQUARE
16TH FLOOR
ST. LOUIS, MISSOURI 63102
314-231-5400

FACSIMILE 314-231-4342
http://www.senniger.com

August 30, 2000

UTILITY PATENT APPLICATION TRANSMITTAL
(new nonprovisional applications under 37 CFR 1.53(b))

Attorney Docket Number: MTC 6718
First Named Inventor: Michael Lassner
Express Mail Label Number: EL615274183US

TO: Assistant Commissioner for Patents
Box Patent Application
Washington, D.C. 20231

APPLICATION ELEMENTS

1. [X] Fee Transmittal Form
(original and duplicate)

2. [X] Pages of Application [Total Pages 73]

3. [X] Drawings [Total Sheets 7]

4. Oath or Declaration [Total Pages 4]

a. [ ] Newly executed (original or copy)
[X] New (unexecuted)

b. [ ] Copy from a prior application
(for continuation/divisional with
Box 17 completed)
i. [ ] DELETION OF INVENTOR(s)
Signed statement attached
deleting inventor(s) named
in prior application.

5. [ ] Incorporation By Reference
(useable if Box 4b is marked)
The entire disclosure of the prior application, from which a copy of the oath or declaration is supplied under Box 4b, is considered as being part of the disclosure of the accompanying application and is hereby incorporated by reference therein.

6. [ ] Microfiche Computer Program (Appendix)

7. [X] Nucleotide and/or Amino Acid Sequence Submission (if applicable, all necessary)
   a. [X] Computer Readable Copy
   b. [X] Paper Copy (identical to computer copy)
   c. [X] Statement verifying identity of above copies

ACCOMPANYING APPLICATION PARTS

8. [ ] Assignment Papers (cover sheet & document(s))

9. [ ] 37 CFR 3.73(b) Statement [ ] Power of Attorney

10. [ ] English Translation Document (if applicable)

11. [ ] IDS with PTO-1449 [ ] Copies of IDS Citations

12. [ ] Preliminary Amendment

13. [X] Return Receipt Postcard

14. [ ] Small Entity Statement(s)
    [ ] Statement filed in prior application; status still proper and desired

15. [ ] Certified Copy of Priority Document(s) if foreign priority is claimed

16. [ ] Other: _____

IF A CONTINUING APPLICATION, CHECK APPROPRIATE BOXES AND SUPPLY THE REQUISITE INFORMATION

17. [ ] Continuation [ ] Divisional [ ] Continuation-in-Part of prior application No.: _____

    [X] Complete Application based on provisional Application No. 60/152,493 filed August 30, 1999

CORRESPONDENCE ADDRESS

18. Correspondence Address: Customer Number 321
    Attention: James E. Butler, Ph.D.

Respectfully submitted,

James E. Butler, Reg. No. 40,931

JEBmkd

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Application Number Not Yet Assigned
Filed Herewith
First Named Inventor Michael Lassner
Attorney Docket Number MTC 6718

METHOD OF PAYMENT

1. [ ] The Commissioner is hereby authorized to charge the indicated fees to Deposit Account No. 19-1345, in the name of Senniger, Powers, Leavitt & Roedel.

   [ ] The Commissioner is hereby authorized to charge any additional filing and claim fees under 37 CFR 1.16 and application processing fees under 37 CFR 1.17 to Deposit Account No. 19-1345, in the name of Senniger, Powers, Leavitt & Roedel.

2. [X] Check Enclosed. The Commissioner is hereby authorized to charge any under payment or credit any over payment to Deposit Account No. 19-1345, in the name of Senniger, Powers, Leavitt & Roedel.

FEE CALCULATION

1. [X] BASIC FILING FEE $ 690.00 (Type: Formal)
   Entity Status: Large

2. [X] CLAIM FEE $ 4,490.00

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   Independent Claims 30
   Multiple Dependent Claims 18

3. [ ] ADDITIONAL FEES $_____

   [ ] Surcharge - late filing fee or oath
   [ ] Surcharge - late provisional filing fee or cover sheet
   [ ] Extension for reply within _____ month
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   [ ] Request for Reexamination
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   [ ] Submission of Information Disclosure Statement
   [ ] Recording each patent assignment per property
   [ ] Other: _____

TOTAL AMOUNT OF PAYMENT $ 5,180.00

James E. Butler, Ph.D., Reg. No. 40,931 — 8/30/00 Date

JEBmkd

PLANT STEROL ACYLTRANSFERASES

CROSS-REFERENCE TO RELATED APPLICATIONS

This application claims priority to U.S. provisional application Serial No. 60/152,493, filed August 30, 1999 and herein incorporated by reference in its entirety for all purposes.

BACKGROUND

Technical Field

The present invention is directed to plant acyltransferase-like nucleic acid and amino acid sequences and constructs, and methods related to their use in altering sterol composition and/or content, and oil composition and/or content in host cells and plants.

Related Art

Through the development of plant genetic engineering techniques, it is now possible to produce transgenic varieties of plant species to provide plants which have novel and desirable characteristics. For example, it is now possible to genetically engineer plants for tolerance to environmental stresses, such as resistance to pathogens and tolerance to herbicides. It is also possible to improve the nutritional characteristics of the plant, for example to provide improved fatty acid, carotenoid, sterol and tocopherol compositions. However, the number of useful nucleotide sequences for the engineering of such characteristics is thus far limited.

There is a need for improved means to obtain or manipulate compositions of sterols from biosynthetic or natural plant sources. The ability to increase sterol production or alter the sterol compositions in plants may provide for novel sources of sterols for use in human and animal nutrition.

Sterol biosynthesis branches from the farnesyl diphosphate intermediate in the isoprenoid pathway. Sterol biosynthesis occurs via a mevalonate dependent pathway in mammals and higher plants (Goodwin,(1981) *Biosynthesis of Isoprenoid Compounds*, vol 1 (Porter, J.W. & Spurgeon, S.L., eds) pp.443-480, John Wiley and Sons, New York), while in green algae sterol biosynthesis is thought to occur via a mevalonate independent pathway (Schwender, *et al.* (1997) *Physiology, Biochemistry, and Molecular Biology of Plant Lipids*, (Williams, J.P., Khan, M.U., and Lem, N.W., eds) pp. 180-182, Kluwer Academic Publishers, Norwell, MA).

The solubility characteristics of sterol esters suggests that this is the storage form of sterols (Chang, *et al.*, (1997) *Annu. Rev. Biochem.*, 66:613-638). Sterol O-acyltransferase enzymes such as acyl CoA:cholesterol acyltransferase (ACAT) and lecithin:cholesterol acyltransferase (LCAT) catalyze the formation of cholesterol esters, and thus are key to controlling the intracellular cholesterol storage. In yeast, it has been reported that overexpression of *LRO1*, a homolog of human LCAT, and phospholipid:diacylglycerol acyltransferase increased lipid synthesis (Oelkers et al., (2000) *J. Biol. Chem.*, 26:15609-15612; Dahlqvist et al., (2000) *Proc. Natl. Acad. Sci. USA*, 97:6487-6492).

The characterization of various acyltransferase proteins is useful for the further study of plant sterol synthesis systems and for the development of novel and/or alternative sterol sources. Studies of plant mechanisms may provide means to further enhance, control, modify, or otherwise alter the sterol composition of plant cells. Furthermore, such alterations in sterol content and/or composition may provide a means for obtaining tolerance to stress and insect damage. Of particular interest are the nucleic acid sequences of genes encoding proteins which may be useful for applications in genetic engineering.

SUMMARY OF THE INVENTION

The present invention is directed to lecithin:cholesterol acyltransferase-like polypeptides (also referred to herein as LCAT) and acyl CoA:cholesterol acyltransferase-like polypeptides (also referred to herein as ACAT). In particular the invention is related to polynucleotides encoding such sterol:acyltransferases, polypeptides encoded by such polynucleotides, and the use of such polynucleotides to alter sterol composition and oil production. The polynucleotides of the present invention include those derived from plant sources.

One aspect of the invention, therefore, is an isolated nucleic acid sequence encoding a plant lecithin:cholesterol acyltransferase-like polypeptide, a fragment of a plant lecithin:cholesterol acyltransferase-like polypeptide, a plant acyl CoA:cholesterol acyltransferase-like polypeptide or a fragment of a plant acyl CoA:cholesterol acyltransferase-like polypeptide.

Another aspect provides an isolated nucleic acid sequence consisting essentially of SEQ ID NO: 2, 4, 6, 8, 10-29, 43-51, 73 or 75. Also provided is an isolated nucleic acid sequence consisting of SEQ ID NO: 2, 4, 6, 8, 10-29, 43-51, 73 or 75.

Still another aspect provides an isolated nucleic acid sequence comprising a polynucleotide selected from the group consisting of an isolated polynucleotide encoding a polypeptide of SEQ ID NO: 3 or SEQ ID NO: 3 with at least one conservative amino acid substitution; SEQ ID NO: 2; an isolated polynucleotide that has at least 70%, 80%, 90%, or 95% sequence identity with SEQ ID NO: 2; an isolated polynucleotide of at least 10 amino acids that hybridizes under stringent conditions to SEQ ID NO: 2; an isolated polynucleotide complementary to any of the foregoing; and an isolated polynucleotide that hybridizes under stringent conditions to SEQ ID NO: 2 and encodes a plant lecithin:cholesterol acyltransferase-like polypeptide.

Still another aspect provides an isolated nucleic acid sequence consisting essentially of a polynucleotide of the formula 5' X-$(R_1)_n$-$(R_2)_n$-$(R_3)_n$-Y 3' where X is a hydrogen, Y is a hydrogen or a metal, R_1 and R_2 are any nucleic acid, n is an integer between 0-3000, and R_2 is selected from the group consisting of an isolated polynucleotide encoding a polypeptide of SEQ ID NO: 3 or SEQ ID NO: 3 with at least one conservative amino acid substitution; SEQ ID NO: 2; an isolated polynucleotide that has at least 70%, 80%, 90%, or 95% sequence identity with SEQ ID NO: 2; an isolated polynucleotide of at least 10 amino acids that hybridizes under stringent conditions to SEQ ID NO: 2; an isolated polynucleotide complementary to any of the foregoing; and an isolated polynucleotide that hybridizes under stringent conditions to SEQ ID NO: 2 and encodes a plant lecithin:cholesterol acyltransferase-like polypeptide.

Another aspect provides an isolated nucleic acid sequence comprising a polynucleotide selected from the group consisting of an isolated polynucleotide encoding a polypeptide of SEQ ID NO:5 or SEQ ID NO: 5 with at least one conservative amino acid substitution; SEQ ID NO: 4; an isolated polynucleotide that has at least 70%, 80%, 90%, or 95% sequence identity with SEQ ID NO: 4; an isolated polynucleotide of at least 10 amino acids that hybridizes under stringent conditions to SEQ ID NO: 4; an isolated polynucleotide complementary to any of the foregoing; and an isolated polynucleotide that hybridizes under stringent conditions to SEQ ID NO: 4 and encodes a plant lecithin:cholesterol acyltransferase-like polypeptide.

Another aspect provides an isolated nucleic acid sequence consisting essentially of a polynucleotide of the formula 5' X-$(R_1)_n$-$(R_2)_n$-$(R_3)_n$-Y 3' where X is a hydrogen, Y is a hydrogen or a metal, R_1 and R_2 are any nucleic acid, n is an integer between 0-3000, and R_2 is selected from the group consisting of an isolated polynucleotide encoding a polypeptide of

SEQ ID NO: 5 or SEQ ID NO: 5 with at least one conservative amino acid substitution; SEQ ID NO: 4; an isolated polynucleotide that has at least 70%, 80%, 90%, or 95% sequence identity with SEQ ID NO: 4; an isolated polynucleotide of at least 10 amino acids that hybridizes under stringent conditions to SEQ ID NO: 4; an isolated polynucleotide complementary to any of the foregoing; and an isolated polynucleotide that hybridizes under stringent conditions to SEQ ID NO: 4 and encodes a plant lecithin:cholesterol acyltransferase-like polypeptide.

Another aspect provides an isolated nucleic acid sequence comprising a polynucleotide selected from the group consisting of an isolated polynucleotide encoding a polypeptide of SEQ ID NO:7 or SEQ ID NO: 7 with at least one conservative amino acid substitution; SEQ ID NO: 6; an isolated polynucleotide that has at least 70%, 80%, 90%, or 95% sequence identity with SEQ ID NO: 6; an isolated polynucleotide of at least 10 amino acids that hybridizes under stringent conditions to SEQ ID NO: 6; an isolated polynucleotide complementary to any of the foregoing; and an isolated polynucleotide that hybridizes under stringent conditions to SEQ ID NO: 6 and encodes a plant lecithin:cholesterol acyltransferase-like polypeptide.

Another aspect provides an isolated nucleic acid sequence consisting essentially of a polynucleotide of the formula 5' $X-(R_1)_n-(R_2)_n-(R_3)_n-Y$ 3' where X is a hydrogen, Y is a hydrogen or a metal, R_1 and R_2 are any nucleic acid, n is an integer between 0-3000, and R_2 is selected from the group consisting of an isolated polynucleotide encoding a polypeptide of SEQ ID NO: 7 or SEQ ID NO: 7 with at least one conservative amino acid substitution; SEQ ID NO: 6; an isolated polynucleotide that has at least 70%, 80%, 90%, or 95% sequence identity with SEQ ID NO: 6; an isolated polynucleotide of at least 10 amino acids that hybridizes under stringent conditions to SEQ ID NO: 6; an isolated polynucleotide complementary to any of the foregoing; and an isolated polynucleotide that hybridizes under stringent conditions to SEQ ID NO: 6 and encodes a plant lecithin:cholesterol acyltransferase-like polypeptide.

Another aspect provides an isolated nucleic acid sequence comprising a polynucleotide selected from the group consisting of an isolated polynucleotide encoding a polypeptide of SEQ ID NO:9 or SEQ ID NO: 9 with at least one conservative amino acid substitution; SEQ ID NO: 8; an isolated polynucleotide that has at least 70%, 80%, 90%, or 95% sequence identity with SEQ ID NO: 8; an isolated polynucleotide of at least 10 amino acids that hybridizes under stringent conditions to SEQ ID NO: 8; an isolated polynucleotide

complementary to any of the foregoing; and an isolated polynucleotide that hybridizes under stringent conditions to SEQ ID NO: 8 and encodes a plant lecithin:cholesterol acyltransferase-like polypeptide.

Another aspect provides an isolated nucleic acid sequence consisting essentially of a polynucleotide of the formula 5' X-$(R_1)_n$-$(R_2)_n$-$(R_3)_n$-Y 3' where X is a hydrogen, Y is a hydrogen or a metal, R_1 and R_2 are any nucleic acid, n is an integer between 0-3000, and R_2 is selected from the group consisting of an isolated polynucleotide encoding a polypeptide of SEQ ID NO: 9 or SEQ ID NO: 9 with at least one conservative amino acid substitution; SEQ ID NO: 8; an isolated polynucleotide that has at least 70%, 80%, 90%, or 95% sequence identity with SEQ ID NO: 8; an isolated polynucleotide of at least 10 amino acids that hybridizes under stringent conditions to SEQ ID NO: 8; an isolated polynucleotide complementary to any of the foregoing; and an isolated polynucleotide that hybridizes under stringent conditions to SEQ ID NO: 8 and encodes a plant lecithin:cholesterol acyltransferase-like polypeptide.

Another aspect provides an isolated nucleic acid sequence comprising a polynucleotide selected from the group consisting of an isolated polynucleotide encoding a polypeptide of SEQ ID NO:74 or SEQ ID NO: 74 with at least one conservative amino acid substitution; SEQ ID NO: 73; an isolated polynucleotide that has at least 70%, 80%, 90%, or 95% sequence identity with SEQ ID NO: 73; an isolated polynucleotide of at least 10 amino acids that hybridizes under stringent conditions to SEQ ID NO: 73; an isolated polynucleotide complementary to any of the foregoing; and an isolated polynucleotide that hybridizes under stringent conditions to SEQ ID NO: 73 and encodes a plant lecithin:cholesterol acyltransferase-like polypeptide.

Another aspect provides an isolated nucleic acid sequence consisting essentially of a polynucleotide of the formula 5' X-$(R_1)_n$-$(R_2)_n$-$(R_3)_n$-Y 3' where X is a hydrogen, Y is a hydrogen or a metal, R_1 and R_2 are any nucleic acid, n is an integer between 0-3000, and R_2 is selected from the group consisting of an isolated polynucleotide encoding a polypeptide of SEQ ID NO: 74 or SEQ ID NO: 74 with at least one conservative amino acid substitution; SEQ ID NO: 73; an isolated polynucleotide that has at least 70%, 80%, 90%, or 95% sequence identity with SEQ ID NO: 73; an isolated polynucleotide of at least 10 amino acids that hybridizes under stringent conditions to SEQ ID NO: 73; an isolated polynucleotide complementary to any of the foregoing; and an isolated polynucleotide that hybridizes under

stringent conditions to SEQ ID NO: 73 and encodes a plant lecithin:cholesterol acyltransferase-like polypeptide.

Another aspect provides an isolated nucleic acid sequence comprising a polynucleotide selected from the group consisting of an isolated polynucleotide encoding a polypeptide of SEQ ID NO:76 or SEQ ID NO: 76 with at least one conservative amino acid substitution; SEQ ID NO: 75; an isolated polynucleotide that has at least 70%, 80%, 90%, or 95% sequence identity with SEQ ID NO: 75; an isolated polynucleotide of at least 10 amino acids that hybridizes under stringent conditions to SEQ ID NO: 75; an isolated polynucleotide complementary to any of the foregoing; and an isolated polynucleotide that hybridizes under stringent conditions to SEQ ID NO: 75 and encodes a plant lecithin:cholesterol acyltransferase-like polypeptide.

Another aspect provides an isolated nucleic acid sequence consisting essentially of a polynucleotide of the formula 5' X-$(R_1)_n$-$(R_2)_n$-$(R_3)_n$-Y 3' where X is a hydrogen, Y is a hydrogen or a metal, R_1 and R_2 are any nucleic acid, n is an integer between 0-3000, and R_2 is selected from the group consisting of an isolated polynucleotide encoding a polypeptide of SEQ ID NO: 76 or SEQ ID NO: 76 with at least one conservative amino acid substitution; SEQ ID NO: 75; an isolated polynucleotide that has at least 70%, 80%, 90%, or 95% sequence identity with SEQ ID NO: 75; an isolated polynucleotide of at least 10 amino acids that hybridizes under stringent conditions to SEQ ID NO: 75; an isolated polynucleotide complementary to any of the foregoing; and an isolated polynucleotide that hybridizes under stringent conditions to SEQ ID NO: 75 and encodes a plant lecithin:cholesterol acyltransferase-like polypeptide.

Another aspect provides an isolated nucleic acid sequence comprising a polynucleotide selected from the group consisting of SEQ ID NO: 42 or a degenerate variant thereof; an isolated polynucleotide that has at least 70%, 80%, 90%, or 95% sequence identity with SEQ ID NO: 42; an isolated polynucleotide of at least 10 amino acids that hybridizes under stringent conditions to SEQ ID NO: 42; an isolated polynucleotide complementary to any of the foregoing; and an isolated polynucleotide that hybridizes under stringent conditions to SEQ ID NO: 42 and encodes an acyl CoA:cholesterol acyltransferase-like polypeptide.

Another aspect provides an isolated nucleic acid sequence consisting essentially of a polynucleotide of the formula 5' X-$(R_1)_n$-$(R_2)_n$-$(R_3)_n$-Y 3' where X is a hydrogen, Y is a hydrogen or a metal, R_1 and R_2 are any nucleic acid, n is an integer between 0-3000, and R_2 is

selected from the group consisting of SEQ ID NO: 42 or a degenerate variant thereof; an isolated polynucleotide that has at least 70%, 80%, 90%, or 95% sequence identity with SEQ ID NO: 42; an isolated polynucleotide of at least 10 amino acids that hybridizes under stringent conditions to SEQ ID NO: 42; an isolated polynucleotide complementary to any of the foregoing; and an isolated polynucleotide that hybridizes under stringent conditions to SEQ ID NO: 42 and encodes a acyl CoA:cholesterol acyltransferase-like polypeptide.

Also provided is a recombinant nucleic acid construct comprising a regulatory sequence operably linked to a polynucleotide encoding a lecithin:cholesterol acyltransferase-like polypeptide and/or an acyl CoA:cholesterol acyltransferase-like polypeptide. In one embodiment, the sterol acyl transferases are plant sterol acyl transferases. In another embodiment, the recombinant nucleic acid constructs further comprises a termination sequence. The regulatory sequence can be a constitutive promoter, an inducible promoter, a developmentally regulated promoter, a tissue specific promoter, an organelle specific promoter, a seed specific promoter or a combination of any of the foregoing. Also provided is a plant containing this recombinant nucleic acid construct and the seed and progeny from such a plant. This recombinant nucleic acid construct can also be introduced into a suitable host cell to provide yet another aspect of the invention. If the host cell is a plant host cell, the cell can be used to generate a plant to provide another aspect of the invention. Further aspects include seed and progeny from such a plant.

Yet another aspect is a purified polypeptide comprising, SEQ ID NO: 3, SEQ ID NO: 5, SEQ ID NO: 7, SEQ ID NO: 9, SEQ ID NO: 74, SEQ ID NO: 76, or any of the preceding sequences with at least one conservative amino acid substitution.

Still another aspect provides a purified immunogenic polypeptide comprising at least 10 consecutive amino acids from an amino acid sequence selected from the group consisting of SEQ ID NO: 3, 5, 7, 9, 74, 76 and any of the preceding sequences containing at least one conservative amino acid substitution. Also provided are antibodies, either polyclonal or monoclonal, that specifically bind the preceding immunogenic polypeptides.

One aspect provides a method for producing a lecithin:cholesterol acyltransferase-like polypeptide or an acyl CoA:cholesterol acyltransferase-like polypeptide comprising culturing a host cell containing any recombinant nucleic acid construct of the present invention under condition permitting expression of said lecithin:cholesterol acyltransferase-like polypeptide or acyl CoA:cholesterol acyltransferase-like polypeptide.

Another aspect provides a method for modifying the sterol content of a host cell, comprising transforming a host cell with a recombinant construct containing a regulatory sequence operably linked to a polynucleotide encoding a lecithin:cholesterol acyltransferase-like polypeptide and culturing said host cell under conditions wherein said host cell expresses a lecithin:cholesterol acyltransferase-like polypeptide such that said host cell has a modified sterol composition as compared to host cells without the recombinant construct.

An additional aspect is a method for modifying the sterol content of a host cell comprising transforming a host cell with a recombinant construct containing a regulatory sequence operably linked to a polynucleotide encoding an acyl CoA:cholesterol acyltransferase-like polypeptide and culturing said host cell under conditions wherein said host cell expresses an acyl CoA:cholesterol acyltransferase-like polypeptide such that said host cell has a modified sterol composition as compared to host cells without the recombinant construct.

A further aspect is a plant comprising a recombinant construct containing a regulatory sequence operably linked to a polynucleotide encoding a lecithin:cholesterol acyltransferase-like polypeptide wherein expression of said recombinant construct results in modified sterol composition of said plant as compared to the same plant without said recombinant construct.

Another aspect provides a plant comprising a recombinant construct containing a regulatory sequence operably linked to a polynucleotide encoding an acyl CoA:cholesterol acyltransferase-like polypeptide wherein expression of said recombinant construct results in modified sterol composition of said plant as compared to the same plant without said recombinant construct.

In a further aspect is provided an oil obtained from any of the plants or host cells of the present invention.

In still another aspect is provided a method for producing an oil with a modified sterol composition comprising providing any of the plants or host cells of the present invention and extracting oil from the plant by any known method. Also provided is an oil produced by the preceding method.

Still another aspect provides a method for altering oil production by a host cell comprising, transforming a host cell with a recombinant construct containing a regulatory sequence operably linked to a polynucleotide encoding a lecithin:cholesterol acyltransferase-like polypeptide and culturing the host cell under conditions wherein the host cell expresses a

lecithin:cholesterol acyltransferase-like polypeptide such that the host cell has an altered oil production as compared to host cells without the recombinant construct.

Another aspect provides a method for altering oil production by a host cell comprising, transforming a host cell with a recombinant construct containing a regulatory sequence operably linked to a polynucleotide encoding an acyl CoA:cholesterol acyltransferase-like polypeptide and culturing the host cell under conditions wherein the host cell expresses an acyl CoA:cholesterol acyltransferase-like polypeptide such that the host cell has an altered oil production as compared to host cells without the recombinant construct.

Also provided is a plant comprising a recombinant construct containing a regulatory sequence operably linked to a polynucleotide encoding a lecithin:cholesterol acyltransferase-like polypeptide wherein expression of said recombinant construct results in an altered production of oil by said plant as compared to the same plant without said recombinant construct.

In a further aspect is provided a plant comprising a recombinant construct containing a regulatory sequence operably linked to a polynucleotide encoding an acyl CoA:cholesterol acyltransferase-like polypeptide wherein expression of said recombinant construct results in an altered production of oil by said plant as compared to the same plant without said recombinant construct.

Additional aspects provide a food, food ingredient or food product comprising any oil, plant or host cell of the present invention; a nutritional or dietary supplement comprising any oil, plant or host cell of the present invention; and a pharmaceutical composition comprising any oil, plant or host cell of the present invention along with a suitable diluent, carrier or excipient.

Additional aspects will be apparent from the descriptions and examples that follow.

BRIEF DESCRIPTION OF THE DRAWINGS

These and other features, aspects, and advantages of the present invention will become better understood with regard to the following description, appended claims and accompanying figures where:

Figure 1 shows an alignment of yeast, human and rat lecithin:cholesterol acyltransferase protein sequences with *Arabidopsis* LCAT1, LCAT2, LCAT3, and LCAT4 deduced amino acid sequences.

Figure 2 shows the results of NMR sterol ester analysis on T2 seed from plant expressing LCAT4 under the control of the napin promoter (pCGN9998).

Figure 3 shows the results of HPLC/MS sterol analysis on oil extracted from T2 seed from control lines (pCGN8640) and lines expressing LCAT3 (pCGN9968) under the control of the napin promoter.

Figure 4 shows the results of HPLC/MS sterol analysis on oil extracted from T2 seed from control lines (pCGN8640), and plant line expressing LCAT1 (pCGN9962), LCAT2 (pCGN9983), LCAT3 (pCGN9968), and LCAT4 (pCGN9998) under the control of the napin promoter. Additionally, data from 3 lines expressing LCAT4 under the control of the 35S promoter (pCGN9996) are shown.

Figure 5 shows the results of Nir analysis of the oil content of T2 seed from control lines (pCGN8640), and plant lines expressing LCAT1 (pCGN9962), LCAT2 (pCGN9983), and LCAT3 (pCGN9968) under the control of the napin promoter. Additionally, data from 16 lines expressing LCAT2 under the control of the 35S promoter (pCGN9981) are shown.

DETAILED DESCRIPTION

The following detailed description is provided to aid those skilled in the art in practicing the present invention. Even so, this detailed description should not be construed to unduly limit the present invention as modifications and variations in the embodiments discussed herein can be made by those of ordinary skill in the art without departing from the spirit or scope of the present inventive discovery.

All publications, patents, patent applications and other references cited in this application are herein incorporated by reference in their entirety as if each individual publication, patent, patent application or other reference were specifically and individually indicated to be incorporated by reference.

The present invention relates to lecithin:cholesterol acyltransferase, particularly the isolated nucleic acid sequences encoding lecithin:cholesterol-like polypeptides (LCAT) from plant sources and acyl CoA:cholesterol:acyltransferase, particularly the isolated nucleic acid sequences encoding acyl CoA:cholesterol acyltransferase-like polypeptides (ACAT) from plant sources. Lecithin:cholesterol acyltransferase-like as used herein includes any nucleic acid sequence encoding an amino acid sequence from a plant source, such as a protein, polypeptide or peptide, obtainable from a cell source, which demonstrates the ability to utilize

lecithin (phosphatidyl choline) as an acyl donor for acylation of sterols or glycerides to form esters under enzyme reactive conditions along with such proteins polypeptides and peptides. Acyl CoA:cholesterol acyltransferase-like as used herein includes any nucleic acid sequence encoding an amino acid sequence from a plant source, such as a protein, polypeptide or peptide, obtainable from a cell source, which demonstrates the ability to utilize acyl CoA as an acyl donor for acylation of sterols or glycerides to form esters under enzyme reactive conditions along with such proteins polypeptides and peptides. By "enzyme reactive conditions" is meant that any necessary conditions are available in an environment (i.e., such factors as temperature, pH, lack of inhibiting substances) which will permit the enzyme to function.

The term "sterol" as applied to plants refers to any chiral tetracyclic isopentenoid which may be formed by cyclization of squalene oxide through the transition state possessing stereochemistry similar to the *trans-syn-trans-anti-trans-anti* configuration, for example, protosteroid cation, and which retains a polar group at C-3 (hydroxyl or keto), an *all-trans-anti* stereochemistry in the ring system, and a side-chain 20R-configuration (Parker, *et al.* (1992) *In* Nes, *et al.*, Eds., *Regulation of Isopentenoid Metabolism*, ACS Symposium Series No. 497, p. 110; American Chemical Society, Washington, D.C.).

Sterols may or may not contain a C-5-C-6 double bond, as this is a feature introduced late in the biosynthetic pathway. Sterols contain a C_8-C_{10} side chain at the C-17 position.

The term "phytosterol," which applies to sterols found uniquely in plants, refers to a sterol containing a C-5, and in some cases a C-22, double bond. Phytosterols are further characterized by alkylation of the C-17 side-chain with a methyl or ethyl substituent at the C-24 position. Major phytosterols include, but are not limited to, sitosterol, stigmasterol, campesterol, brassicasterol, etc. Cholesterol, which lacks a C-24 methyl or ethyl side-chain, is found in plants, but is not unique thereto, and is not a "phytosterol."

"Phytostanols" are saturated forms of phytosterols wherein the C-5 and, when present, C-22 double bond(s) is (are) reduced, and include, but are not limited to, sitostanol, campestanol, and 22-dihydrobrassicastanol.

"Sterol esters" are further characterized by the presence of a fatty acid or phenolic acid moiety rather than a hydroxyl group at the C-3 position.

The term "sterol" includes sterols, phytosterols, phytosterol esters, phytostanols, and phytostanol esters.

The term "sterol compounds" includes sterols, phyotsterols, phytosterol esters, phytostanols, and phytostanol esters.

The term "phytosterol compound" refers to at least one phytosterol, at least one phytosterol ester, or a mixture thereof.

The term "phytostanol compound" refers to at least one phytostanol, at least one phytostanol ester, or a mixture thereof.

The term "glyceride" refers to a fatty acid ester of glycerol and includes mono-, di-, and tri- acylglycerols.

As used herein, "recombinant construct" is defined either by its method of production or its structure. In reference to its method of production, e.g., a product made by a process, the process is use of recombinant nucleic acid techniques, e.g., involving human intervention in the nucleotide sequence, typically selection or production. Alternatively, in terms of structure, it can be a sequence comprising fusion of two or more nucleic acid sequences which are not naturally contiguous or operatively linked to each other

As used herein, "regulatory sequence" means a sequence of DNA concerned with controlling expression of a gene; e.g. promoters, operators and attenuators. A "heterologous regulatory sequence" is one which differs from the regulatory sequence naturally associated with a gene.

As used herein, "polynucleotide" and "oligonucleotide" are used interchangeably and mean a polymer of at least two nucleotides joined together by a phosphodiester bond and may consist of either ribonucleotides or deoxynucleotides.

As used herein, "sequence" means the linear order in which monomers appear in a polymer, for example, the order of amino acids in a polypeptide or the order of nucleotides in a polynucleotide.

As used herein, "polypeptide", "peptide", and "protein" are used interchangeably and mean a compound that consist of two or more amino acids that are linked by means of peptide bonds.

As used herein, the terms "complementary" or "complementarity" refer to the pairing of bases, purines and pyrimidines, that associate through hydrogen bonding in double stranded nucleic acids. For example, the following base pairs are complementary: guanine and cytosine; adenine and thymine; and adenine and uracil. The terms, as used herein, include complete and partial complementarity.

Isolated proteins, Polypeptides and Polynucleotides

A first aspect of the present invention relates to isolated LCAT polynucleotides. The polynucleotide sequences of the present invention include isolated polynucleotides that encode the polypeptides of the invention having a deduced amino acid sequence selected from the group of sequences set forth in the Sequence Listing and to other polynucleotide sequences closely related to such sequences and variants thereof.

The invention provides a polynucleotide sequence identical over its entire length to each coding sequence as set forth in the Sequence Listing. The invention also provides the coding sequence for the mature polypeptide or a fragment thereof, as well as the coding sequence for the mature polypeptide or a fragment thereof in a reading frame with other coding sequences, such as those encoding a leader or secretory sequence, a pre-, pro-, or prepro- protein sequence. The polynucleotide can also include non-coding sequences, including for example, but not limited to, non-coding 5' and 3' sequences, such as the transcribed, untranslated sequences, termination signals, ribosome binding sites, sequences that stabilize mRNA, introns, polyadenylation signals, and additional coding sequence that encodes additional amino acids. For example, a marker sequence can be included to facilitate the purification of the fused polypeptide. Polynucleotides of the present invention also include polynucleotides comprising a structural gene and the naturally associated sequences that control gene expression.

The invention also includes polynucleotides of the formula:

$$X\text{-}(R_1)_n\text{-}(R_2)\text{-}(R_3)_n\text{-}Y$$

wherein, at the 5' end, X is hydrogen, and at the 3' end, Y is hydrogen or a metal, R_1 and R_3 are any nucleic acid residue, n is an integer between 0 and 3000, preferably between 1 and 1000 and R_2 is a nucleic acid sequence of the invention, particularly a nucleic acid sequence selected from the group set forth in the Sequence Listing and preferably SEQ ID NOs: 2, 4, 6, 8, 10-29, 33, 42-51, 73 and 75. In the formula, R_2 is oriented so that its 5' end residue is at the left, bound to R_1, and its 3' end residue is at the right, bound to R_3. Any stretch of nucleic acid residues denoted by either R group, where R is greater than 1, may be either a heteropolymer or a homopolymer, preferably a heteropolymer.

The invention also relates to variants of the polynucleotides described herein that encode for variants of the polypeptides of the invention. Variants that are fragments of the

polynucleotides of the invention can be used to synthesize full-length polynucleotides of the invention. Preferred embodiments are polynucleotides encoding polypeptide variants wherein 5 to 10, 1 to 5, 1 to 3, 2, 1 or no amino acid residues of a polypeptide sequence of the invention are substituted, added or deleted, in any combination. Particularly preferred are substitutions, additions, and deletions that are silent such that they do not alter the properties or activities of the polynucleotide or polypeptide.

Further preferred embodiments of the invention that are at least 50%, 60%, or 70% identical over their entire length to a polynucleotide encoding a polypeptide of the invention, and polynucleotides that are complementary to such polynucleotides. More preferable are polynucleotides that comprise a region that is at least 80% identical over its entire length to a polynucleotide encoding a polypeptide of the invention and polynucleotides that are complementary thereto. In this regard, polynucleotides at least 90% identical over their entire length are particularly preferred, those at least 95% identical are especially preferred. Further, those with at least 97% identity are highly preferred and those with at least 98% and 99% identity are particularly highly preferred, with those at least 99% being the most highly preferred.

Preferred embodiments are polynucleotides that encode polypeptides that retain substantially the same biological function or activity as determined by the methods described herein as the mature polypeptides encoded by the polynucleotides set forth in the Sequence Listing.

The invention further relates to polynucleotides that hybridize to the above-described sequences. In particular, the invention relates to polynucleotides that hybridize under stringent conditions to the above-described polynucleotides. An example of stringent hybridization conditions is overnight incubation at 42°C in a solution comprising 50% formamide, 5x SSC (150 mM NaCl, 15 mM trisodium citrate), 50 mM sodium phosphate (pH 7.6), 5x Denhardt's solution, 10% dextran sulfate, and 20 micrograms/milliliter denatured, sheared salmon sperm DNA, followed by washing the hybridization support in 0.1x SSC at approximately 65°C. Also included are polynucleotides that hybridize under a wash stringency of 0.1X SSC or 0.1X SSPE (at 50°C. Other hybridization and wash conditions are well known and are exemplified in Sambrook, *et al.*, Molecular Cloning: A Laboratory Manual, Second Edition, Cold Spring Harbor, NY (1989), particularly Chapter 11.

The invention also provides a polynucleotide consisting essentially of a polynucleotide sequence obtainable by screening an appropriate library containing the

complete gene for a polynucleotide sequence set for in the Sequence Listing under stringent hybridization conditions with a probe having the sequence of said polynucleotide sequence or a fragment thereof; and isolating said polynucleotide sequence. Methods for screening libraries are well known in the art and can be found for example in Sambrook, *et al.*, *Molecular Cloning: A Laboratory Manual*, Second Edition, Cold Spring Harbor, NY (1989), particularly Chapter 8 and Ausubel et al., *Short Protocols in Molecular Biology*, 3rd ed, Wiley and Sons, 1995, chapter 6. Nucleic acid sequences useful for obtaining such a polynucleotide include, for example, probes and primers as described herein and in particular SEQ ID NO: 2, 4, 6, 8, 10-29, 33, 42-51, 73 and 75. These sequences are particularly useful in screening libraries obtained from *Arabidopsis*, soybean and corn for sequences encoding lecithin:cholesterol acyltransferase and lecithin:cholesterol acyltransferase-like polypeptides and for screening libraries for sequences encoding acyl CoA:cholesterol acyl transferase and acyl CoA:cholesterol acyl transferase-like polypeptides.

As discussed herein regarding polynucleotide assays of the invention, for example, polynucleotides of the invention can be used as a hybridization probe for RNA, cDNA, or genomic DNA to isolate full length cDNAs or genomic clones encoding a polypeptide and to isolate cDNA or genomic clones of other genes that have a high sequence similarity to a polynucleotide set forth in the Sequence Listing and in particular SEQ ID NO: 2, 4, 6, 8, 10-29, 33, 42-51, 73 and 75. Such probes will generally comprise at least 15 bases. Preferably such probes will have at least 30 bases and can have at least 50 bases. Particularly preferred probes will have between 30 bases and 50 bases, inclusive.

The coding region of each gene that comprises or is comprised by a polynucleotide sequence set forth in the Sequence Listing may be isolated by screening using a DNA sequence provided in the Sequence Listing to synthesize an oligonucleotide probe. A labeled oligonucleotide having a sequence complementary to that of a gene of the invention is then used to screen a library of cDNA, genomic DNA or mRNA to identify members of the library which hybridize to the probe. For example, synthetic oligonucleotides are prepared which correspond to the LCAT EST sequences. The oligonucleotides are used as primers in polymerase chain reaction (PCR) techniques to obtain 5' and 3' terminal sequence of LCAT genes. Alternatively, where oligonucleotides of low degeneracy can be prepared from particular LCAT peptides, such probes may be used directly to screen gene libraries for LCAT gene sequences. In particular, screening of cDNA libraries in phage vectors is useful in such methods due to lower levels of background hybridization.

Typically, a LCAT sequence obtainable from the use of nucleic acid probes will show 60-70% sequence identity between the target LCAT sequence and the encoding sequence used as a probe. However, lengthy sequences with as little as 50-60% sequence identity may also be obtained. The nucleic acid probes may be a lengthy fragment of the nucleic acid sequence, or may also be a shorter, oligonucleotide probe. When longer nucleic acid fragments are employed as probes (greater than about 100 bp), one may screen at lower stringencies in order to obtain sequences from the target sample which have 20-50% deviation (i.e., 50-80% sequence homology) from the sequences used as probe. Oligonucleotide probes can be considerably shorter than the entire nucleic acid sequence encoding an LCAT enzyme, but should be at least about 10, preferably at least about 15, and more preferably at least about 20 nucleotides. A higher degree of sequence identity is desired when shorter regions are used as opposed to longer regions. It may thus be desirable to identify regions of highly conserved amino acid sequence to design oligonucleotide probes for detecting and recovering other related LCAT genes. Shorter probes are often particularly useful for polymerase chain reactions (PCR), especially when highly conserved sequences can be identified. (*See*, Gould, *et al.*, *PNAS USA* (1989) *86*:1934-1938.).

Another aspect of the present invention relates to LCAT polypeptides. Such polypeptides include isolated polypeptides set forth in the Sequence Listing, as well as polypeptides and fragments thereof, particularly those polypeptides which exhibit LCAT activity and also those polypeptides which have at least 50%, 60% or 70% identity, preferably at least 80% identity, more preferably at least 90% identity, and most preferably at least 95% identity to a polypeptide sequence selected from the group of sequences set forth in the Sequence Listing, and also include portions of such polypeptides, wherein such portion of the polypeptide preferably includes at least 30 amino acids and more preferably includes at least 50 amino acids.

"Identity", as is well understood in the art, is a relationship between two or more polypeptide sequences or two or more polynucleotide sequences, as determined by comparing the sequences. In the art, "identity" also means the degree of sequence relatedness between polypeptide or polynucleotide sequences, as determined by the match between strings of such sequences. "Identity" can be readily calculated by known methods including, but not limited to, those described in *Computational Molecular Biology*, Lesk, A.M., ed., Oxford University Press, New York (1988); *Biocomputing: Informatics and Genome Projects*, Smith, D.W., ed., Academic Press, New York, 1993; *Computer Analysis of Sequence Data, Part I*, Griffin,

A.M. and Griffin, H.G., eds., Humana Press, New Jersey (1994); *Sequence Analysis in Molecular Biology*, von Heinje, G., Academic Press (1987); *Sequence Analysis Primer*, Gribskov, M. and Devereux, J., eds., Stockton Press, New York (1991); and Carillo, H., and Lipman, D., SIAM *J Applied Math*, 48:1073 (1988). Methods to determine identity are designed to give the largest match between the sequences tested. Moreover, methods to determine identity are codified in publicly available programs. Computer programs which can be used to determine identity between two sequences include, but are not limited to, GCG (Devereux, J., et al., *Nucleic Acids Research* 12(1):387 (1984); suite of five BLAST programs, three designed for nucleotide sequences queries (BLASTN, BLASTX, and TBLASTX) and two designed for protein sequence queries (BLASTP and TBLASTN) (Coulson, *Trends in Biotechnology, 12:* 76-80 (1994); Birren, *et al., Genome Analysis, 1:* 543-559 (1997)). The BLAST X program is publicly available from NCBI and other sources (*BLAST Manual*, Altschul, S., *et al.*, NCBI NLM NIH, Bethesda, MD 20894; Altschul, S., *et al., J. Mol. Biol.*, 215:403-410 (1990)). The well known Smith Waterman algorithm can also be used to determine identity.

Parameters for polypeptide sequence comparison typically include the following:

Algorithm: Needleman and Wunsch, *J. Mol. Biol.* 48:443-453 (1970)

Comparison matrix: BLOSSUM62 from Hentikoff and Hentikoff, *Proc. Natl. Acad. Sci USA* 89:10915-10919 (1992)

Gap Penalty: 12

Gap Length Penalty: 4

A program which can be used with these parameters is publicly available as the "gap" program from Genetics Computer Group, Madison Wisconsin. The above parameters along with no penalty for end gap are the default parameters for peptide comparisons.

Parameters for polynucleotide sequence comparison include the following:

Algorithm: Needleman and Wunsch, J. Mol. Biol. 48:443-453 (1970)

Comparison matrix: matches = +10; mismatches = 0

Gap Penalty: 50

Gap Length Penalty: 3

A program which can be used with these parameters is publicly available as the "gap" program from Genetics Computer Group, Madison Wisconsin. The above parameters are the default parameters for nucleic acid comparisons.

The invention also includes polypeptides of the formula:

$$X\text{-}(R_1)_n\text{-}(R_2)\text{-}(R_3)_n\text{-}Y$$

wherein, at the amino terminus, X is hydrogen, and at the carboxyl terminus, Y is hydrogen or a metal, R_1 and R_3 are any amino acid residue, n is an integer between 0 and 1000, and R_2 is an amino acid sequence of the invention, particularly an amino acid sequence selected from the group set forth in the Sequence Listing and preferably SEQ ID NOs: 3, 5, 7, 9, 74 and 76. In the formula, R_2 is oriented so that its amino terminal residue is at the left, bound to R_1, and its carboxy terminal residue is at the right, bound to R_3. Any stretch of amino acid residues denoted by either R group, where R is greater than 1, may be either a heteropolymer or a homopolymer, preferably a heteropolymer.

Polypeptides of the present invention include isolated polypeptides encoded by a polynucleotide comprising a sequence selected from the group of a sequence contained in SEQ ID NOs: 2, 4, 6, 8, 73 and 75.

The polypeptides of the present invention can be mature protein or can be part of a fusion protein.

Fragments and variants of the polypeptides are also considered to be a part of the invention. A fragment is a variant polypeptide which has an amino acid sequence that is entirely the same as part but not all of the amino acid sequence of the previously described polypeptides. The fragments can be "free-standing" or comprised within a larger polypeptide of which the fragment forms a part or a region, most preferably as a single continuous region. Preferred fragments are biologically active fragments which are those fragments that mediate activities of the polypeptides of the invention, including those with similar activity or improved activity or with a decreased activity. Also included are those polypeptides and polypeptide fragments that are antigenic or immunogenic in an animal, particularly a human and antibodies, either polyclonal or monoclonal that specifically bind the antigenic fragments. In one preferred embodiment, such antigenic or immunogenic fragments comprise at least 10 consecutive amino acids from the amino acid sequences disclosed herein or such sequences with at least one conservative amino acid substitution. In additional embodiments, such antigenic or immunogenic fragments comprise at least 15, at least 25, at least 50 or at least 100 consecutive amino acids from the amino acid sequences disclosed herein or such sequences with at least one conservative amino acid substitution. Methods for the production of antibodies from polypeptides and polypeptides conjugated to carrier molecules are well known in the art and can be found for example in Ausubel et al., *Short Protocols in Molecular Biology*, 3rd ed., Wiley & Sons, 1995, particularly chapter 11.

Variants of the polypeptide also include polypeptides that vary from the sequences set forth in the Sequence Listing by conservative amino acid substitutions, substitution of a residue by another with like characteristics. Those of ordinary skill in the art are aware that modifications in the amino acid sequence of a peptide, polypeptide, or protein can result in equivalent, or possibly improved, second generation peptides, etc., that display equivalent or superior functional characteristics when compared to the original amino acid sequence. The present invention accordingly encompasses such modified amino acid sequences. Alterations can include amino acid insertions, deletions, substitutions, truncations, fusions, shuffling of subunit sequences, and the like, provided that the peptide sequences produced by such modifications have substantially the same functional properties as the naturally occurring counterpart sequences disclosed herein.

One factor that can be considered in making such changes is the hydropathic index of amino acids. The importance of the hydropathic amino acid index in conferring interactive biological function on a protein has been discussed by Kyte and Doolittle (*J. Mol. Biol.*, 157: 105-132, 1982). It is accepted that the relative hydropathic character of amino acids contributes to the secondary structure of the resultant protein. This, in turn, affects the interaction of the protein with molecules such as enzymes, substrates, receptors, DNA, antibodies, antigens, etc.

Based on its hydrophobicity and charge characteristics, each amino acid has been assigned a hydropathic index as follows: isoleucine (+4.5); valine (+4.2); leucine (+3.8); phenylalanine (+2.8); cysteine/cystine (+2.5); methionine (+1.9); alanine (+1.8); glycine (-0.4); threonine (-0.7); serine (-0.8); tryptophan (-0.9); tyrosine (-1.3); proline (-1.6); histidine (-3.2); glutamate/glutamine/aspartate/asparagine (-3.5); lysine (-3.9); and arginine (-4.5).

As is known in the art, certain amino acids in a peptide or protein can be substituted for other amino acids having a similar hydropathic index or score and produce a resultant peptide or protein having similar biological activity, i.e., which still retains biological functionality. In making such changes, it is preferable that amino acids having hydropathic indices within ±2 are substituted for one another. More preferred substitutions are those wherein the amino acids have hydropathic indices within ±1. Most preferred substitutions are those wherein the amino acids have hydropathic indices within ±0.5.

Like amino acids can also be substituted on the basis of hydrophilicity. U.S. Patent No. 4,554,101 discloses that the greatest local average hydrophilicity of a protein, as governed by the hydrophilicity of its adjacent amino acids, correlates with a biological

property of the protein. The following hydrophilicity values have been assigned to amino acids: arginine/lysine (+3.0); aspartate/glutamate (+3.0 ±1); serine (+0.3); asparagine/glutamine (+0.2); glycine (0); threonine (-0.4); proline (-0.5 ±1); alanine/histidine (-0.5); cysteine (-1.0); methionine (-1.3); valine (-1.5); leucine/isoleucine (-1.8); tyrosine (-2.3); phenylalanine (-2.5); and tryptophan (-3.4). Thus, one amino acid in a peptide, polypeptide, or protein can be substituted by another amino acid having a similar hydrophilicity score and still produce a resultant protein having similar biological activity, i.e., still retaining correct biological function. In making such changes, amino acids having hydropathic indices within ±2 are preferably substituted for one another, those within ±1 are more preferred, and those within ±0.5 are most preferred.

As outlined above, amino acid substitutions in the peptides of the present invention can be based on the relative similarity of the amino acid side-chain substituents, for example, their hydrophobicity, hydrophilicity, charge, size, etc. Exemplary substitutions that take various of the foregoing characteristics into consideration in order to produce conservative amino acid changes resulting in silent changes within the present peptides, etc., can be selected from other members of the class to which the naturally occurring amino acid belongs. Amino acids can be divided into the following four groups: (1) acidic amino acids; (2) basic amino acids; (3) neutral polar amino acids; and (4) neutral non-polar amino acids. Representative amino acids within these various groups include, but are not limited to: (1) acidic (negatively charged) amino acids such as aspartic acid and glutamic acid; (2) basic (positively charged) amino acids such as arginine, histidine, and lysine; (3) neutral polar amino acids such as glycine, serine, threonine, cysteine, cystine, tyrosine, asparagine, and glutamine; and (4) neutral non-polar amino acids such as alanine, leucine, isoleucine, valine, proline, phenylalanine, tryptophan, and methionine. It should be noted that changes which are not expected to be advantageous can also be useful if these result in the production of functional sequences.

Variants that are fragments of the polypeptides of the invention can be used to produce the corresponding full length polypeptide by peptide synthesis. Therefore, these variants can be used as intermediates for producing the full-length polypeptides of the invention.

The polynucleotides and polypeptides of the invention can be used, for example, in the transformation of host cells, such as plant cells, animal cells, yeast cells, bacteria, bacteriophage, and viruses, as further discussed herein.

The invention also provides polynucleotides that encode a polypeptide that is a mature protein plus additional amino or carboxyl-terminal amino acids, or amino acids within the mature polypeptide (for example, when the mature form of the protein has more than one polypeptide chain). Such sequences can, for example, play a role in the processing of a protein from a precursor to a mature form, allow protein transport, shorten or lengthen protein half-life, or facilitate manipulation of the protein in assays or production. It is contemplated that cellular enzymes can be used to remove any additional amino acids from the mature protein.

A precursor protein, having the mature form of the polypeptide fused to one or more prosequences may be an inactive form of the polypeptide. The inactive precursors generally are activated when the prosequences are removed. Some or all of the prosequences may be removed prior to activation. Such precursor protein are generally called proproteins.

Preparation of Expression Constructs and Methods of Use

Of interest is the use of the nucleotide sequences in recombinant DNA constructs to direct the transcription or transcription and translation (expression) of the acyltransferase sequences of the present invention in a host cell. Of particular interest is the use of the polynucleotide sequences of the present invention in recombinant DNA constructs to direct the transcription or transcription and translation (expression) of the acyltransferase sequences of the present invention in a host plant cell.

The expression constructs generally comprise a regulatory sequence functional in a host cell operably linked to a nucleic acid sequence encoding a lecithin:cholesterol acyltransferase-like polypeptide or acyl CoA:cholesterol acyltransferase-like polypeptide of the present invention and a transcriptional termination region functional in a host plant cell. Of particular interest is the use of promoters (also referred to as transcriptional initiation regions) functional in plant host cells.

Those skilled in the art will recognize that there are a number of promoters which are functional in plant cells, and have been described in the literature including constitutive, inducible, tissue specific, organelle specific, developmentally regulated and environmentally regulated promoters. Chloroplast and plastid specific promoters, chloroplast or plastid functional promoters, and chloroplast or plastid operable promoters are also envisioned.

One set of promoters are constitutive promoters such as the CaMV35S or FMV35S promoters that yield high levels of expression in most plant organs. Enhanced or duplicated versions of the CaMV35S and FMV35S promoters are useful in the practice of this invention (Odell, *et al.* (1985) *Nature* 313:810-812; Rogers, U.S. Patent Number 5,378, 619). Other useful constitutive promoters include, but are not limited to, the mannopine synthase (*mas*) promoter, the nopaline synthase (*nos*) promoter, and the octopine synthase (*ocs*) promoter.

Useful inducible promoters include heat-shock promoters (Ou-Lee et al. (1986) *Proc. Natl. Acad. Sci. USA* 83: 6815; Ainley et al. (1990) *Plant Mol. Biol.* 14: 949), a nitrate-inducible promoter derived from the spinach nitrite reductase gene (Back et al. (1991) *Plant Mol. Biol.* 17: 9), hormone-inducible promoters (Yamaguchi-Shinozaki et al. (1990) *Plant Mol. Biol.* 15: 905; Kares et al. (1990) *Plant Mol. Biol.* 15: 905), and light-inducible promoters associated with the small subunit of RuBP carboxylase and LHCP gene families (Kuhlemeier et al. (1989) *Plant Cell* 1: 471; Feinbaum et al. (1991) *Mol. Gen. Genet.* 226: 449; Weisshaar et al. (1991) *EMBO J.* 10: 1777; Lam and Chua (1990) *Science* 248: 471; Castresana et al. (1988) *EMBO J.* 7: 1929; Schulze-Lefert et al. (1989) *EMBO J.* 8: 651).

In addition, it may also be preferred to bring about expression of the acyltransferase gene in specific tissues of the plant, such as leaf, stem, root, tuber, seed, fruit, etc., and the promoter chosen should have the desired tissue and developmental specificity. Examples of useful tissue-specific, developmentally-regulated promoters include fruit-specific promoters such as the E4 promoter (Cordes et al. (1989) *Plant Cell* 1:1025), the E8 promoter (Deikman et al. (1988) *EMBO J.* 7: 3315), the kiwifruit actinidin promoter (Lin et al. (1993) *PNAS* 90: 5939), the 2A11 promoter (Houck et al., U.S. Patent 4,943,674), and the tomato pZ130 promoter (U.S. Patents 5,175, 095 and 5,530,185); the β-conglycinin 7S promoter (Doyle et al. (1986) *J. Biol. Chem.* 261: 9228; Slighton and Beachy (1987) *Planta* 172: 356), and seed-specific promoters (Knutzon et al. (1992) *Proc. Natl. Acad. Sci. USA* 89: 2624; Bustos et al. (1991) *EMBO J.* 10: 1469; Lam and Chua (1991) *J. Biol. Chem.* 266: 17131; Stayton et al. (1991) *Aust. J. Plant. Physiol.* 18: 507). Fruit-specific gene regulation is discussed in U.S. Patent 5,753,475. Other useful seed-specific promoters include, but are not limited to, the napin, phaseolin, zein, soybean trypsin inhibitor, 7S, ADR12, ACP, stearoyl-ACP desaturase, oleosin, *Lasquerella* hydroxylase, and barley aldose reductase promoters (Bartels (1995) *Plant J.* 7: 809-822), the EA9 promoter (U.S. Patent 5,420,034), and the Bce4 promoter (U.S. Patent 5,530,194). Useful embryo-specific promoters include the corn globulin 1 and oleosin promoters. Useful endosperm-specific promoters include the rice glutelin-1 promoter, the

promoters for the low-pI β amylase gene (Amy32b) (Rogers et al. (1984) *J. Biol. Chem.* 259: 12234), the high-pI β amylase gene (Amy 64) (Khurseed et al. (1988) *J. Biol. Chem.* 263: 18953), and the promoter for a barley thiol protease gene ("Aleurain") (Whittier et al. (1987) *Nucleic Acids Res.* 15: 2515).

Of particular interest is the expression of the nucleic acid sequences of the present invention from transcription initiation regions which are preferentially expressed in a plant seed tissue. Examples of such seed preferential transcription initiation sequences include those sequences derived from sequences encoding plant storage protein genes or from genes involved in fatty acid biosynthesis in oilseeds. Examples of such promoters include the 5' regulatory regions from such genes as napin (Kridl *et al.*, *Seed Sci. Res. 1*:209:219 (1991)), phaseolin, zein, soybean trypsin inhibitor, ACP, stearoyl-ACP desaturase, soybean α' subunit of β-conglycinin (soy 7s, (Chen *et al.*, *Proc. Natl. Acad. Sci.*, 83:8560-8564 (1986))) and oleosin. Seed-specific gene regulation is discussed in EP 0 255 378 B1 and U.S. Patents 5,420,034 and 5,608,152 . Promoter hybrids can also be constructed to enhance transcriptional activity (Hoffman, U.S. Patent No. 5,106,739), or to combine desired transcriptional activity and tissue specificity.

It may be advantageous to direct the localization of proteins conferring LCAT to a particular subcellular compartment, for example, to the mitochondrion, endoplasmic reticulum, vacuoles, chloroplast or other plastidic compartment. For example, where the genes of interest of the present invention will be targeted to plastids, such as chloroplasts, for expression, the constructs will also employ the use of sequences to direct the gene to the plastid. Such sequences are referred to herein as chloroplast transit peptides (CTP) or plastid transit peptides (PTP). In this manner, where the gene of interest is not directly inserted into the plastid, the expression construct will additionally contain a gene encoding a transit peptide to direct the gene of interest to the plastid. The chloroplast transit peptides may be derived from the gene of interest, or may be derived from a heterologous sequence having a CTP. Such transit peptides are known in the art. See, for example, Von Heijne *et al.* (1991) *Plant Mol. Biol. Rep. 9*:104-126; Clark *et al.* (1989) *J. Biol. Chem. 264*:17544-17550; della-Cioppa *et al.* (1987) *Plant Physiol. 84*:965-968; Romer *et al.* (1993) *Biochem. Biophys. Res Commun. 196*:1414-1421; and, Shah *et al.* (1986) *Science 233*:478-481.

Depending upon the intended use, the constructs may contain the nucleic acid sequence which encodes the entire LCAT protein, a portion of the LCAT protein, the entire ACAT protein, or a portion of the ACAT protein. For example, where antisense inhibition of

a given LCAT or ACAT protein is desired, the entire sequence is not required. Furthermore, where LCAT or ACAT sequences used in constructs are intended for use as probes, it may be advantageous to prepare constructs containing only a particular portion of a LCAT or ACAT encoding sequence, for example a sequence which is discovered to encode a highly conserved region.

The skilled artisan will recognize that there are various methods for the inhibition of expression of endogenous sequences in a host cell. Such methods include, but are not limited to antisense suppression (Smith, *et al.* (1988) *Nature* 334:724-726) , co-suppression (Napoli, *et al.* (1989) *Plant Cell* 2:279-289), ribozymes (PCT Publication WO 97/10328), and combinations of sense and antisense Waterhouse, *et al.* (1998) *Proc. Natl. Acad. Sci. USA* 95:13959-13964. Methods for the suppression of endogenous sequences in a host cell typically employ the transcription or transcription and translation of at least a portion of the sequence to be suppressed. Such sequences may be homologous to coding as well as non-coding regions of the endogenous sequence.

Regulatory transcript termination regions may be provided in plant expression constructs of this invention as well. Transcript termination regions may be provided by the DNA sequence encoding the diacylglycerol acyltransferase or a convenient transcription termination region derived from a different gene source, for example, the transcript termination region which is naturally associated with the transcript initiation region. The skilled artisan will recognize that any convenient transcript termination region which is capable of terminating transcription in a plant cell may be employed in the constructs of the present invention.

Alternatively, constructs may be prepared to direct the expression of the LCAT or ACAT sequences directly from the host plant cell plastid. Such constructs and methods are known in the art and are generally described, for example, in Svab, *et al.* (1990) *Proc. Natl. Acad. Sci. USA* 87:8526-8530 and Svab and Maliga (1993) *Proc. Natl. Acad. Sci. USA* 90:913-917 and in U.S. Patent Number 5,693,507.

A plant cell, tissue, organ, or plant into which the recombinant DNA constructs containing the expression constructs have been introduced is considered transformed, transfected, or transgenic. A transgenic or transformed cell or plant also includes progeny of the cell or plant and progeny produced from a breeding program employing such a transgenic plant as a parent in a cross and exhibiting an altered phenotype resulting from the presence of a LCAT nucleic acid sequence.

Plant expression or transcription constructs having a plant LCAT as the DNA sequence of interest for increased or decreased expression thereof may be employed with a wide variety of plant life, particularly, plant life involved in the production of vegetable oils for edible and industrial uses. Most especially preferred are temperate oilseed crops. Plants of interest include, but are not limited to, rapeseed (Canola and High Erucic Acid varieties), sunflower, safflower, cotton, soybean, peanut, coconut and oil palms, and corn. Depending on the method for introducing the recombinant constructs into the host cell, other DNA sequences may be required. Importantly, this invention is applicable to dicotyledyons and monocotyledons species alike and will be readily applicable to new and/or improved transformation and regulation techniques.

Of particular interest, is the use of plant LCAT and ACAT constructs in plants to produce plants or plant parts, including, but not limited to leaves, stems, roots, reproductive, and seed, with a modified content of lipid and/or sterol esters and to alter the oil production by such plants.

Of particular interest in the present invention, is the use of ACAT genes in conjunction with the LCAT sequences to increase the sterol content of seeds. Thus, overexpression of a nucleic acid sequence encoding an ACAT and LCAT in an oilseed crop may find use in the present invention to increase sterol levels in plant tissues and/or increase oil production.

It is contemplated that the gene sequences may be synthesized, either completely or in part, especially where it is desirable to provide plant-preferred sequences. Thus, all or a portion of the desired structural gene (that portion of the gene which encodes the LCAT or ACAT protein) may be synthesized using codons preferred by a selected host. Host-preferred codons may be determined, for example, from the codons used most frequently in the proteins expressed in a desired host species.

One skilled in the art will readily recognize that antibody preparations, nucleic acid probes (DNA and RNA) and the like may be prepared and used to screen and recover "homologous" or "related" sequences from a variety of plant sources. Homologous sequences are found when there is an identity of sequence, which may be determined upon comparison of sequence information, nucleic acid or amino acid, or through hybridization reactions between a known LCAT and a candidate source. Conservative changes, such as Glu/Asp, Val/Ile, Ser/Thr, Arg/Lys and Gln/Asn may also be considered in determining sequence homology. Amino acid sequences are considered homologous by as little as 25% sequence

identity between the two complete mature proteins. (*See* generally, Doolittle, R.F., *OF URFS and ORFS* (University Science Books, CA, 1986.)

Thus, other LCATs may be obtained from the specific sequences provided herein. Furthermore, it will be apparent that one can obtain natural and synthetic sequences, including modified amino acid sequences and starting materials for synthetic-protein modeling from the exemplified LCAT and ACAT sequences and from sequences which are obtained through the use of such exemplified sequences. Modified amino acid sequences include sequences which have been mutated, truncated, increased and the like, whether such sequences were partially or wholly synthesized. Sequences which are actually purified from plant preparations or are identical or encode identical proteins thereto, regardless of the method used to obtain the protein or sequence, are equally considered naturally derived.

For immunological screening, antibodies to the protein can be prepared by injecting rabbits or mice with the purified protein or portion thereof, such methods of preparing antibodies being well known to those in the art. Either monoclonal or polyclonal antibodies can be produced, although typically polyclonal antibodies are more useful for gene isolation. Western analysis may be conducted to determine that a related protein is present in a crude extract of the desired plant species, as determined by cross-reaction with the antibodies to the encoded proteins. When cross-reactivity is observed, genes encoding the related proteins are isolated by screening expression libraries representing the desired plant species. Expression libraries can be constructed in a variety of commercially available vectors, including lambda gt11, as described in Sambrook, *et al.* (*Molecular Cloning: A Laboratory Manual*, Second Edition (1989) Cold Spring Harbor Laboratory, Cold Spring Harbor, New York).

To confirm the activity and specificity of the proteins encoded by the identified nucleic acid sequences as acyltransferase enzymes, *in vitro* assays are performed in insect cell cultures using baculovirus expression systems. Such baculovirus expression systems are known in the art and are described by Lee, *et al.* U.S. Patent Number 5,348,886, the entirety of which is herein incorporated by reference.

In addition, other expression constructs may be prepared to assay for protein activity utilizing different expression systems. Such expression constructs are transformed into yeast or prokaryotic host and assayed for acyltransferase activity. Such expression systems are known in the art and are readily available through commercial sources.

The method of transformation in obtaining such transgenic plants is not critical to the instant invention, and various methods of plant transformation are currently available.

Furthermore, as newer methods become available to transform crops, they may also be directly applied hereunder. For example, many plant species naturally susceptible to *Agrobacterium* infection may be successfully transformed via tripartite or binary vector methods of *Agrobacterium* mediated transformation. In many instances, it will be desirable to have the construct bordered on one or both sides by T-DNA, particularly having the left and right borders, more particularly the right border. This is particularly useful when the construct uses *A. tumefaciens* or *A. rhizogenes* as a mode for transformation, although the T-DNA borders may find use with other modes of transformation. In addition, techniques of microinjection, DNA particle bombardment, and electroporation have been developed which allow for the transformation of various monocot and dicot plant species.

Normally, included with the DNA construct will be a structural gene having the necessary regulatory regions for expression in a host and providing for selection of transformant cells. The gene may provide for resistance to a cytotoxic agent, e.g. antibiotic, heavy metal, toxin, etc., complementation providing prototrophy to an auxotrophic host, viral immunity or the like. Depending upon the number of different host species the expression construct or components thereof are introduced, one or more markers may be employed, where different conditions for selection are used for the different hosts.

Non-limiting examples of suitable selection markers include genes that confer resistance to bleomycin, gentamycin, glyphosate, hygromycin, kanamycin, methotrexate, phleomycin, phosphinotricin, spectinomycin, streptomycin, sulfonamide and sulfonylureas. Maliga et al., *Methods in Plant Molecular Biology*, Cold Spring Harbor Laboratory Press, 1995, p. 39. Examples of markers include, but are not limited to, alkaline phosphatase (AP), myc, hemagglutinin (HA), β glucuronidase (GUS), luciferase, and green fluorescent protein (GFP).

Where *Agrobacterium* is used for plant cell transformation, a vector may be used which may be introduced into the *Agrobacterium* host for homologous recombination with T-DNA or the Ti- or Ri-plasmid present in the *Agrobacterium* host. The Ti- or Ri-plasmid containing the T-DNA for recombination may be armed (capable of causing gall formation) or disarmed (incapable of causing gall formation), the latter being permissible, so long as the *vir* genes are present in the transformed *Agrobacterium* host. The armed plasmid can give a mixture of normal plant cells and gall.

In some instances where *Agrobacterium* is used as the vehicle for transforming host plant cells, the expression or transcription construct bordered by the T-DNA border region(s)

will be inserted into a broad host range vector capable of replication in *E. coli* and *Agrobacterium*, there being broad host range vectors described in the literature. Commonly used is pRK2 or derivatives thereof. See, for example, Ditta, *et al.*, (*Proc. Nat. Acad. Sci., U.S.A.* (1980) *77*:7347-7351) and EPA 0 120 515, which are incorporated herein by reference. Alternatively, one may insert the sequences to be expressed in plant cells into a vector containing separate replication sequences, one of which stabilizes the vector in *E. coli*, and the other in *Agrobacterium*. See, for example, McBride and Summerfelt (*Plant Mol. Biol.* (1990) *14*:269-276), wherein the pRiHRI (Jouanin, *et al., Mol. Gen. Genet.* (1985) 201:370-374) origin of replication is utilized and provides for added stability of the plant expression vectors in host *Agrobacterium* cells.

Included with the expression construct and the T-DNA can be one or more markers, which allow for selection of transformed Agrobacterium and transformed plant cells. A number of markers have been developed for use with plant cells, such as resistance to chloramphenicol, kanamycin, the aminoglycoside G418, hygromycin, or the like. The particular marker employed is not essential to this invention, one or another marker being preferred depending on the particular host and the manner of construction.

For transformation of plant cells using *Agrobacterium*, explants may be combined and incubated with the transformed *Agrobacterium* for sufficient time for transformation, the bacteria killed, and the plant cells cultured in an appropriate selective medium. Once callus forms, shoot formation can be encouraged by employing the appropriate plant hormones in accordance with known methods and the shoots transferred to rooting medium for regeneration of plants. The plants may then be grown to seed and the seed used to establish repetitive generations and for isolation of vegetable oils.

Thus, in another aspect of the present invention, methods for modifying the sterol and/or stanol composition of a host cell. Of particular interest are methods for modifying the sterol and/or stanol composition of a host plant cell. In general the methods involve either increasing the levels of sterol ester compounds as a proportion of the total sterol compounds. The method generally comprises the use of expression constructs to direct the expression of the polynucleotides of the present invention in a host cell.

Also provided are methods for reducing the proportion of sterol ester compounds as a percentage of total sterol compounds in a host plant cell. The method generally comprises the use of expression constructs to direct the suppression of endogenous acyltransferase proteins in a host cell.

Of particular interest is the use of expression constructs to modify the levels of sterol compounds in a host plant cell. Most particular, the methods find use in modifying the levels of sterol compounds in seed oils obtained from plant seeds.

Also of interest is the use of expression constructs of the present invention to alter oil production in a host cell and in particular to increase oil production. Of particular interest is the use of expression constructs containing nucleic acid sequences encoding LCAT and/or ACAT polypeptides to transform host plant cells and to use these host cells to regenerate whole plants having increase oil production as compared to the same plant not containing the expression construct.

The oils obtained from transgenic plants having modified sterol compound content find use in a wide variety of applications. Of particular interest in the present invention is the use of the oils containing modified levels of sterol compounds in applications involved in improving human nutrition and cardiovascular health. For example, phytostanols are beneficial for lowering serum cholesterol (Ling, *et al.* (1995) *Life Sciences* 57:195-206).

Cholesterol-lowering compositions comprise the oils and sterol ester compound compositions obtained using the methods of the present invention. Such cholesterol lowering compositions include, but are not limited to foods, food products, processed foods, food ingredients, food additive compositions, or dietary/nutritional supplements that contain oils and/or fats. Non-limiting examples include margarines; butters; shortenings; cooking oils; frying oils; dressings, such as salad dressings; spreads; mayonnaises; and vitamin/mineral supplements. Patent documents relating to such compositions include, U.S. Patents 4,588,717 and 5,244,887, and PCT International Publication Nos. WO 96/38047, WO 97/42830, WO 98/06405, and WO 98/06714. Additional non-limiting examples include toppings; dairy products such as cheese and processed cheese; processed meat; pastas; sauces; cereals; desserts, including frozen and shelf-stable desserts; dips; chips; baked goods; pastries; cookies; snack bars; confections; chocolates; beverages; unextracted seed; and unextracted seed that has been ground, cracked, milled, rolled, extruded, pelleted, defatted, dehydrated, or otherwise processed, but which still contains the oils, etc., disclosed herein.

The cholesterol-lowering compositions can also take the form of pharmaceutical compositions comprising a cholesterol-lowering effective amount of the oils or sterol compound compositions obtained using the methods of the present invention, along with a pharmaceutically acceptable carrier, excipient, or diluent. These pharmaceutical

compositions can be in the form of a liquid or a solid. Liquids can be solutions or suspensions; solids can be in the form of a powder, a granule, a pill, a tablet, a gel, or an extrudate. U.S. Patent 5,270,041 relates to sterol-containing pharmaceutical compositions.

Thus, by expression of the nucleic acid sequences encoding acyltransferase-like sequences of the present invention in a host cell, it is possible to modify the lipid content and/or composition as well as the sterol content and/or composition of the host cell.

The invention now being generally described, it will be more readily understood by reference to the following examples which are included for purposes of illustration only and are not intended to limit the present invention.

EXAMPLES

Example 1: RNA Isolations

Total RNA from the inflorescence and developing seeds of *Arabidopsis thaliana* was isolated for use in construction of complementary (cDNA) libraries. The procedure was an adaptation of the DNA isolation protocol of Webb and Knapp (D.M. Webb and S.J. Knapp, (1990) Plant Molec. Reporter, 8, 180-185). The following description assumes the use of 1g fresh weight of tissue. Frozen seed tissue was powdered by grinding under liquid nitrogen. The powder was added to 10ml REC buffer (50mM Tris-HCl, pH 9, 0.8M NaCl, 10mM EDTA, 0.5% w/v CTAB (cetyltrimethyl-ammonium bromide)) along with 0.2g insoluble polyvinylpolypyrrolidone, and ground at room temperature. The homogenate was centrifuged for 5 minutes at 12,000 x*g* to pellet insoluble material. The resulting supernatant fraction was extracted with chloroform, and the top phase was recovered.

The RNA was then precipitated by addition of 1 volume RecP (50mM Tris-HCL pH9, 10mM EDTA and 0.5% (w/v) CTAB) and collected by brief centrifugation as before. The RNA pellet was redissolved in 0.4 ml of 1M NaCl. The RNA pellet was redissolved in water and extracted with phenol/chloroform. Sufficient 3M potassium acetate (pH 5) ws added to make the mixture 0.3M in acetate, followed by addition of two volumes of ethanol to precipitate the RNA. After washing with ethanol, this final RNA precipitate was dissolved in water and stored frozen.

Alternatively, total RNA may be obtained using TRIzol reagent (BRL-Lifetechnologies, Gaithersburg, MD) following the manufacturer's protocol. The RNA precipitate was dissolved in water and stored frozen.

Example 2: Identification of LCAT Sequences

Searches were performed on a Silicon Graphics Unix computer using additional Bioaccellerator hardware and GenWeb software supplied by Compugen Ltd. This software and hardware enabled the use of the Smith-Waterman algorithm in searching DNA and protein databases using profiles as queries. The program used to query protein databases was profilesearch. This is a search where the query is not a single sequence but a profile based on a multiple alignment of amino acid or nucleic acid sequences. The profile was used to query a sequence data set, i.e., a sequence database. The profile contained all the pertinent information for scoring each position in a sequence, in effect replacing the "scoring matrix" used for the standard query searches. The program used to query nucleotide databases with a protein profile was tprofilesearch. Tprofilesearch searches nucleic acid databases using an amino acid profile query. As the search is running, sequences in the database are translated to amino acid sequences in six reading frames. The output file for tprofilesearch is identical to the output file for profilesearch except for an additional column that indicates the frame in which the best alignment occurred.

The Smith-Waterman algorithm, (Smith and Waterman (1981) *J. Molec. Biol.*, 147:195-197), was used to search for similarities between one sequence from the query and a group of sequences contained in the database.

A protein sequence of Lecithin: cholesterol acyltransferase from human (McLean J, *et al.* (1986) *Nucleic Acids Res.* 14(23):9397-406 SEQ ID NO:1)) was used to search the NCBI non-redundant protein database using BLAST. Three sequences were identified from *Arabidopsis*, GenBank accessions AC004557 (referred to herein as LCAT1, SEQ ID NO:2), AC003027 (referred to herein as LCAT2, SEQ ID NO:4), and AL024486 (referred to herein as LCAT3, SEQ ID NO:6). The deduced amino acid sequences are provided in SEQ ID NOs: 3, 5, and 7, respectively.

The profile generated from the queries using PSI-BLAST was excised from the hyper text markup language (html) file. The worldwide web (www)/html interface to psiblast at ncbi stores the current generated profile matrix in a hidden field in the html file that is

returned after each iteration of psiblast. However, this matrix has been encoded into string62 (s62) format for ease of transport through html. String62 format is a simple conversion of the values of the matrix into html legal ascii characters.

The encoded matrix width (x axis) is 26 characters, and comprise the consensus characters, the probabilities of each amino acid in the order A,B,C,D,E,F,G,H,I,K,L,M,N, P,Q,R,S,T,V,W,X,Y,Z (where B represents D and N, and Z represents Q and E, and X represents any amino acid), gap creation value, and gap extension value.

The length (y axis) of the matrix corresponds to the length of the sequences identified by PSI-BLAST. The order of the amino acids corresponds to the conserved amino acid sequence of the sequences identified using PSI-BLAST, with the N-terminal end at the top of the matrix. The probabilities of other amino acids at that position are represented for each amino acid along the x axis, below the respective single letter amino acid abbreviation.

Thus, each row of the profile consists of the highest scoring (consensus) amino acid, followed by the scores for each possible amino acid at that position in sequence matrix, the score for opening a gap that that position, and the score for continuing a gap at that position.

The string62 file is converted back into a profile for use in subsequent searches. The gap open field is set to 11 and the gap extension field is set to 1 along the x axis. The gap creation and gap extension values are known, based on the settings given to the PSI-BLAST algorithm. The matrix is exported to the standard GCG profile form. This format can be read by GenWeb.

The algorithm used to convert the string62 formatted file to the matrix is outlined in Table 1.

Table 1

1. if encoded character z then the value is blast score min
2. if encoded character Z then the value is blast score max
3. else if the encoded character is uppercase then its value is (64-(ascii # of char))
4. else if the encoded character is a digit the value is ((ascii # of char)-48)
5. else if the encoded character is not uppercase then the value is ((ascii # of char) - 87)
6. ALL B positions are set to min of D and N amino acids at that row in sequence matrix
7. ALL Z positions are set to min of Q amd E amino acids at that row in sequence matrix
8. ALL X positions are set to min of all amino acids at that row in sequence matrix
9. kBLAST_SCORE_MAX=999;
10. kBLAST_SCORE_MIN=-999;
11. all gap opens are set to 11
12. all gap lens are set to 1

The protein sequences of LCAT1, LCAT2, and LCAT3 as well as the PSI-BLAST profile were used to search public and proprietary databases for additional LCAT sequences. Two EST sequences were identified which appear to be identical to LCAT1 and LCAT3, respectively. One additional *Arabidopsis* sequence was identified from the proprietary databases, LCAT4 (SEQ ID NO:8). The deduced protein sequence of LCAT4 is provided in SEQ ID NO:9. Two additional genomic sequences were identified using the PSI-BLAST profile from libraries of *Arabidopsis* ecotypes Columbia and Landsberg, LCAT7 (SEQ ID NO:10) and LCAT8 (SEQ ID NO:11). The LCAT7 sequence was present in both the Columbia and Landsberg genomic libraries, while the LCAT8 sequence was only present in the Columbia library.

An open reading frame was predicted from the genomic sequence of LCAT7 in the Arabidopsis public database and this sequence was called MSH12 (referred to herein as LCAT5, SEQ ID NO: 73). The deduced protein sequence of LCAT5 is provided in SEQ ID NO: 74.

The PSI-BLAST profile and the LCAT sequences were used to query the public yeast database and proprietary libraries containing corn and soy EST sequences. The yeast genome contains only one gene, *LRO1* (LCAT Related Open reading frame, YNR008W, Figure 1) with distinct similarity to the human LCAT. The DNA sequence of *LRO1* is provided in SEQ

ID NO: 75 and the protein sequence is provided in SEQ ID NO: 76. Seven EST sequences were identified from soybean libraries as being LCAT sequences. Two sequences from soy (SEQ ID NOs: 12 and 13) are most closely related to the *Arabidopsis* LCAT1 sequence, a single sequence was identified as being most closely related to LCAT2 (SEQ ID NO:14), three were closely related to LCAT3 (SEQ ID NOs: 15-17), and an additional single sequence was identified (SEQ ID NO:18). A total of 11 corn EST sequences were identified as being related to the *Arabidopsis* LCAT sequences. Two corn EST sequences (SEQ ID NOs: 19 and 20) were most closely related to LCAT1, two sequences were identified as closely related to LCAT2 (SEQ ID NOs: 21 and 22), four corn EST sequences were identified as closely related to LCAT3 (SEQ ID NOs: 23-26), and an additional three corn EST sequences were also identified (SEQ ID NOs: 27-29).

Example 3: Identification of ACAT Sequences

Since plant ACATs are unknown in the art, searches were performed to identify known and related ACAT sequences from mammalian sources from public databases. These sequences were then used to search public and proprietary EST databases to identify plant ACAT-like sequences.

A public database containing mouse Expressed Sequence Tag (EST) sequences (dBEST) was searched for ACAT-like sequences. The search identified two sequences (SEQ ID 30 and 31) which were related (approximately 20% identical), but divergent, to known ACAT sequences.

In order to identify ACAT-like sequences from other organisms, the two mouse ACAT sequences were used to search public and proprietary databases containing EST sequences from human and rat tissues. Results of the search identified several sequences from the human database and from the rat database which were closely related to the mouse sequences. The human and rat ACAT-like EST sequences were assembled, using the GCG assembly program, to construct a complete inferred cDNA sequence by identifying overlapping sequences (SEQ ID NOs: 32 and 33, respectively).

The protein sequence of the human ACAT-like sequence was aligned with known ACAT sequences from human (Chang, *et al.* (1993) *J. Biol. Chem.* 268:20747-20755, SEQ ID NO:34), mouse (Uelmen, *et al.* (1995) *J. Biol. Chem.* 270:26192-26201 SEQ ID NO:35) and yeast (Yu, *et al.* (1996) *J. Biol. Chem.* 271:24157-24163, SEQ ID NO:36 and Yang, *et al.*

(1996) *Science* 272:1353-1356, SEQ ID NO:37) using MacVector (Oxford Molecular, Inc.). Results of the alignment demonstrated that the sequence was related to the known sequences, however the related sequence was only about 25% similar to the known sequences.

The protein sequence of the human sterol O-acyltransferase (ACAT, Acyl CoA:Cholesterol acyltransferase, Accession number A48026) related sequence was used to search protein and nucleic acid Genbank databases. A single plant homologue was identified in the public *Arabidopsis* EST database (Accession A042298, SEQ ID NO:38). The protein sequence (SEQ ID NO:39)was translated from the EST sequence, and was found to contain a peptide sequence conserved in both mammalian and yeast ACATs (Chang *et al.*, (1997) *Ann. Rev. Biochem.*, 66:613-638).

To obtain the entire coding region corresponding to the *Arabidopsis* ACAT-like EST, synthetic oligo-nucleotide primers were designed to amplify the 5' and 3' ends of partial cDNA clones containing ACAT-like sequences. Primers were designed according to the *Arabidopsis* ACAT-like EST sequence and were used in Rapid Amplification of cDNA Ends (RACE) reactions (Frohman *et al.* (1988) *Proc. Natl. Acad. Sci. USA* 85:8998-9002).

Primers were designed (5'-TGCAAATTGACGAGCACACCAACCCCTTC-3' (SEQ ID NO:40) and 5'-AAGGATGCTTTGAGTTCCTGACAATAGG-3' (SEQ ID NO:41)) to amplify the 5' end from the Arabidopsis ACAT EST sequence. Amplification of flanking sequences from cDNA clones were performed using the Marathon cDNA Amplification kit (Clontech, CA).

The sequence derived from the 5'-RACE amplification was used to search proprietary *Arabidopsis* EST libraries. A single EST accession, LIB25-088-C7 (SEQ ID NO:42), was identified which contained a sequence identical to the 5'-RACE sequence. Furthermore, LIB25-088-C7 was found to contain the complete putative coding sequence for the *Arabidopsis* ACAT-like product.

The nucleic acid as well as the putative translation product sequences of A042298 were used to search public and proprietary databases. Four EST sequences were identified in both soybean (SEQ ID NOs:43-46) and maize (SEQ ID NOs:47-50) proprietary databases, and a single ACAT-like sequence was identified from *Mortierrella alpina* EST sequences (SEQ ID NO:51).

Sequence alignments between ACAT sequences from several different sources were compared to identify the similarity between the sequences. Nucleotide sequences from

known human and mouse ACATs, as well as nucleotide sequences from known yeast ACATs were compared to the ACAT-like EST sequences from human and *Arabidopsis*.

Analysis of the sequence alignments revealed several classes of ACATs based on sequence similarity. The known human and mouse ACATs, being 88% similar in the nucleotide sequence, formed one class of ACATs. Another class of ACATs included the yeast ACATs which are less than 20% similar to the known human and mouse class ACATs.

The final class of ACATs included the Arabidopsis and human sequences disclosed in the present invention. This class is approximately 22% similar to the known human and mouse ACAT class and approximately 23% similar to the yeast class of ACATs. Thus, the ACAT sequences disclosed in the present invention represent a novel class of ACAT enzymes. Partial mouse sequences of this class are also provided.

Example 4: Expression Construct Preparation

Constructs were prepared to direct expression of the LCAT1, LCAT2, LCAT3, LCAT4, LCAT5 and the yeast LRO1 sequences in plants and cultured insect cells. The entire coding region of each LCAT was amplified from the appropriate EST clone or an Arabidopsis genomic cDNA library using the following oligonucleotide primers in a polymerase chain reactions (PCR). The LCAT1 coding sequence was amplified from the EST clone Lib25-082-Q1-E1-G4 using the primers 5'-GGATCCGCGGCCGCACAATGAAAAAAATATCTTCACATTATTCGG-3' (SEQ ID NO:52) and 5'-GGATCCCCTGCAGGTCATTCATTGACGGCATTAACATTGG-3' (SEQ ID NO:53). The LCAT2 coding sequence was amplified from an Arabidopsis genomic cDNA library using the synthetic oligo nucleotide primers 5'-GGATCCGCGGCCGCACAATGGGAGCGAATTCGAAATCAGTAACG-3' (SEQ ID NO:54) and 5'-GGATCCCCTGCAGGTTAATACCCACTTTTATCAAGCTCCC-3' (SEQ ID NO:55). The LCAT3 coding sequence was amplified from the EST clone LIB22-004-Q1-E1-B4 using the synthetic oligo nucleotide primers 5'-GGATCCGCGGCCGCACAATGTCTCTATTACTGGAA GAGATC-3' (SEQ ID NO:56) and 5'-GGATCCCCTGCAGGTTATGCATC AACAGAGACACTTACAGC-3' (SEQ ID NO:57) . The LCAT4 coding sequence was amplified from the EST clone LIB23-007-Q1-E1-B5 using the synthetic oligo nucleotide primers

5'-GGATCCGCGGCCGCACAATGGGCTGGATTCCGTGTCCGTGC-3' (SEQ ID NO:58) and 5'-GGATCCCCTGCAGGTTAACCAGAATCAACTACTTTGTG-3' (SEQ ID NO:59). The LCAT5 coding sequence was amplified from LIB23-053-Q1-E1-E3 using the synthetic oligo nucleotide primers

5'-GGATCCGCGGCCGCACAATGCCCCTTATTCATCGG-3' (SEQ ID NO:77) and 5'-GGATCCCCTGCAGGTCACAGCTTCAGGTCAATACG-3' (SEQ ID NO:78).

The yeast LROI coding sequence was amplified from genomic yeast DNA using the synthetic oligo nucleotide primers

5'GGATCCGCGGCCGCACAATGGGCACACTGTTTCGAAG3' (SEQ ID NO:79) and 5'GGATCCCCTGCAGGTTACATTGGGAAGGGCATCTGAG3' (SEQ ID NO:80).

The entire coding region of the *Arabidopsis* ACAT sequence (SEQ ID NO: 42) was amplified from the EST clone LIB25-088-C7 using oligonucleotide primers 5'-TCGACCTGCAGGAAGCTTAGAAATGGCGATTTTGGATTC-3' (SEQ ID NO: 60) and 5'-GGATCCGCGGCCGCTCATGACATCGATCCTTTTCGG-3' (SEQ ID NO: 61) in a polymerase chain reaction (PCR).

Each resulting PCR product was subcloned into pCR2.1Topo (Invitrogen) and labeled pCGN9964 (LCAT1), pCGN9985 (LCAT2), pCGN9965 (LCAT3), pCGN9995 (LCAT4), pCGN10964 (LCAT5), pCGN10963 (*LRO1*), and pCGN8626 (ACAT). Double stranded DNA sequence was obtained to verify that no errors were introduced by the PCR amplification.

4A. Baculovirus Expression Constructs

Constructs are prepared to direct the expression of the *Arabidopsis* LCAT and yeast LCAT sequences in cultured insect cells. The entire coding region of the LCAT proteins was removed from the respective constructs by digestion with *Not*I and *Sse*8387I, followed by gel electrophoresis and gel purification. The fragments containing the LCAT coding sequences were cloned into *Not*I and *Pst*I digested baculovirus expression vector pFastBac1 (Gibco-BRL, Gaithersburg, MD). The resulting baculovirus expression constructs were referred to as pCGN9992 (LCAT1), pCGN9993 (LCAT2), pCGN9994 (LCAT3), pCGN10900 (LCAT4), pCGN10967 (LCAT5), and pCGN10962 (*LRO1*).

4B. Plant Expression Construct Preparation

A plasmid containing the napin cassette derived from pCGN3223 (described in U.S. Patent No. 5,639,790, the entirety of which is incorporated herein by reference) was modified to make it more useful for cloning large DNA fragments containing multiple restriction sites, and to allow the cloning of multiple napin fusion genes into plant binary transformation vectors. An adapter comprised of the self annealed oligonucleotide of sequence 5'-CGCGATTTAAATGGCGCGCCCTGCAGGCGGCCGCCTGCAGGGCGCGCCATTTAA AT-3' (SEQ ID NO:62) was ligated into the cloning vector pBC SK+ (Stratagene) after digestion with the restriction endonuclease BssHII to construct vector pCGN7765. Plamids pCGN3223 and pCGN7765 were digested with NotI and ligated together. The resultant vector, pCGN7770, contained the pCGN7765 backbone with the napin seed specific expression cassette from pCGN3223.

The cloning cassette, pCGN7787, contained essentially the same regulatory elements as pCGN7770, with the exception of the napin regulatory regions of pCGN7770 have been replaced with the double CAMV 35S promoter and the tml polyadenylation and transcriptional termination region.

A binary vector for plant transformation, pCGN5139, was constructed from pCGN1558 (McBride and Summerfelt, (1990) Plant Molecular Biology, 14:269-276). In pCGN5139, the polylinker of pCGN1558 was replaced as a HindIII/Asp718 fragment with a polylinker containing unique restriction endonuclease sites, AscI, PacI, XbaI, SwaI, BamHI,and NotI. The Asp718 and HindIII restriction endonuclease sites are retained in pCGN5139.

A series of turbo binary vectors was constructed to allow for the rapid cloning of DNA sequences into binary vectors containing transcriptional initiation regions (promoters) and transcriptional termination regions.

The plasmid pCGN8618 was constructed by ligating oligonucleotides 5'-TCGAGGATCCGCGGCCGCAAGCTTCCTGCAGG-3' (SEQ ID NO:63) and 5'-TCGACCTGCAGGAAGCTTGCGGCCGCGGATCC-3' (SEQ ID NO:64) into SalI/XhoI-digested pCGN7770. A fragment containing the napin promoter, polylinker and napin 3' region was excised from pCGN8618 by digestion with Asp718I; the fragment was blunt-ended by filling in the 5' overhangs with Klenow fragment then ligated into pCGN5139 that had been digested with Asp718I and HindIII and blunt-ended by filling in the 5'

overhangs with Klenow fragment. A plasmid containing the insert oriented so that the napin promoter was closest to the blunted Asp718I site of pCGN5139 and the napin 3' was closest to the blunted HindIII site was subjected to sequence analysis to confirm both the insert orientation and the integrity of cloning junctions. The resulting plasmid was designated pCGN8622.

The plasmid pCGN8619 was constructed by ligating oligonucleotides 5'-TCGACCTGCAGGAAGCTTGCGGCCGCGGATCC-3' (SEQ ID NO:65) and 5'-TCGAGGATCCGCGGCCGCAAGCTTCCTGCAGG-3' (SEQ ID NO:66) into SalI/XhoI-digested pCGN7770. A fragment containing the napin promoter, polylinker and napin 3' region was removed from pCGN8619 by digestion with Asp718I; the fragment was blunt-ended by filling in the 5' overhangs with Klenow fragment then ligated into pCGN5139 that had been digested with Asp718I and HindIII and blunt-ended by filling in the 5' overhangs with Klenow fragment. A plasmid containing the insert oriented so that the napin promoter was closest to the blunted Asp718I site of pCGN5139 and the napin 3' was closest to the blunted HindIII site was subjected to sequence analysis to confirm both the insert orientation and the integrity of cloning junctions. The resulting plasmid was designated pCGN8623.

The plasmid pCGN8620 was constructed by ligating oligonucleotides 5'-TCGAGGATCCGCGGCCGCAAGCTTCCTGCAGGAGCT -3' (SEQ ID NO:67) and 5'-CCTGCAGGAAGCTTGCGGCCGCGGATCC-3' (SEQ ID NO:68) into SalI/SacI-digested pCGN7787. A fragment containing the d35S promoter, polylinker and tml 3' region was removed from pCGN8620 by complete digestion with Asp718I and partial digestion with NotI. The fragment was blunt-ended by filling in the 5' overhangs with Klenow fragment then ligated into pCGN5139 that had been digested with Asp718I and HindIII and blunt-ended by filling in the 5' overhangs with Klenow fragment. A plasmid containing the insert oriented so that the d35S promoter was closest to the blunted Asp718I site of pCGN5139 and the tml 3' was closest to the blunted HindIII site was subjected to sequence analysis to confirm both the insert orientation and the integrity of cloning junctions. The resulting plasmid was designated pCGN8624.

The plasmid pCGN8621 was constructed by ligating oligonucleotides 5'-TCGACCTGCAGGAAGCTTGCGGCCGCGGATCCAGCT-3' (SEQ ID NO:69) and 5'-GGATCCGCGGCCGCAAGCTTCCTGCAGG-3' (SEQ ID NO:70) into SalI/SacI-digested pCGN7787. A fragment containing the d35S promoter, polylinker and tml 3' region

was removed from pCGN8621 by complete digestion with Asp718I and partial digestion with NotI. The fragment was blunt-ended by filling in the 5' overhangs with Klenow fragment then ligated into pCGN5139 that had been digested with Asp718I and HindIII and blunt-ended by filling in the 5' overhangs with Klenow fragment. A plasmid containing the insert oriented so that the d35S promoter was closest to the blunted Asp718I site of pCGN5139 and the tml 3' was closest to the blunted HindIII site was subjected to sequence analysis to confirm both the insert orientation and the integrity of cloning junctions. The resulting plasmid was designated pCGN8625.

The plasmid construct pCGN8640 is a modification of pCGN8624 described above. A 938bp PstI fragment isolated from transposon Tn7 which encodes bacterial spectinomycin and streptomycin resistance (Fling et al. (1985), *Nucleic Acids Research* 13(19):7095-7106), a determinant for E. coli and Agrobacterium selection, was blunt ended with Pfu polymerase. The blunt ended fragment was ligated into pCGN8624 that had been digested with SpeI and blunt ended with Pfu polymerase. The region containing the PstI fragment was sequenced to confirm both the insert orientation and the integrity of cloning junctions.

The spectinomycin resistance marker was introduced into pCGN8622 and pCGN8623 as follows. A 7.7 Kbp AvrII-SnaBI fragment from pCGN8640 was ligated to a 10.9 Kbp AvrII-SnaBI fragment from pCGN8623 or pCGN8622, described above. The resulting plasmids were pCGN8641 and pCGN8643, respectively.

The plasmid pCGN8644 was constructed by ligating oligonucleotides 5'-GATCACCTGCAGGAAGCTTGCGGCCGCGGATCCAATGCA-3' (SEQ ID NO:71) and 5'-TTGGATCCGCGGCCGCAAGCTTCCTGCAGGT-3' (SEQ ID NO:72) into BamHI-PstI digested pCGN8640.

4C. Plant LCAT Expression Construct Preparation

The coding sequence of LCAT1 was cloned from pCGN9964 as a *NotI*/ *Sse*8387I fragment into pCGN8640, pCGN8641, pCGN8643, and pCGN8644 to create the expression constructs pCGN9960, pCGN9961, pCGN9962, and pCGN9963, respectively. The construct pCGN9960 was designed to express the LCAT1 coding sequence in the sense orientation from the constitutive promoter CaMV 35S. The construct pCGN9961 was designed to express the LCAT1 coding sequence in the antisense orientation from the napin promoter. The construct pCGN9962 was designed to express the LCAT1 coding sequence in the sense

orientation from the napin promoter. The construct pCGN9963 was designed to express the LCAT1 coding sequence in the antisense orientation from the constitutive promoter CaMV 35S.

The coding sequence of LCAT2 was cloned from pCGN9985 as a *NotI*/ *Sse*8387I fragment into pCGN8640, pCGN8641, pCGN8643, and pCGN8644 to create the expression constructs pCGN9981, pCGN9982, pCGN9983, and pCGN9984, respectively. The construct pCGN9981 was designed to express the LCAT2 coding sequence in the sense orientation from the constitutive promoter CaMV 35S. The construct pCGN9982 was designed to express the LCAT2 coding sequence in the antisense orientation from the napin promoter. The construct pCGN9983 was designed to express the LCAT2 coding sequence in the sense orientation from the napin promoter. The construct pCGN9984 was designed to express the LCAT2 coding sequence in the antisense orientation from the constitutive promoter CaMV 35S.

The coding sequence of LCAT3 was cloned from pCGN9965 as a *NotI*/ *Sse*8387I fragment into pCGN8640, pCGN8641, pCGN8643, and pCGN8644 to create the expression constructs pCGN9966, pCGN9967, pCGN9968, and pCGN9969, respectively. The construct pCGN9966 was designed to express the LCAT3 coding sequence in the sense orientation from the constitutive promoter CaMV 35S. The construct pCGN9967 was designed to express the LCAT3 coding sequence in the antisense orientation from the napin promoter. The construct pCGN9968 was designed to express the LCAT3 coding sequence in the sense orientation from the napin promoter. The construct pCGN9969 was designed to express the LCAT3 coding sequence in the antisense orientation from the constitutive promoter CaMV 35S.

The coding sequence of LCAT4 was cloned from pCGN9995 as a *NotI*/ *Sse*8387I fragment into pCGN8640, pCGN8641, pCGN8643, and pCGN8644 to create the expression constructs pCGN9996, pCGN9997, pCGN9998, and pCGN9999, respectively. The construct pCGN9996 was designed to express the LCAT4 coding sequence in the sense orientation from the constitutive promoter CaMV 35S. The construct pCGN9997 was designed to express the LCAT4 coding sequence in the antisense orientation from the napin promoter. The construct pCGN9998 was designed to express the LCAT4 coding sequence in the sense orientation from the napin promoter. The construct pCGN9999 was designed to express the LCAT4 coding sequence in the antisense orientation from the constitutive promoter CaMV 35S.

The coding sequence of LCAT5 was cloned from pCGN10964 as a *NotI*/ *Sse*8387I fragment into pCGN9977 and pCGN9979, to create the expression constructs pCGN10965, and pCGN10966, respectively. The construct pCGN10965 was designed to express the LCAT5 coding sequence in the sense orientation from the constitutive promoter CaMV 35S. The construct pCGN10966 was designed to express the LCAT5 coding sequence in the sense orientation from the napin promoter.

The coding sequence of *LRO1* was cloned from pCGN10963 as a *NotI*/ *Sse*8387I fragment into pCGN9977 and pCGN9979, to create the expression constructs pCGN10960, and pCGN10961, respectively. The construct pCGN10960 was designed to express the *LRO1* coding sequence in the sense orientation from the constitutive promoter CaMV 35S. The construct pCGN10961 was designed to express the *LRO1* coding sequence in the sense orientation from the napin promoter.

4D. Plant ACAT Expression Construct Preparation

A fragment containing the *Arabidopsis* ACAT-like coding region was removed from pCGN8626 by digestion with Sse8387I and Not I. The fragment containing the ACAT-like sequence was ligated into PstI-Not I digested pCGN8622. The resulting plasmid was designated pCGN8627. DNA sequence analysis confirmed the integrity of the cloning junctions.

A fragment containing the *Arabidopsis* ACAT-like coding region (SEQ ID NO: 42) was removed from pCGN8626 by digestion with Sse8387I and Not I. The fragment was ligated into PstI-Not I digested pCGN8623. The resulting plasmid was designated pCGN8628. DNA sequence analysis confirmed the integrity of the cloning junctions.

A fragment containing the *Arabidopsis* ACAT-like coding region was removed from pCGN8626 by digestion with Sse8387 and Not I. The fragment was ligated into PstI-Not I digested pCGN8624. The resulting plasmid was designated pCGN8629. DNA sequence analysis confirmed the integrity of the cloning junctions.

A fragment containing the *Arabidopsis* ACAT-like coding region was removed from pCGN8626 by digestion with Sse8387 and Not I. The fragment was ligated into PstI-Not I digested pCGN8625. The resulting plasmid was designated pCGN8630. DNA sequence analysis confirmed the integrity of the cloning junctions.

An additional expression construct for the suppression of endogenous ACAT-like activity was also prepared. The construct pCGN8660 was constructed by cloning approximately 1 Kb of the *Arabidopsis* ACAT-like coding region from pCGN8626 in the sense orientation, and the full-length *Arabidopsis* ACAT-like coding region in the antisense orientation under the regulatory control of the napin transcription initiation sequence.

For expression of the rat ACAT-like sequence in plants, the NotI-Sse8387I fragment of pCGN8592 was cloned into NotI-PstI digested binary vectors pCGN8621, pCGN8622, and pCGN8624 to yield plasmids, pCGN 9700, pCGN9701, and pCGN9702, respectively. Plasmid pCGN9700 expresses a sense transcript of the rat ACAT-like cDNA under control of a napin promoter, plasmid pCGN9701 expresses an antisense transcript of the rat ACAT-like cDNA under control of a napin promoter, and plasmid pCGN9702 expresses a sense transcript of the rat ACAT-like cDNA under control of a double 35S promoter. Plasmids pCGN 9700, pCGN9701, and pCGN9702 were introduced in *Agrobacterium tumefaciens* EHA101.

Constructs were prepared to direct the expression of the rat ACAT-like sequence in the seed embryo of soybean and the endosperm of corn. For expression of the rat ACAT-like DNA sequence in soybean, a 1.5 kb *NotI*/*Sse*8387I fragment from pCGN8592 containing the coding sequence of the rat ACAT-like sequence was blunt ended using Mung bean nuclease, and ligated into the *Sma*I site of the turbo 7S binary/cloning vector pCGN8809 to create the vector pCGN8817 for transformation into soybean by particle bombardment. The vector pCGN8817 contained the operably linked components of the promoter region of the soybean α' subunit of β-conglycinin (7S promoter, (Chen *et al.*, (1986), *Proc. Natl. Acad. Sci.*, 83:8560-8564), the DNA sequence coding for the entire rat ACAT-like protein, and the transcriptional termination region of pea RuBisCo small subunit, referred to as E9 3' (Coruzzi, *et al.* (1984) *EMBO J.* 3:1671-1679 and Morelli, *et al.* (1985) *Nature* 315:200-204). This construct further contained sequences for the selection of positive transformed plants by screening for resistance to glyphosate using the CP4 EPSPS (U.S. Patent 5,633,435) expressed under the control of the figwort mosaic virus (FMV) promoter (U.S. Patent Number 5,378,619) and the transcriptional termination region of E9.

For expression of the rat ACAT-like sequence in the corn endosperm, a 1.5 kb *NotI*/*Sse*8387I fragment from pCGN8592 containing the coding sequence of the rat ACAT-like sequence was blunt ended using Mung bean nuclease, and ligated into the *Bam*HI site of the rice pGt1 expression cassette pCGN8592 for expression from the pGt1 promoter (Leisy,

D.J. et al., Plant Mol. Biol. 14 (1989) 41-50) and the HSP70 intron sequence (U.S. Patent Number 5,593,874). This cassette also included the transcriptional termination region downstream of the cloning site of nopaline synthase, *nos* 3' (Depicker et al., *J. Molec. Appl. Genet.* (1982) 1: 562-573). A 7.5 kb fragment containing the pGt1 promoter, the DNA sequence encoding the rat ACAT-like protein, and the *nos* transcriptional termination sequence was cloned into the binary vector pCGN8816 to create the vector pCGN8818 for transformation into corn. This construct also contained sequences for the selection of positive transformants with kanamycin using the kanamycin resistance gene from Tn5 bacteria under the control of the CAMV 35S promoter and tml transcriptional termination regions.

Example 5: Expression in Insect Cell Culture

A baculovirus expression system was used to express the LCAT cDNAs in cultured insect cells.

The baculovirus expression constructs pCGN9992, pCGN9993, pCGN9994, pCGN10900, pCGN10962, and pCGN10967 were transformed and expressed using the BAC-to-BAC Baculovirus Expression System (Gibco-BRL, Gaithersburg, MD) according to the manufacturer's directions.

The transformed insect cells were used to assay for acyltransferase activities using methods known in the art (see Example 8).

Example 6: Plant Transformation

A variety of methods have been developed to insert a DNA sequence of interest into the genome of a plant host to obtain the transcription or transcription and translation of the sequence to effect phenotypic changes. Transgenic plants were obtained by *Agrobacterium*-mediated transformation as described by Radke *et al.* (*Theor. Appl. Genet.* (1988) *75*:685-694; *Plant Cell Reports* (1992) *11*:499-505). Alternatively, microprojectile bombardment methods, such as described by Klein *et al.* (*Bio/Technology 10*:286-291) may also be used to obtain nuclear transformed plants. Other plant species may be similarly transformed using related techniques.

The plant binary constructs described above were used in plant transformation to direct the expression of the sterol acyltransferases in plant tissues. Binary vector constructs were transformed into strain EHA101 *Agrobacterium* cells (Hood *et al., J. Bacteriol* (1986) *168*:1291-

1301), by the method of Holsters *et al.* (*Mol. Gen. Genet.* (1978) *163*:181-187). Transgenic *Arabidopsis thaliana* plants were obtained by *Agrobacterium*-mediated transformation as described by Valverkens *et al.*, (*Proc. Nat. Acad. Sci.* (1988) *85*:5536-5540), Bent et al. ((1994), *Science* 265:1856-1860), and Bechtold et al. ((1993), *C. R.Acad. Sci., Life Sciences* 316:1194-1199).

EXAMPLE 7: Plant Assays for Modified Sterol Content/Profile

7a: NMR of T2 seed

Seed from plants expressing LCAT 1 through 4 under the control of the napin promoter were analyzed by NMR. Arabidopsis seeds from transgenic plants were placed directly into wide-mouth MAS NMR sample tubes.

High-resolution spectra were measured at 11.7 T (1H=500 MHz, 13C=125 mHz) using Varian NMR Instruments (Palo Alto, CA) Inova™ NMR spectrometers equipped with carbon-observe MAS Nanoprobes™. The 13C spectra were acquired without a field-frequency lock at ambient temperature (approx. 21-22°C) for 14 hours using the following conditions: spectral width = 29.996 kHz, acquisition time = 2.185 seconds, p/2 pulse (3.8 ms) with no relaxation delay, 1H g B2 = 2.5 kHz with Waltz decoupling. Data processing conditions were typically: digital resolution = 0.11 Hz, 0.3 to 1.5 Hz line broadening and time-reversed linear prediction of the first three data points. Chemical shifts were referenced by adding neat tetramethylsilane (TMS) to Arabidopsis seeds and using the resulting referencing parameters for subsequent spectra. The 13C resolution was 2-3 Hz for the most narrow seed resonances. Spectral resolution was independent of MAS spinning speeds (1.5-3.5 kHz) and data were typically obtained with 1.5 kHz spinning speeds. Spinning sidebands were approx. 1% of the main resonance. Phytosterol 13C assignments were based on model samples composed of triolein, β-sitosterol and cholesterol oleate. Triacylglycerol 13C assignments were made from comparison with literature assignments or with shifts computed from a 13C NMR database (Advanced Chemical Development, Inc., version 3.50, Toronto Canada).

The results of these analyses are displayed in Figure 2 and show that there was a trend of an approximately 2 fold increase of phytosterols in the seeds derived from plant line 5 expressing the LCAT 4 gene (pCGN9998) under the control of the napin promoter. During the course of this analysis it was also noted that the average oil content of seed from plants

expressing the LCAT2 construct (pCGN9983) under the control of the napin promoter was higher than that of controls. This is the first *in planta* evidence supporting the concept that overexpression of a nucleotide sequence encoding a lecithin:cholesterol acyltransferase-like polypeptide can increase oil content.

7b: HPLC/MS of T2 seed

Seed oil from T2 plants expressing LCAT1 through 4 under the control of the napin promoter was extracted using an accelerated solvent extractor (ASE) method. Seed samples were ground, using a mortar and pestle, to achieve a fine homogeneous meal. Oil was obtained using a Dionex Accelerated Solvent Extractor (ASE). Clean ground seed was added to an equal amount of diatomaceous earth. The ground seed sample and the diatomaceous earth were thoroughly mixed until a homogeneous texture was achieved. The sample was then loaded into the instrument and oil extraction was achieved using hexane under validated laboratory protocols.

Oil from these seed samples was then analyzed for sterol ester analysis using HPLC/MS for free campesterol, stigmasterol, and sitosterol and their fatty acid esters. To the autosampler vial containing approximately 0.1 grams oil was added 0.3 mLs $CDCl_3$. One-hundred microliters of this solution was added to 900 microliters $CHCl_3$. Five microliters of this diluted sample was subsequently injected into an HPLC/MS with positive ion atmospheric pressure ionization. The individual components in the oils were separated using two 4.6 x 50 mm C_8 Zorbax columns in series and a gradient using acetonitrile and acetonitrile with 40% $CHCl_3$. The sterol concentrations were calculated assuming each sterol and its fatty acids have the same molar responses. This was observed to be the case with cholesterol and its esters and was assumed to be the case for campesterol, stigmasterol, and sitosterol. In the present study, the sterol identified as stigmasterol was actually an isomer of this compound.

The results of these analyses are displayed in Figures 3 and 4 and show that there were sterol ester enhancements on the order of 50%. in the seeds derived from six out of seven T2 plant lines expressing LCAT3 (pCGN9968) under the control of the napin promoter.

EXAMPLE 8: Baculovirus Insect Cell Culture for Sterol Esterification Activity

Baculovirus expression construct pCGN9992, pCGN9993, pCGN9994 and pCGN10900 (see Example 4) were transformed and expressed using the BAC-TOBAC Baculovirus Expression System (Gibco-BRL, Gaithersburg, MD) according to the manufacturer's instructions except harvesting of recombinant viruses was done 5 days post-transfection. The supernatant from the transfection mixture was used for generating virus stock which in turn was used for infecting Sf9 cells used in the assay.

The transformed cells were assayed for lecithin:sterol acyltransferase activities using the method described herein. Insect cells were centrifuged and the resulting cell pellet was either used immediately or stored at -70 C for later analysis. Cells were resuspended in Medium A (100 mM Tricine/NaOH, pH 7.8, 10% (w/v) glycerol, 280 mM NaCl with : 0.1 μM Aprotinin, 1 μM Leupeptin, and 100 μM Pefabloc (all from Boehringer Mannheim, Germany) and lysed by sonication (2 x 10 sec). Cell walls and other debris were pelleted by centrifugation (14,000 x g , 10 min, 4°C). The supernatant was transferred to a new vial and membranes pelleted by centrifugation (100,000 x g, Ti 70.1 rotor, 46,000 rpm for 1 hour at 4°C). Total membranes were resuspended in Medium A. Lecithin:sterol acyltransferase activity was assayed in a 0.1 ml reaction mixture containing 100 mM Tris/HCl, pH 7, 28 mM NaCl, 0.03% Triton X-100, 0.1 mM sitosterol, 20 μM 1,2-[^{14}C]-palmitoyl-phosphatidyl choline (246420 dpm/nmole), and 0.05-20 mg of membrane protein. After 15 minutes at 30 °C, the reaction was terminated by addition of a 0.5 ml solution of methylene chloride:methanol (4:1, v/v) containing 100 μg cholesterol and cholesterol ester as cold carriers. A portion (0.1 ml) of the bottom organic layer was removed and evaporated under nitrogen gas. The concentrated extract was resuspended in 30 μl of hexane and spotted onto a silica gel-G thin layer chromatographic plate. The plate was migrated in hexane:diethyl ether:acetic acid (80:20:1) to the top, then air dried. Radioactivity was determined by exposure to a Low Energy Phosphor-imaging Screen. Following exposure, the screen was read on a phosphorimager.

The LCAT 4 protein from pCGN10900 in baculovirus membranes showed a radioactive spot in the region of the TLC plate where cholesterol ester migrates indicating that LCAT 4 has the ability to catalyze the transfer of an acyl group from lecithin (PC) to sitosterol to make a sitosterol ester.

EXAMPLE 9: Plant Assay for Modified Lipid Content

Nir (near infrared spectroscopy spectral scanning) can be used to determine the total oil content of Arabidopsis seeds in a non-destructive way provided that a spectral calibration curve has been developed and validated for seed oil content. A seed oil spectral calibartion curve was developed using seed samples from 85 Arabidopsis plants. Seed was cleaned and scanned using a Foss NIR model 6500 (Foss-Nirs Systems, Inc.). Approximately 50 to 100 milligrams of whole seeds, per sample, were packed in a mini sample ring cup with quartz lens [IH-0307] consisting a mini-insert [IH-0337] and scanned in reflectance mode to obtain the spectral data. The seed samples were then ground, using a mortar and pestle, to achieve a fine homogeneous meal. The ground samples were measured for oil using an accelerated solvent extractor (ASE).

Measurement for the total oil content was performed on the Dionex Accelerated Solvent Extractor (ASE). Approximately 500 mg of clean ground seed was weighed to the nearest 0.1 mg onto a 9 x 9 cm weigh boat. An equal amount of diatomaceous earth was added using a top-loading balance accurate to the nearest 0.01 g. The ground seed sample and the diatomaceous earth were thoroughly mixed until a homogeneous texture was achieved. The sample was loaded on to the instrument and oil extraction was achieved using hexane under validated laboratory protocols. Standard Rapeseed samples were obtained from the Community Bureau of Reference (BCR). The ASE extraction method was validated using the BCR reference standards. A total percent oil recovery of 99% to 100% was achieved. "As-is" oil content was calculated to the nearest 0.01 mass percentage using the formula:

Oil Content = 100% x (vial plus extracted oil wt - initial vial wt) / (sample wt)

The analytical data generated by ASE were used to perform spectral calibrations. Nir calibration equations were generated using the built-in statistical package within the NirSytems winisi software. The spectral calibration portion of the software is capable of calibration and self-validation. From a total of 85 samples, 57 samples were used to generate the total percent oil calibration. The remaining samples were used to validate the oil calibrations. Optimized smoothing, derivative size, and mathematical treatment (modified partial least square) was utilized to generate the calibration. The samples that were not used

in building respective calibrations were used as a validation set. Statistical tools such as correlation coefficient (R), coefficient of determination (R^2), standard error of prediction (SEP), and the standard error of prediction corrected for bias (SEPC) were used to evaluate the calibration equations.

T2 seeds from plants that had been transformed with the LCAT genes were cleaned and scanned using a Foss NIR model 6500 (Foss-Nirs Systems, Inc.). Approximately 50 to 100 milligrams of whole seeds, per sample, were packed in a mini sample ring cup with quartz lens [IH-0307] consisting a mini-insert [IH-0337] and scanned in reflectance mode to obtain the spectral data. Oil percentage in each seed sample was determined using the seed oil spectral calibration curve detailed above.

The results of these analyses are found in Figure 5 and Table 2 and show that there was a significant increase in the oil level in seed from T2 plants expressing the LCAT2 gene. This increase in oil was seen in plants when LCAT2 was driven by either the 35S constitutive promoter or the seed-specific napin promoter. These results show that overexpression of a nucleic acid sequence encoding a lecithin:cholesterol acyltransferase-like polypeptide can increase seed oil production in plants.

Table 2

	Construct number	Seed Oil Percentage (%)
CONTROL		24.7
CONTROL		28.0
CONTROL		31.8
CONTROL		32.4
NAPIN LCAT1	PCGN9962	28.5
NAPIN LCAT1	PCGN9962	28.9
NAPIN LCAT1	PCGN9962	29.6
NAPIN LCAT1	PCGN9962	30.1
NAPIN LCAT1	PCGN9962	30.1
NAPIN LCAT1	PCGN9962	30.1
NAPIN LCAT1	PCGN9962	30.8
NAPIN LCAT1	PCGN9962	31.0
NAPIN LCAT1	pCGN9962	32.1
NAPIN LCAT1	pCGN9962	34.2
NAPIN LCAT3	pCGN9968	26.8
NAPIN LCAT3	pCGN9968	27.4
NAPIN LCAT3	pCGN9968	29.0
NAPIN LCAT3	pCGN9968	29.0
NAPIN LCAT3	pCGN9968	32.6
NAPIN LCAT2	pCGN9983	26.5
NAPIN LCAT2	pCGN9983	34.7
NAPIN LCAT2	pCGN9983	34.8
NAPIN LCAT2	pCGN9983	35.7
NAPIN LCAT2	pCGN9983	35.8
NAPIN LCAT2	pCGN9983	36.3
NAPIN LCAT2	pCGN9983	36.7
NAPIN LCAT2	pCGN9983	37.0
NAPIN LCAT2	pCGN9983	37.2
NAPIN LCAT2	pCGN9983	37.3
NAPIN LCAT2	pCGN9983	37.3
NAPIN LCAT2	pCGN9983	37.4
NAPIN LCAT2	pCGN9983	37.8
NAPIN LCAT2	pCGN9983	38.0
NAPIN LCAT2	pCGN9983	38.0
35S LCAT2	pCGN9981	27.3
35S LCAT2	pCGN9981	28.1
35S LCAT2	pCGN9981	28.2
35S LCAT2	pCGN9981	28.6
35S LCAT2	pCGN9981	29.8
35S LCAT2	pCGN9981	30.3
35S LCAT2	pCGN9981	32.4
35S LCAT2	pCGN9981	32.5
35S LCAT2	pCGN9981	33.6
35S LCAT2	pCGN9981	34.1
35S LCAT2	pCGN9981	35.5
35S LCAT2	pCGN9981	36.4

35S LCAT2	pCGN9981	37.1
35S LCAT2	pCGN9981	38.3
35S LCAT2	pCGN9981	38.5
35S LCAT2	pCGN9981	39.1

In light of the detailed description of the invention and the examples presented above, it can be appreciated that the several aspects of the invention are achieved.

It is to be understood that the present invention has been described in detail by way of illustration and example in order to acquaint others skilled in the art with the invention, its principles, and its practical application. Particular formulations and processes of the present invention are not limited to the descriptions of the specific embodiments presented, but rather the descriptions and examples should be viewed in terms of the claims that follow and their equivalents. While some of the examples and descriptions above include some conclusions about the way the invention may function, the inventors do not intend to be bound by those conclusions and functions, but put them forth only as possible explanations.

It is to be further understood that the specific embodiments of the present invention as set forth are not intended as being exhaustive or limiting of the invention, and that many alternatives, modifications, and variations will be apparent to those of ordinary skill in the art in light of the foregoing examples and detailed description. Accordingly, this invention is intended to embrace all such alternatives, modifications, and variations that fall within the spirit and scope of the following claims.

What is claimed is:

1. An isolated nucleic acid sequence comprising a polynucleotide encoding a plant lecithin:cholesterol acyltransferase-like polypeptide or fragment thereof.

2. The isolated nucleic acid sequence of claim 1, wherein said plant lecithin:cholesterol acyltransferase-like polypeptide is selected from the group consisting of *Arabidopsis*, soybean and corn.

3. An isolated nucleic acid sequence comprising a polynucleotide encoding a plant acyl CoA:cholesterol acyltransferase-like polypeptide.

4. The isolated nucleic acid sequence of claim 3, wherein said polynucleotide is SEQ ID NO: 42 or degenerate variants thereof.

5. The isolated nucleic acid sequence of claim 1, wherein said polynucleotide is selected from the group consisting of SEQ ID NO: 2, 4, 6, 8, 10, 11, 12, 13, 14, 15, 16, 17, 18, 19, 20, 21, 22, 23, 24, 25, 26, 27, 28, 29, 43, 44, 45, 46, 47, 48, 49, 50, 51, 73 and 75 or degenerate variants thereof.

6. An isolated nucleic acid sequence consisting essentially of SEQ ID NO: 2, 4, 6, 8, 10, 11, 12, 13, 14, 15, 16, 17, 18, 19, 20, 21, 22, 23, 24, 25, 26, 27, 28, 29, 42, 43, 44, 45, 46, 47, 48, 49, 50, 51 73 or 75.

7. An isolated nucleic acid sequence consisting of SEQ ID NO: 2, 4, 6, 8, 10, 11, 12, 13, 14, 15, 16, 17, 18, 19, 20, 21, 22, 23, 24, 25, 26, 27, 28, 29, 42, 43, 44, 45, 46, 47, 48, 49, 50, 51 73 or 75.

8. An isolated nucleic acid sequence comprising a polynucleotide selected from the group consisting of:
   a) an isolated polynucleotide encoding a polypeptide of SEQ ID NO 3 or SEQ ID NO 3 with at least one conservative amino acid substitution;
   b) SEQ ID NO: 2;
   c) an isolated polynucleotide that has at least 70% sequence identity to SEQ ID NO: 2;

d) an isolated polynucleotide that has at least 80% sequence identity to SEQ ID NO: 2;

e) an isolated polynucleotide that has at least 90% sequence identity to SEQ ID NO: 2;

f) an isolated polynucleotide that has at least 95% sequence identity to SEQ ID NO: 2;

g) an isolated polynucleotide of at least 10 nucleic acids that hybridizes under stringent conditions to SEQ ID NO: 2;

h) an isolated polynucleotide complementary to a polynucleotide of (a), (b), (c), (d), (e), (f) or (g); and

i) an isolated polynucleotide that hybridizes under stringent conditions to SEQ ID NO: 2 and encodes a plant lecithin:cholesterol acyltransferase-like polypeptide.

9. An isolated nucleic acid sequence consisting essentially of a polynucleotide of the formula 5' X-$(R_1)_n$-$(R_2)_n$-$(R_3)_n$-Y 3', where X is hydrogen, Y is hydrogen or a metal, R_1 and R_3 are any nucleic acid, n is an integer between 0-3000, and R_2 is selected from the group consisting of:

a) an isolated polynucleotide encoding a polypeptide of SEQ ID NO: 3 or SEQ ID NO: 3 with at least one conservative amino acid substitution;

b) SEQ ID NO: 2;

c) an isolated polynucleotide that has at least 70% sequence identity to SEQ ID NO: 2;

d) an isolated polynucleotide that has at least 80% sequence identity to SEQ ID NO: 2;

e) an isolated polynucleotide that has at least 90% sequence identity to SEQ ID NO: 2;

f) an isolated polynucleotide that has at least 95% sequence identity to SEQ ID NO: 2;

g) an isolated polynucleotide of at least 10 nucleic acids that hybridizes under stringent conditions to SEQ ID NO: 2;

h) an isolated polynucleotide complementary to a polynucleotide of (a), (b), (c), (d), (e), (f) or (g); and

i) an isolated polynucleotide that hybridizes under stringent conditions to SEQ ID NO: 2 and encodes a plant lecithin:cholesterol acyltransferase-like polypeptide.

10. An isolated nucleic acid sequence comprising a polynucleotide selected from the group consisting of:

a) an isolated polynucleotide encoding a polypeptide of SEQ ID NO: 5 or SEQ ID NO: 5 with at least one conservative amino acid substitution;

b) SEQ ID NO: 4;

c) an isolated polynucleotide having at least 70% sequence identity with SEQ ID NO: 4;

d) an isolated polynucleotide having at least 80% sequence identity with SEQ ID NO: 4;

e) an isolated polynucleotide having at least 90% sequence identity with SEQ ID NO: 4;

f) an isolated polynucleotide having at least 95% sequence identity with SEQ ID NO: 4;

g) an isolated polynucleotide of at least 10 nucleic acids that hybridizes under stringent conditions to SEQ ID NO: 4;

h) an isolated polynucleotide complementary to a polynucleotide of (a), (b), (c), (d), (e), (f) or (g); and

i) an isolated polynucleotide that hybridizes under stringent conditions to SEQ ID NO: 4 and encodes a plant lecithin:cholesterol acyltransferase-like polypeptide.

11. An isolated nucleic acid sequence consisting essentially of a polynucleotide of the formula 5' $X-(R_1)_n-(R_2)_n-(R_3)_n-Y$ 3', where X is hydrogen, Y is hydrogen or a metal, R_1 and R_3 are any nucleic acid, n is an integer between 0-3000, and R_2 is selected from the group consisting of:

a) an isolated polynucleotide encoding a polypeptide of SEQ ID NO: 5 or SEQ ID NO: 5 with at least one conservative amino acid substitution;

b) SEQ ID NO: 4;

c) an isolated polynucleotide having at least 70% sequence identity with SEQ ID NO: 4;

d) an isolated polynucleotide having at least 80% sequence identity with SEQ ID NO: 4;

e) an isolated polynucleotide having at least 90% sequence identity with SEQ ID NO: 4;

f) an isolated polynucleotide having at least 95% sequence identity with SEQ ID NO: 4;

g) an isolated polynucleotide of at least 10 nucleic acids that hybridizes under stringent conditions to SEQ ID NO: 4;

h) an isolated polynucleotide complementary to a polynucleotide of (a), (b), (c), (d), (e), (f) or (g); and

i) an isolated polynucleotide that hybridizes under stringent conditions to SEQ ID NO: 4 and encodes a plant lecithin:cholesterol acyltransferase-like polypeptide.

12. An isolated nucleic acid sequence comprising a polynucleotide selected from the group consisting of:

a) an isolated polynucleotide encoding a polypeptide of SEQ ID NO: 7 or SEQ ID NO: 7 with at least one conservative amino acid substitution;

b) SEQ ID NO: 6;

c) an isolated polynucleotide having at least 70% sequence identity with SEQ ID NO: 6;

d) an isolated polynucleotide having at least 80% sequence identity with SEQ ID NO: 6;

e) an isolated polynucleotide having at least 90% sequence identity with SEQ ID NO: 6;

f) an isolated polynucleotide having at least 95% sequence identity with SEQ ID NO: 6;

g) an isolated polynucleotide of at least 10 nucleic acids that hybridizes under stringent conditions to SEQ ID NO: 6;

h) an isolated polynucleotide complementary to a polynucleotide of (a), (b), (c), (d), (e), (f) or (g); and

i) an isolated polynucleotide that hybridizes under stringent conditions to SEQ ID NO: 6 and encodes a plant lecithin:cholesterol acyltransferase-like polypeptide.

13. An isolated nucleic acid sequence consisting essentially of a polynucleotide of the formula 5' X-$(R_1)_n$-$(R_2)_n$-$(R_3)_n$-Y 3', where X is hydrogen, Y is hydrogen or a metal, R_1 and R_3 are any nucleic acid, n is an integer between 0-3000, and R_2 is selected from the group consisting of:

a) an isolated polynucleotide encoding a polypeptide of SEQ ID NO: 7 or SEQ ID NO: 7 with at least one conservative amino acid substitution;

b) SEQ ID NO: 6;

c) an isolated polynucleotide having at least 70% sequence identity with SEQ ID NO: 6;

d) an isolated polynucleotide having at least 80% sequence identity with SEQ ID NO:6;

e) an isolated polynucleotide having at least 90% sequence identity with SEQ ID NO: 6;

f) an isolated polynucleotide having at least 95% sequence identity with SEQ ID NO: 6;

g) an isolated polynucleotide of at least 10 nucleic acids that hybridizes under stringent conditions to SEQ ID NO: 6;

h) an isolated polynucleotide complementary to a polynucleotide of (a), (b), (c), (d), (e), (f) or (g); and

i) an isolated polynucleotide that hybridizes under stringent conditions to SEQ ID NO: 6 and encodes a plant lecithin:cholesterol acyltransferase-like polypeptide.

14. An isolated nucleic acid sequence comprising a polynucleotide selected from the group consisting of

a) an isolated polynucleotide encoding a polypeptide of SEQ ID NO: 9 or SEQ ID NO: 9 with at least one conservative amino acid substitution;

b) SEQ ID NO 8;

c) an isolated polynucleotide having at least 70% sequence identity with SEQ ID NO: 8;

d) an isolated polynucleotide having at least 80% sequence identity with SEQ ID NO: 8;

e) an isolated polynucleotide having at least 90% sequence identity with SEQ ID NO: 8;

f) an isolated polynucleotide having at least 95% sequence identity with SEQ ID NO: 8;

g) an isolated polynucleotide of at least 10 nucleic acids that hybridizes under stringent conditions to SEQ ID NO: 8;

h) an isolated polynucleotide complementary to a polynucleotide of (a), (b), (c), (d), (e), (f) or (g); and

i) an isolated polynucleotide that hybridizes under stringent conditions to SEQ ID NO: 8 and encodes a plant lecithin:cholesterol acyltransferase-like polypeptide.

15. An isolated nucleic acid sequence consisting essentially of a polynucleotide of the formula 5' $X-(R_1)_n-(R_2)_n-(R_3)_n-Y$ 3', where X is hydrogen, Y is hydrogen or a metal, R_1 and R_3 are any nucleic acid, n is an integer between 0-3000, and R_2 is selected from the group consisting of:

a) an isolated polynucleotide encoding a polypeptide of SEQ ID NO: 9 or SEQ ID NO: 9 with at least one conservative amino acid substitution;

b) SEQ ID NO: 8;

c) an isolated polynucleotide having at least 70% sequence identity with SEQ ID NO: 8;

d) an isolated polynucleotide having at least 80% sequence identity with SEQ ID NO: 8;

e) an isolated polynucleotide having at least 90% sequence identity with SEQ ID NO: 8;

f) an isolated polynucleotide having at least 95% sequence identity with SEQ ID NO: 8;

g) an isolated polynucleotide of at least 10 nucleic acids that hybridizes under stringent conditions to SEQ ID NO: 8;

h) an isolated polynucleotide complementary to a polynucleotide of (a), (b), (c), (d), (e), (f) or (g); and

i) an isolated polynucleotide that hybridizes under stringent conditions to SEQ ID NO: 8 and encodes a plant lecithin:cholesterol acyltransferase-like polypeptide.

16. An isolated nucleic acid sequence comprising a polynucleotide selected from the group consisting of:

a) an isolated polynucleotide encoding a polypeptide of SEQ ID NO: 74 or SEQ ID NO: 74 with at least one conservative amino acid substitution;

b) SEQ ID NO: 73;

c) an isolated polynucleotide having at least 70% sequence identity with SEQ ID NO: 73;

d) an isolated polynucleotide having at least 80% sequence identity with SEQ ID NO: 73;

e) an isolated polynucleotide having at least 90% sequence identity with SEQ ID NO: 73;

f) an isolated polynucleotide having at least 95% sequence identity with SEQ ID NO: 73;

g) an isolated polynucleotide of at least 10 nucleic acids that hybridizes under stringent conditions to SEQ ID NO: 73;

h) an isolated polynucleotide complementary to a polynucleotide of (a), (b), (c), (d), (e), (f) or (g); and

i) an isolated polynucleotide that hybridizes under stringent conditions to SEQ ID NO: 73 and encodes a plant lecithin:cholesterol acyltransferase-like polypeptide.

17. An isolated nucleic acid sequence consisting essentially of a polynucleotide of the formula 5' $X-(R_1)_n-(R_2)_n-(R_3)_n-Y$ 3', where X is hydrogen, Y is hydrogen or a metal, R_1 and R_3 are any nucleic acid, n is an integer between 0-3000, and R_2 is selected from the group consisting of:

a) an isolated polynucleotide encoding a polypeptide of SEQ ID NO: 74 or SEQ ID NO: 74 with at least one conservative amino acid substitution;

b) SEQ ID NO: 73;

c) an isolated polynucleotide having at least 70% sequence identity with SEQ ID NO: 73;

d) an isolated polynucleotide having at least 80% sequence identity with SEQ ID NO: 73;

e) an isolated polynucleotide having at least 90% sequence identity with SEQ ID NO: 73;

f) an isolated polynucleotide having at least 95% sequence identity with SEQ ID NO: 73;

g) an isolated polynucleotide of at least 10 nucleic acids that hybridizes under stringent conditions to SEQ ID NO: 73;

h) an isolated polynucleotide complementary to a polynucleotide of (a), (b), (c), (d), (e), (f) or (g); and

i) an isolated polynucleotide that hybridizes under stringent conditions to SEQ ID NO: 73 and encodes a plant lecithin:cholesterol acyltransferase-like polypeptide.

18. A isolated nucleic acid sequence comprising a polynucleotide selected from the group consisting of:

a) an isolated polynucleotide encoding a polypeptide of SEQ ID NO: 76 or SEQ ID NO: 76 with at least one conservative amino acid substitution;

b) SEQ ID NO: 75;

c) an isolated polynucleotide having at least 70% sequence identity with SEQ ID NO: 75;

d) an isolated polynucleotide having at least 80% sequence identity with SEQ ID NO: 75;

e) an isolated polynucleotide having at least 90% sequence identity with SEQ ID NO: 75;

f) an isolated polynucleotide having at least 95% sequence identity with SEQ ID NO: 75;

g) an isolated polynucleotide of at least 10 nucleic acids that hybridizes under stringent conditions to SEQ ID NO: 75;

h) an isolated polynucleotide complementary to a polynucleotide of (a), (b), (c), (d), (e), (f) or (g); and

i) an isolated polynucleotide that hybridizes under stringent conditions to SEQ ID NO: 75 and encodes a plant lecithin:cholesterol acyltransferase-like polypeptide.

19. An isolated nucleic acid sequence consisting essentially of a polynucleotide of the formula 5' X-$(R_1)_n$-$(R_2)_n$-$(R_3)_n$-Y 3', where X is hydrogen, Y is hydrogen or a metal, R_1 and R_3 are any nucleic acid, n is an integer between 0-3000, and R_2 is selected from the group consisting of:

a) an isolated polynucleotide encoding a polypeptide of SEQ ID NO: 76 or SEQ ID NO: 76 with at least one conservative amino acid substitution;

b) SEQ ID NO: 75;

c) an isolated polynucleotide having at least 70% sequence identity with SEQ ID NO: 75;

d) an isolated polynucleotide having at least 80% sequence identity with SEQ ID NO: 75;

e) an isolated polynucleotide having at least 90% sequence identity with SEQ ID NO: 75;

f) an isolated polynucleotide having at least 95% sequence identity with SEQ ID NO: 75;

g) an isolated polynucleotide of at least 10 nucleic acids that hybridizes under stringent conditions to SEQ ID NO: 75;

h) an isolated polynucleotide complementary to a polynucleotide of (a), (b), (c), (d), (e), (f) or (g); and

i) an isolated polynucleotide that hybridizes under stringent conditions to SEQ ID NO: 75 and encodes a plant lecithin:cholesterol acyltransferase-like polypeptide.

20. An isolated nucleic acid sequence comprising a polynucleotide selected from the group consisting of:

a) SEQ ID NO: 42 or a degenerate variant thereof;

b) an isolated polynucleotide having at least 70% sequence identity with SEQ ID NO: 42;

c) an isolated polynucleotide having at least 80% sequence identity with SEQ ID NO: 42;

d) an isolated polynucleotide having at least 90% sequence identity with SEQ ID NO: 42;

e) an isolated polynucleotide having at least 95% sequence identity with SEQ ID NO: 42;

f) an isolated polynucleotide of at least 10 nucleic acids that hybridizes under stringent conditions to SEQ ID NO: 42;

g) an isolated polynucleotide complementary to a polynucleotide of (a), (b), (c), (d), (e), or (f); and

h) an isolated polynucleotide that hybridizes under stringent conditions to SEQ ID NO: 42 and encodes an acyl CoA:cholesterol acyltransferase-like polypeptide.

21. An isolated nucleic acid sequence consisting essentially of a polynucleotide of the formula 5' X-$(R_1)_n$-$(R_2)_n$-$(R_3)_n$-Y 3', where X is hydrogen, Y is hydrogen or a metal R_1 and R_3 are any nucleic acid, n is an integer between 0 and 3000, and R_2 is selected from the group consisting of:

a) SEQ ID NO: 42 or degenerate variants thereof;

b) an isolated polynucleotide having at least 70% sequence identity to SEQ ID NO: 42;

c) an isolated polynucleotide having at least 80% sequence identity to SEQ ID NO: 42;

d) an isolated polynucleotide having at least 90% sequence identity to SEQ ID NO: 42;

e) an isolated polynucleotide having at least 95% sequence identity to SEQ ID NO: 42;

f) an isolated polynucleotide of at least 10 nucleic acids that hybridizes under stringent conditions to SEQ ID NO: 42;

g) an isolated polynucleotide complementary to a polynucleotide of (a), (b), (c), (d), (e), or (f); and

h) an isolated polynucleotide that hybridizes under stringent conditions to SEQ ID NO: 42 and encodes an acyl CoA:cholesterol acyltransferase-like polypeptide.

22. A recombinant nucleic acid construct comprising a regulatory sequence operably linked to polynucleotide encoding a lecithin:cholesterol acyltransferase-like polypeptide or a fragment thereof.

23. The recombinant nucleic acid construct of claim 22, wherein said lecithin:cholesterol acyltransferase-like polypeptide is a plant lecithin:cholesterol acyltransferase-like polypeptide.

24. A recombinant nucleic acid construct comprising a regulatory sequence operably linked to a polynucleotide encoding an acyl CoA:cholesterol acyltransferase-like polypeptide.

25. The recombinant nucleic acid construct of claim 24, wherein said acyl CoA:cholesterol acyltransferase-like polypeptide is a plant acyl CoA:cholesterol acyltransferase-like polypeptide.

26. The recombinant construct of claim 22, wherein said regulatory sequence comprises a heterologous regulatory sequence.

27. The recombinant construct of claim 24, wherein said regulatory sequence comprises a heterologous regulatory sequence.

28. The recombinant construct of claim 22, wherein said regulatory sequence is functional in a plant cell.

29. The recombinant construct of claim 24, wherein said regulatory sequence is functional in a plant cell.

30. The recombinant construct of claim 22, further comprising a termination sequence.

31. The recombinant construct of claim 24 further comprising a termination sequence.

32. The recombinant construct of claim 22 wherein said polynucleotide is selected from the group consisting of SEQ ID NO: 2, 4, 6, 8, 10, 11, 12, 13, 14, 15, 16, 17, 18, 19, 20, 21, 22, 23, 24, 25, 26, 27, 28, 29, 43, 44, 45, 46, 47, 48, 49, 50, 51, 73 and 75.

33. The recombinant construct of claim 24, wherein said polynucleotide is selected from the group consisting of SEQ ID NO: 33 and 42.

34. The recombinant construct of claim 22, wherein said regulatory sequence comprises a constitutive promoter.

35. The recombinant construct of claim 24, wherein said regulatory sequence comprises a constitutive promoter.

36. The recombinant construct of claim 22, wherein said regulatory sequence comprises an inducible promoter.

37. The recombinant construct of claim 24, wherein said regulatory sequence comprises an inducible promoter.

38. The recombinant construct of claim 22, wherein said regulatory sequence is selected from the group consisting of a tissue specific promoter, a developmentally regulated promoter, an organelle specific promoter, and a seed specific promoter.

39. The recombinant construct of claim 24, wherein said regulatory sequence is selected from the group consisting of a tissue specific promoter, a developmentally regulated promoter, an organelle specific promoter, and a seed specific promoter.

40. A host cell containing the recombinant construct of claim 22 or 24.

41. The host cell of claim 40, wherein said host cell is selected from the group consisting of plant cells, animal cells, insect cells, yeast, bacteria, bacteriophage and viruses.

42. The host cell of claim 40, wherein said host cell is a plant cell.

43. The host cell of claim 40, wherein said host cell expresses a lecithin:cholesterol acyltransferase-like polypeptide or an acyl CoA:cholesterol acyltransferase-like polypeptide.

44. The host cell of claim 43, wherein said cholesterol acyltransferase-like polypeptide is a plant acyltransferase-like polypeptide.

45. A plant comprising at least one host cell of claim 40.

46. The progeny of a plant of claim 45.

47. A seed from the plant of claim 45.

48. A plant comprising the recombinant construct of claim 22 or 24.

49. The progeny of a plant of claim 48.

50. A seed from the plant of claim 48.

51. A purified polypeptide comprising an amino acid sequence selected from the group consisting of SEQ ID NO: 3, SEQ ID NO: 3 with at least one conservative amino acid substitution, SEQ ID NO: 5, SEQ ID NO: 5 with at least one conservative amino acid substitution, SEQ ID NO: 7, SEQ ID NO: 7 with at least one conservative amino acid substitution, SEQ ID NO: 9, SEQ ID NO: 9 with at least one conservative amino acid substitution, SEQ ID NO: 74, SEQ ID NO: 74 with at least one conservative amino acid substitution, SEQ ID NO: 76 and SEQ ID NO: 76 with at least one conservative amino acid substitution.

52. A purified immunogenic polypeptide comprising at least 10 consecutive amino acids from an amino acid sequence selected from the group consisting of SEQ ID NO: 3, SEQ ID NO: 3 with at least one conservative amino acid substitution, SEQ ID NO: 5, SEQ ID NO: 5 with at least one conservative amino acid substitution, SEQ ID NO: 7, SEQ ID NO: 7 with at least one conservative amino acid substitution, SEQ ID NO: 9, SEQ ID NO: 9 with at least one conservative amino acid substitution, SEQ ID NO: 74, SEQ ID NO: 74 with at least one conservative amino acid substitution, SEQ ID NO: 76 and SEQ ID NO: 76 with at least one conservative amino acid substitution.

53. An antibody which specifically binds to an immunogenic polypeptide of claim 52.

54. A method for producing a lecithin:cholesterol acyltransferase-like polypeptide or an acyl CoA:cholesterol acyltransferase-like polypeptide comprising culturing a host cell of claim 40 under conditions permitting expression of said lecithin:cholesterol acyltransferase-like polypeptide or acyl CoA:cholesterol acyltransferase-like polypeptide.

55. The method of claim 54, further comprising isolating the cholesterol acyltransferase-like polypeptide from the host cell or from the medium in which the host cell is cultured.

56. A method for modifying the sterol content of a host cell, comprising transforming a host cell with a recombinant construct containing a regulatory sequence operably linked to a polynucleotide encoding a lecithin:cholesterol acyltransferase-like polypeptide and culturing said host cell under conditions wherein said host cell expresses a lecithin:cholesterol acyltransferase-like polypeptide such that said host cell has a modified sterol composition as compared to host cells without the recombinant construct.

57. The method of claim 56, wherein said lecithin:cholesterol acyltransferase-like polypeptide is a plant lecithin:cholesterol acyltransferase-like polypeptide.

58. A method for modifying the sterol content of a host cell, comprising transforming a host cell with a recombinant construct containing a regulatory sequence operably linked to a polynucleotide encoding an acyl CoA:cholesterol acyltransferase-like polypeptide and culturing said host cell under conditions wherein said host cell expresses an acyl CoA:cholesterol acyltransferase-like polypeptide such that said host cell has a modified sterol composition as compared to host cells without the recombinant construct.

59. The method of claim 58, wherein said acyl CoA:cholesterol acyltransferase-like polypeptide is a plant acyl CoA:cholesterol acyltransferase-like polypeptide.

60. The method of claim 56, wherein said modified sterol composition is an increase in sterol esters.

61. The method of claim 58, wherein said modified sterol composition is an increase in sterol esters.

62. The method of claim 56, wherein said polynucleotide encoding a lecithin:cholesterol acyltransferase-like polypeptide is selected from the group consisting of SEQ ID NO: 2, 4, 6, 8, 10, 11, 73 and 75.

63. The method of claim 58 wherein said polynucleotide encoding a acyl CoA:cholesterol acyltransferase-like polypeptide is SEQ ID NO 33 OR 42.

64. The method of claim 56, wherein said regulatory sequence comprises a constitutive promoter.

65. The method of claim 58, wherein said regulatory sequence comprises a constitutive promoter.

66. The method of claim 56, wherein said regulatory sequence is an inducible promoter.

67. The method of claim 58, wherein said regulatory sequence is an inducible promoter.

68. The method of claim 56, wherein said regulatory sequence is a tissue specific promoter.

69. The method of claim 58, wherein said regulatory sequence is a tissue specific promoter.

70. The method of claim 56, wherein said regulatory sequence is a seed specific promoter.

71. The method of claim 58, wherein said regulatory sequence is a seed specific promoter.

72. The method of claim 56, wherein said polynucleotide encoding a lecithin:cholesterol acyltransferase-like polypeptide is in the antisense orientation.

73. The method of claim 58, wherein said polynucleotide encoding an acyl CoA:cholesterol acyltransferase-like polypeptide is in the antisense orientation.

74. The method of claim 72, wherein said modified sterol composition is a decrease in sterol esters.

75. The method of claim 73, wherein said modified sterol composition is a decrease in sterol esters.

76. A plant comprising a recombinant construct containing a regulatory sequence operably linked to a polynucleotide encoding a lecithin:cholesterol acyltransferase-like polypeptide wherein expression of said recombinant construct results in modified sterol composition of said plant as compared to the same plant without said recombinant construct.

77. The plant of claim 76, wherein said lecithin:cholesterol acyltransferase-like polypeptide is a plant lecithin:cholesterol acyltransferase-like polypeptide.

78. The plant of claim 76, wherein said polynucleotide encoding a lecithin:cholesterol acyltransferase-like polypeptide is selected from the group consisting of SEQ ID NO: 2, 4, 6, 8, 10, 11, 73 and 75.

79. A plant comprising a recombinant construct containing a regulatory sequence operably linked to a polynucleotide encoding an acyl CoA:cholesterol acyltransferase-like polypeptide wherein expression of said recombinant construct results in modified sterol composition of said plant as compared to the same plant without said recombinant construct.

80. The plant of claim 79, wherein said acyl CoA:cholesterol acyltransferase-like polypeptide is a plant acyl CoA:cholesterol acyltransferase-like polypeptide.

81. The plant of claim 79, wherein said polynucleotide encoding an acyl CoA:cholesterol acyltransferase-like polypeptide is SEQ ID NO: 33 or 42.

82. The plant of claim 76, wherein said regulatory sequence comprises a tissue specific promoter.

83. The plant of claim 79, wherein said regulatory sequence comprises a tissue specific promoter.

84. The plant of claim 76, wherein said regulatory sequence comprises a seed specific promoter.

85. The plant of claim 79, wherein said regulatory sequence comprises a seed specific promoter.

86. The plant of claim 76, wherein said modified sterol composition is an increase in sterol esters.

87. The plant of claim 79, wherein said modified sterol composition is an increase in sterol esters.

88. The plant of claim 76, wherein the polynucleotide encoding a lecithin:cholesterol acyltransferase-like polypeptide is in the antisense orientation.

89. The plant of claim 79, wherein the polynucleotide encoding an acyl CoA:cholesterol acyltransferase-like polypeptide is in the antisense orientation.

90. An oil obtained from the plant of claim 76 or 79.

91. A method for producing an oil with a modified sterol composition comprising, providing a plant of claim 76 or 79 and extracting the oil from said plant.

92. An oil produced by the method of claim 91.

93. A method for altering oil production by a host cell comprising, transforming a host cell with a recombinant construct containing a regulatory sequence operably linked to a polynucleotide encoding a lecithin:cholesterol acyltransferase-like polypeptide and culturing said host cell under conditions wherein said host cell expresses a lecithin:cholesterol acyltransferase-like polypeptide such that said host cell has an altered oil production as compared to host cells without the recombinant construct.

94. The method of claim 93, wherein said lecithin:cholesterol acyltransferase-like polypeptide is a plant lecithin:cholesterol acyltransferase-like polypeptide.

95. A method for altering oil production by a host cell comprising, transforming a host cell with a recombinant construct containing a regulatory sequence operably linked to a polynucleotide encoding an acyl CoA:cholesterol acyltransferase-like polypeptide and culturing said host cell under conditions wherein said host cell expresses an acyl CoA:cholesterol acyltransferase-like polypeptide such that said host cell has an altered oil production as compared to host cells without the recombinant construct.

96. The method of claim 95, wherein said acyl CoA:cholesterol acyltransferase-like polypeptide is a plant acyl CoA:cholesterol acyltransferase-like polypeptide.

97. The method of claim 93, wherein said oil production is increased.

98. The method of claim 95, wherein said oil production is increased.

99. The method of claim 93, wherein said host cell is a plant cell.

100. The method of claim 95, wherein said host cell is a plant cell.

101. The method of claim 93, wherein said polynucleotide encoding a lecithin:cholesterol acyltransferase-like polypeptide is selected from the group consisting of SEQ ID NO: 2, 4, 6, 8, 10, 11, 73 and 75.

102. The method of claim 95, wherein said polynucleotide encoding an acyl CoA:cholesterol acyltransferase-like polypeptide is SEQ ID NO: 33 or 42.

103. The method of claim 93, wherein said regulatory sequence is a tissue specific promoter.

104. The method of claim 95, wherein said regulatory sequence is a tissue specific promoter.

105. The method of claim 93, wherein said regulatory sequence is a seed specific promoter.

106. The method of claim 95, wherein said regulatory sequence is a seed specific promoter.

107. A plant comprising a recombinant construct containing a regulatory sequence operably linked to a polynucleotide encoding a lecithin:cholesterol acyltransferase-like polypeptide wherein expression of said recombinant construct results in an altered production of oil by said plant as compared to the same plant without said recombinant construct.

108. The plant of claim 107, wherein said lecithin:cholesterol acyltransferase-like polypeptide is a plant lecithin:cholesterol acyltransferase-like polypeptide.

109. A plant comprising a recombinant construct containing a regulatory sequence operably linked to a polynucleotide encoding an acyl CoA:cholesterol acyltransferase-like polypeptide wherein expression of said recombinant construct results in an altered production of oil by said plant as compared to the same plant without said recombinant construct.

110. The plant of claim 109, wherein said acyl CoA:cholesterol acyltransferase-like polypeptide is a plant acyl CoA:cholesterol acyltransferase-like polypeptide.

111. The plant of claim 107, wherein said oil production is increased.

112. The plant of claim 109, wherein said oil production is increased.

113. The plant of claim 107, wherein said polynucleotide encoding a lecithin:cholesterol acyltransferase-like polypeptide is selected from the group consisting of SEQ ID NO: 2, 4, 6, 8, 10, 11, 73 and 75.

114. The plant of claim 109, wherein said polynucleotide encoding an acyl CoA:cholesterol acyltransferase-like polypeptide is SEQ ID NO: 33 or 42.

115. The plant of claim 107, wherein said regulatory sequence is a tissue specific promoter.

116. The plant of claim 109, wherein said regulatory sequence is a tissue specific promoter.

117. The plant of claim 107, wherein said regulatory sequence is a seed specific promoter.

118. The plant of claim 109, wherein said regulatory sequence is a seed specific promoter.

119. A food, food ingredient or food product comprising the oil of claim 90 or 92.

120. A food, food ingredient or food product comprising the plant of claim 107 or 109.

ABSTRACT

The present invention is directed to lecithin:cholesterol acyltransferase-like polypeptides (LCAT) and acyl CoA:cholesterol acyltransferases-like polypeptides (ACAT). The invention provides polynucleotides encoding such cholesterol:acyltransferases-like polypeptides, polypeptides encoded by such polynucleotides, and the use of such polynucleotides to alter sterol composition and oil production in plants and host cells. Also provided are oils produced by the plants and host cells containing the polynucleotides and food products, nutritional supplements, and pharmaceutical composition containing plants or oils of the present invention. The polynucleotides of the present invention include those derived from plant sources

FIG. 1A

ClustalW Formatted Alignments

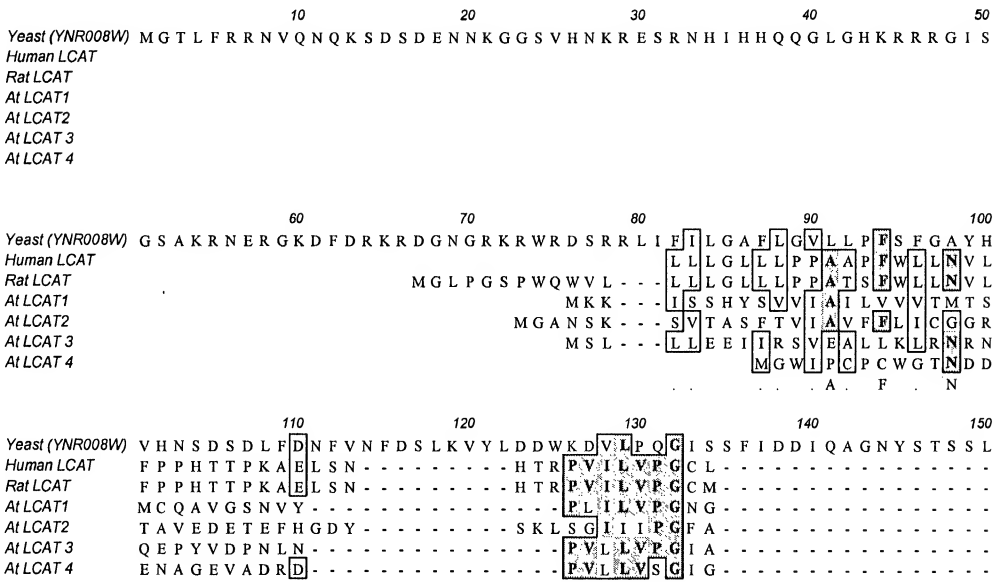


FIG. 1B

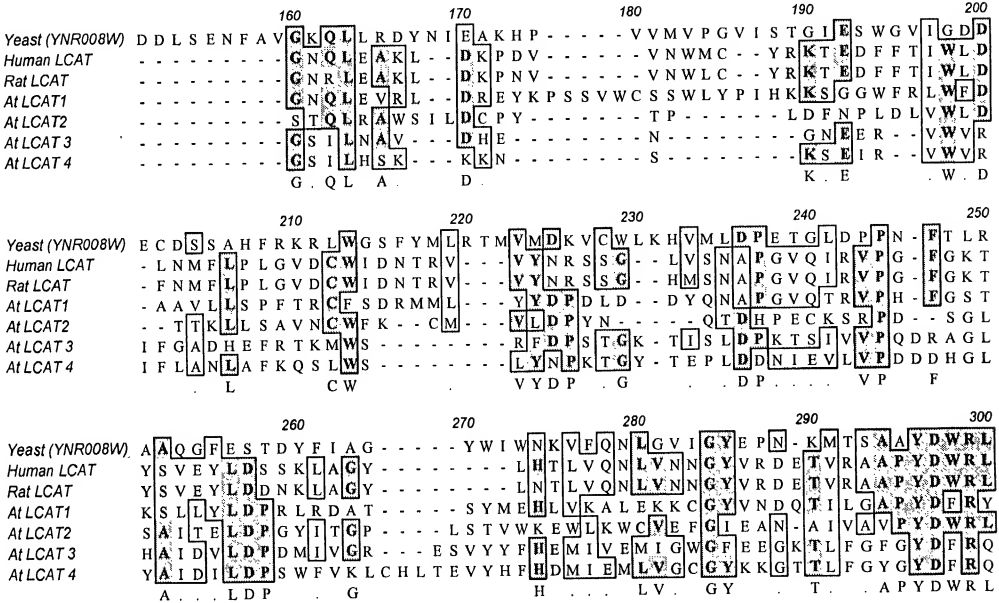


FIG. 1C

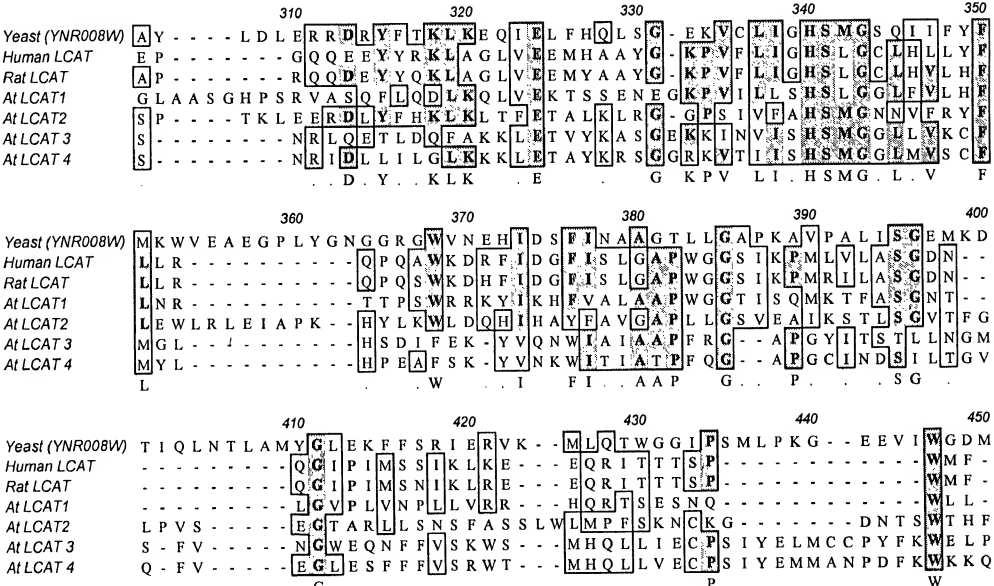
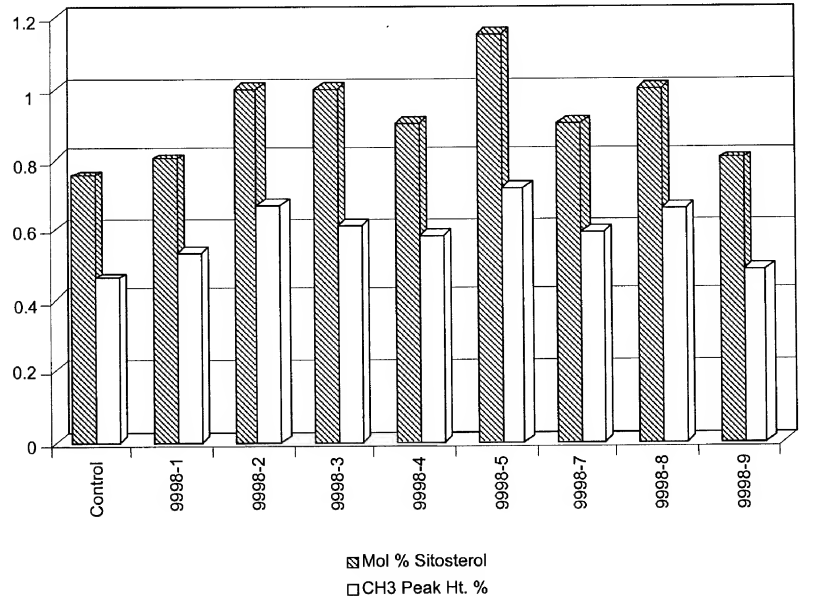


FIG. 2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0.2
0
Control
9998-1
9998-2
9998-3
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9998-5
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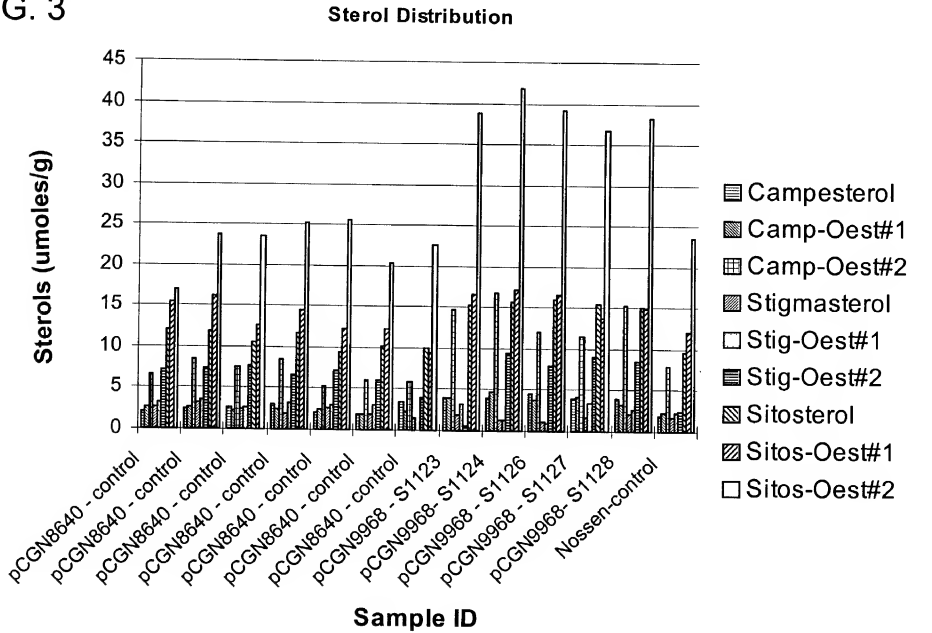
9998-9
Mol % Sitosterol
CH3 Peak Ht. %

FIG. 3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Sterols (umoles/g)
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40
35
30
25
20
15
10
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pCGN8640 - control
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Sample ID
Campesterol
Camp-Oest#1
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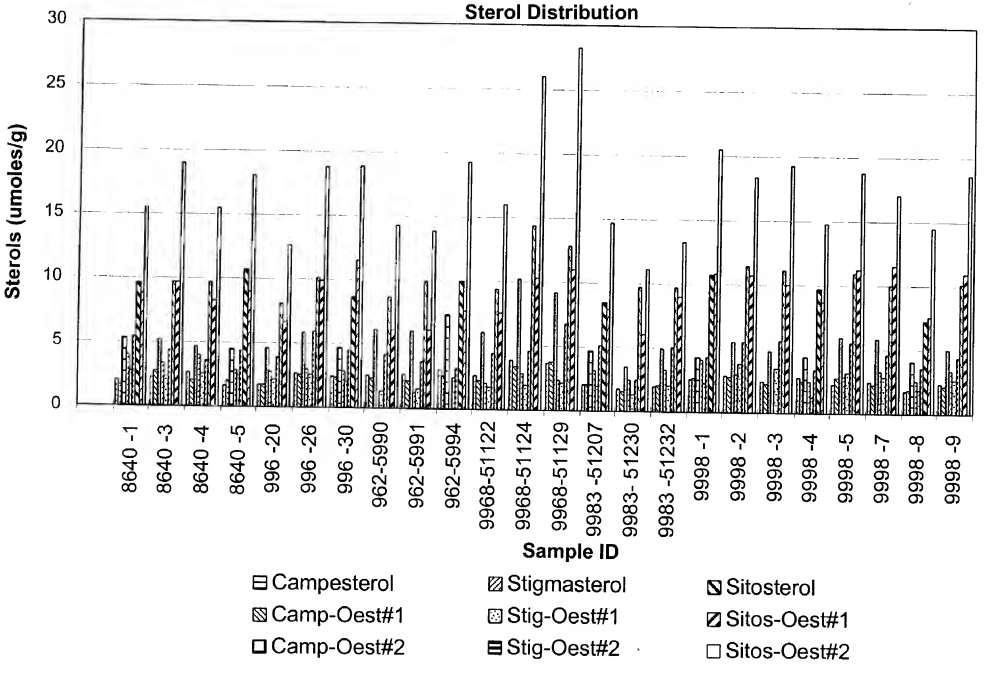
FIG. 4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25
20
15
10
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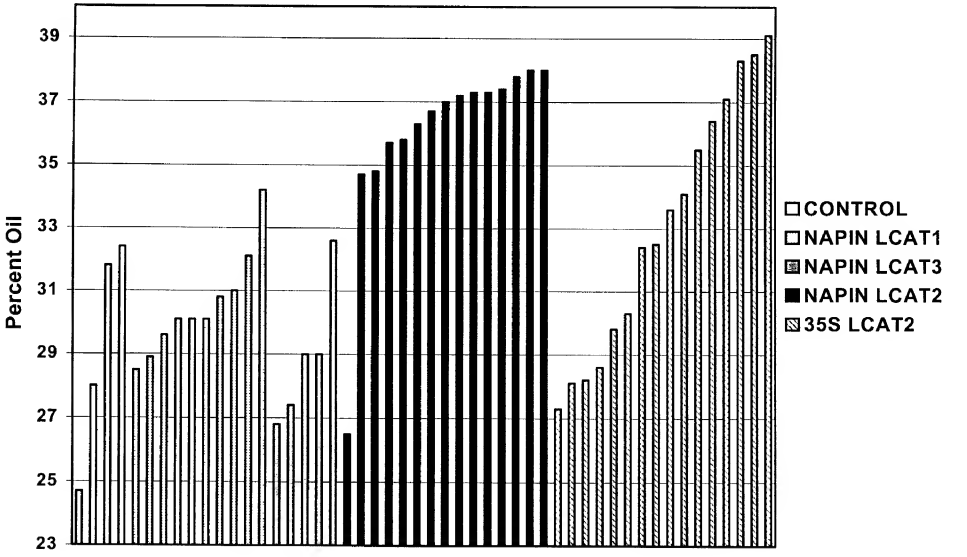
FIG. 5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Percent Oil
39
37
35
33
31
29
27
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23
CONTROL
NAPIN LCAT1
NAPIN LCAT3
NAPIN LCAT2
35S LCAT2

Attorney's Docket No. MTC 6718

DECLARATION AND POWER OF ATTORNEY

REGULAR OR DESIGN APPLICATION

As a below named inventor, I hereby declare that:

My residence, post office address and citizenship are as stated below next to my name.

I believe I am the original, first and sole inventor (if only one name is listed below) or an original, first and joint inventor (if plural names are listed below) of the subject matter which is claimed and for which a patent is sought on the invention entitled:

PLANT STEROL ACYLTRANSFERASES

the specification of which:

(check one)

[ ] is attached hereto
[ ] was filed on _____ as Application Serial No. _____, and was amended on _____.
[ ] was described and claimed in PCT International Application No. _____, filed on _____ and as amended under PCT Article 19 on _____, if any.

ACKNOWLEDGEMENT OF REVIEW OF PAPERS AND DUTY OF CANDOR

I hereby state that I have reviewed and understand the contents of the above identified specification, including the claims, as amended by any amendment referred to above.

I acknowledge the duty to disclose information which is material to patentability as defined in Title 37, Code of Federal Regulations §1.56.

PRIORITY CLAIM

I hereby claim foreign priority benefits under Title 35, United States Code, §119 (a) - (d) or §365(b) of any foreign application for patent or inventor's certificate, or §365(a) of any PCT application which designates at least one country other than the United States of America, listed below and have also identified below any foreign application for patent or inventor's certificate having a filing date before that of the application on which priority is claimed:

Priority Claimed

(Number)	(Country)	(Day/Month/Year Filed)
(Number)	(Country)	(Day/Month/Year Filed)
(Number)	(Country)	(Day/Month/Year Filed)

Priority Not Claimed

ANY FOREIGN APPLICATION(S), ON THE SAME SUBJECT MATTER WHICH HAS A FILING DATE EARLIER THAN THE EARLIEST APPLICATION FROM WHICH PRIORITY IS CLAIMED

(Number)	(Country)	(Day/Month/Year Filed)

CLAIM FOR BENEFIT OF PROVISIONAL APPLICATION(S)

I hereby claim the benefit under Title 35, United States Code, §119(e) of any United States provisional application(s) listed below.

60/152,493 (Application Number)	August 30, 1999 (Filing Date)
(Application Number)	(Filing Date)

CLAIM FOR BENEFIT OF EARLIER U.S. APPLICATION(S) UNDER 35 U.S.C. 120

(complete this part only if this is a divisional, continuation or CIP application)

I hereby claim the benefit under Title 35, United States Code, §120 of any United States application(s), or §365(c) of any PCT international application designating the United States of America, listed below and, insofar as the subject matter of each of the claims of this application is not disclosed in the prior United States application in the manner provided by the first paragraph of Title 35, United States Code §112, I acknowledge the duty to disclose information which is material to patentability as defined in Title 37, Code of Federal Regulations, §1.56 which became available between the filing date of the prior application and the national or PCT International filing date of this application:

(Serial No.)	(Filing Date)	(Status)
(Serial No.)	(Filing Date)	(Status)

POWER OF ATTORNEY

I hereby appoint the following attorneys to prosecute this application and to transact all business in the Patent and Trademark Office connected therewith: Irving Powers (15,700), Donald G. Leavitt (17,626), John K. Roedel, Jr. (25,914), Michael E. Godar (28,416), Edward J. Hejlek (31,525), William E. Lahey (26,757), Richard G. Heywood (18,224), Frank R. Agovino (27,416), Kurt F. James (33,716), G. Harley Blosser (33,650), Paul I. J. Fleischut (35,513), Vincent M. Keil (36,838), Robert M. Evans, Jr. (36,794), Robert M. Bain (36,736), Joseph A. Schaper (30,493), Kathleen M. Petrillo (35,076), David E. Crawford, Jr. (38,118), Paul A. Maddock (37,877), Richard L. Bridge (40,529), Christopher M. Goff (41,785), James E. Butler (40,931), Derick E. Allen (43,468), Matthew L. Cutler (43,574), Michael G. Munsell (43,820), Karen Y. Hui (44,785), Anthony R. Kinney (44,834), Brian P. Klein (44,837), Sarah J. Chickos (46,157), Donald W. Tuegel (45,424), Steven M. Ritchey (46,321), Michael J. Thomas (39,857), Kathryn J. Doty (40,593), and Laura R. Polcyn (P-47,000), all of the law firm of SENNIGER, POWERS, LEAVITT & ROEDEL, One Metropolitan Square, 16th Floor, St. Louis, Missouri 63102.

Send Correspondence To:	Direct Telephone Calls To:
Customer Number: 000321	James E. Butler, Ph.D. (314) 231-5400

I hereby declare that all statements made herein of my own knowledge are true and that all statements made on information and belief are believed to be true; and further that these statements were made with the knowledge that willful false statements and the like so made are punishable by fine or imprisonment, or both, under Section 1001 of Title 18 of the United States Code and that such willful false statements may jeopardize the validity of the application or any patent issued thereon.

- -

Full name of sole or first inventor Michael Lassner

Inventor's signature _____ Date _____

Residence Redwood City, CA Citizenship U.S.A.

Post Office address 515 Galveston Drive

Redwood City, California 94063

Full name of second joint inventor Alison Van Eenennaam

Second inventor's signature _____ Date _____

Residence Davis, California Citizenship Australia

Post Office address 856 Burr Street

Davis, California 95616

SEQUENCE LISTING

<110> Monsanto Company

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<151> 1999-08-30

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<213> Arabidopsis thaliana

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Pro Val Leu Leu Val Pro Gly Ile Ala Gly Ser Ile Leu Asn Ala Val
35 40 45

Asp His Glu Asn Gly Asn Glu Glu Arg Val Trp Val Arg Ile Phe Gly
50 55 60

Ala Asp His Glu Phe Arg Thr Lys Met Trp Ser Arg Phe Asp Pro Ser

65 70 75 80

Thr Gly Lys Thr Ile Ser Leu Asp Pro Lys Thr Ser Ile Val Val Pro
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Gln Asp Arg Ala Gly Leu His Ala Ile Asp Val Leu Asp Pro Asp Met
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Ile Val Gly Arg Glu Ser Val Tyr Tyr Phe His Glu Met Ile Val Glu
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130 135 140

Tyr Asp Phe Arg Gln Ser Asn Arg Leu Gln Glu Thr Leu Asp Gln Phe
145 150 155 160

Ala Lys Lys Leu Glu Thr Val Tyr Lys Ala Ser Gly Glu Lys Lys Ile
165 170 175

Asn Val Ile Ser His Ser Met Gly Gly Leu Leu Val Lys Cys Phe Met
180 185 190

Gly Leu His Ser Asp Ile Phe Glu Lys Tyr Val Gln Asn Trp Ile Ala
195 200 205

Ile Ala Ala Pro Phe Arg Gly Ala Pro Gly Tyr Ile Thr Ser Thr Leu
210 215 220

Leu Asn Gly Met Ser Phe Val Asn Gly Trp Glu Gln Asn Phe Phe Val
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260 265 270

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275 280 285

Val Val Leu Glu Ser Tyr Arg Ser Leu Glu Ser Leu Glu Val Phe Thr
290 295 300

Lys Ser Leu Ser Asn Asn Thr Ala Asp Tyr Cys Gly Glu Ser Ile Asp
305 310 315 320

Leu Pro Phe Asn Trp Lys Ile Met Glu Trp Ala His Lys Thr Lys Gln

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Val Leu Ala Ser Ala Lys Leu Pro Pro Lys Val Lys Phe Tyr Asn Ile
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Lys Met Pro Val Lys Asp Leu Thr Asn Leu Arg Tyr Phe Gln Pro Thr
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Tyr Ile Cys Val Asp Gly Asp Gly Thr Val Pro Met Glu Ser Ala Met
385                 390                 395                 400

Ala Asp Gly Leu Glu Ala Val Ala Arg Val Gly Val Pro Gly Glu His
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Arg Gly Ile Leu Asn Asp His Arg Val Phe Arg Met Leu Lys Lys Trp
            420                 425                 430

Leu Asn Val Gly Glu Pro Asp Pro Phe Tyr Asn Pro Val Asn Asp Tyr
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Val Ile Leu Pro Thr Thr Tyr Glu Phe Glu Lys Phe His Glu Asn Gly
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Leu Glu Val Ala Ser Val Lys Glu Ser Trp Asp Ile Ile Ser Asp Asp
465                 470                 475                 480

Asn Asn Ile Gly Thr Thr Gly Ser Thr Val Asn Ser Ile Ser Val Ser
                485                 490                 495

Gln Pro Gly Asp Asp Gln Asn Pro Gln Ala Glu Ala Arg Ala Thr Leu
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Gly Gly Ser Ile Leu His Ser Lys Lys Lys Asn Ser Lys Ser Glu Ile
         35   40   45

Arg Val Trp Val Arg Ile Phe Leu Ala Asn Leu Ala Phe Lys Gln Ser
     50   55   60

Leu Trp Ser Leu Tyr Asn Pro Lys Thr Gly Tyr Thr Glu Pro Leu Asp
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Asp Asn Ile Glu Val Leu Val Pro Asp Asp Asp His Gly Leu Tyr Ala
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Ile Asp Ile Leu Asp Pro Ser Trp Phe Val Lys Leu Cys His Leu Thr
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Tyr Lys Lys Gly Thr Thr Leu Phe Gly Tyr Gly Tyr Asp Phe Arg Gln
    130                 135                 140

Ser Asn Arg Ile Asp Leu Leu Ile Leu Gly Leu Lys Lys Lys Leu Glu
145                 150                 155                 160

Thr Ala Tyr Lys Arg Ser Gly Gly Arg Lys Val Thr Ile Ile Ser His
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Ser Met Gly Gly Leu Met Val Ser Cys Phe Met Tyr Leu His Pro Glu
            180                 185                 190

Ala Phe Ser Lys Tyr Val Asn Lys Trp Ile Thr Ile Ala Thr Pro Phe
        195                 200                 205

Gln Gly Ala Pro Gly Cys Ile Asn Asp Ser Ile Leu Thr Gly Val Gln
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Phe Val Glu Gly Leu Glu Ser Phe Phe Phe Val Ser Arg Trp Thr Met
225                 230                 235                 240

His Gln Leu Leu Val Glu Cys Pro Ser Ile Tyr Glu Met Met Ala Asn
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Lys Ser Glu Asn Asp Val Asp Thr Ser Val Glu Leu Glu Ser Phe Gly
        275                 280                 285

Leu Ile Glu Ser Ile Asp Leu Phe Asn Asp Ala Leu Lys Asn Asn Glu
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Leu Ser Tyr Gly Gly Asn Lys Ile Ala Leu Pro Phe Asn Phe Ala Ile
305                 310                 315                 320

Leu Asp Trp Ala Ala Lys Thr Arg Glu Ile Leu Asn Lys Ala Gln Leu
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Pro Asp Gly Val Ser Phe Tyr Asn Ile Tyr Gly Val Ser Leu Asn Thr
            340                 345                 350
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Pro Phe Asp Val Cys Tyr Gly Thr Glu Thr Ser Pro Ile Asp Asp Leu
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370 375 380

Gly Thr Val Pro Ala Glu Ser Ala Ala Ala Ala Gln Phe Lys Ala Val
385 390 395 400

Ala Ser Val Gly Val Ser Gly Ser His Arg Gly Leu Leu Arg Asp Glu
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<220>
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<210> 15
<211> 272
<212> DNA
<213> Glycine max

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<212> DNA
<213> Glycine max

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<222> (1)..(237)
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<211> 244
<212> DNA
<213> Glycine max

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<222> (1)..(244)
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<210> 18
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<212> DNA

<213> Glycine max

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<210> 20
<211> 1155
<212> DNA
<213> Zea mays

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<210> 21
<211> 328
<212> DNA
<213> Zea mays

<400> 21
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aaggcctgat agtggtcttc tgcaattaca gagctggacc ctggttatat aacaggtcct 180
ctctcttcag tatggaaaga atgggtcaaa tggtgtgtag agtttggcat tgaagctaat 240
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ctgtactttc acaattaaac aggatcag 328

<210> 22
<211> 356
<212> DNA
<213> Zea mays

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atcgctgttc cgtatgattg gagactgccc ccatcaatgc ttgaggagag agatctgtac 180
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catctgttag gatcagtaag agttaatggc ccatgacgga ttcaggttcc tgctcaccaa 300
cagatcccac aagcatacgg ttaccgccaa tgcctgcagt tggacagtac caaccc 356

<210> 23
<211> 1552
<212> DNA
<213> Zea mays

<400> 23
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<210> 24
<211> 227
<212> DNA
<213> Zea mays

<220>
<221> unsure
<222> (1)..(227)
<223> n=unknown

<400> 24
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tgggggggatt acttgtgaaa tgtntcatct cactgcacag tgatatatnt gaaaaatatg 180
tcaagagttg gntcgcaatt gcngcaccat tccaaggtgc ccctggg 227

<210> 25
<211> 1587
<212> DNA
<213> Zea mays

<220>
<221> unsure
<222> (1)..(1587)

<223> n=unknown

<400> 25
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gacgaccacg gcctgtttgc catcgacatt cttgatcctt cctggtttgt agaacttctc 180
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gagaaaggga ccacactatt tggatatggt tatgattttc gccaaagcaa caggatagac 300
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aaaaaaaaaa aaaaaaaaaa aaaaaaa 1587

<210> 26
<211> 300
<212> DNA
<213> Zea mays

<220>
<221> unsure
<222> (1)..(300)
<223> n=unknown

<400> 26
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aaaaaagtta atttaatctc acattctatg ggtggattgc tagtacgctg cttcatgtct 120
atgaatcatg atgtgagttt tcatgttttc tgtgtttttt ttgcttttgc ataaatatcc 180
atgtcaattt cccccatttt ctaggtattc actangtatg tcaacaaatg gatttgcatt 240
gcttgtccat tccaaggtaa cttatgggac atttcaattg tttattanat natggggncc 300

<210> 27
<211> 1240
<212> DNA
<213> Zea mays

<220>
<221> unsure
<222> (1)..(1240)
<223> n=unknown

<400> 27
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aagascaata ttgaactcat agwagsgaca aatggtggaa atagggtggt ggkmgatccc 180
acnactccat ggggtcnttn attttntgcn ttttacgnaa tggntcgaag ccctcctccg 240
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gacctcacaa gtccagggag tgcctgcctc tgatcctcat catcaggaac aggctcaagt 540
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gtttagattt agagtacact atgtatgcgt tgtttgacaa tgcttattta tgatatattg 1200
aatggtactt atttatatta attaattaaa aaaaaaaaa 1240

<210> 28
<211> 324
<212> DNA
<213> Zea mays

<400> 28
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catatgtcta taagttggcc ccacaggcag aatgttatat acctttccga attgacacct 120
cggctgaagg cggggaggaa aatagctgct tgaaaggggg tgtttactta gccgatggtg 180
atgaaactgt tccagttctt agtgcgggct acatgtgtgc aaaaggatgg cgtggcaaaa 240
ctcgtttcaa ccctgccggc agcaagactt acgtgagaga atacagccat tcaccaccct 300
ctactctcct ggaaggcagg ggca 324

<210> 29
<211> 254
<212> DNA
<213> Zea mays

<400> 29
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tcccacactc catgggggtc ctctattttt tgcattttat gaaatgggtc gaagcacctc 120
ctcccatggg gggtggcggt ggtccagact ggtgtgagaa gcatattaaa gctgtaatga 180
atattggagg acctttctta ggagttccta aggctgttgc tggccttttc tcatctgaag 240
ccaaagatgt tgcc 254

<210> 30
<211> 518
<212> DNA
<213> Mus musculus

<400> 30
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gtcaacctgt ggtgccgcca gcgaagggtc aaggccaaag ctgtctctac agggaagaag 180
gtcagtgggg ctgctgcgag caagctgtga gctatccaga caacctgacc taccgagatc 240
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aagtggggct gatccaacag tggatggtcc ctactatcca gaactccatg gaagcccttt 420
caagagcttc tgcagttttg gagaccgcga gttctacaga gattggtgga atgctgagtc 480
tgtcaccgac ttttggcaga actggaatat ccccgtgg 518

<210> 31
<211> 299
<212> DNA
<213> Mus musculus

<400> 31
ccatgatggc tcaggtccca ctggcctgga ttgtgggccg attcttccaa gggaactatg 60
gcaatgcagc tgtgtgggtg acactcatca ttgggcaacc ggtggctgtc tcatgtatgt 120
ccacgactac tacgtgctca actacgatgc cccagtgggt catgagctac tgccaaaggc 180
agccctccct aacctgggcc tggagttctg gaggggttcc tggctgcctg cacactcctc 240
ctagtctggg aggcctctct gcccctatgc gctactcctg ctcttgggga tggcatttg 299

<210> 32
<211> 1895
<212> DNA

<213> Artificial Sequence

<220>
<223> Description of Artificial Sequence: Inferred cDNA sequence

<220>
<221> unsure
<222> (1)..(1895)
<223> n=unknown

<400> 32
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ggctgccgcc ctgagctcgg gctgggacgt gcccagaggt gttgggagga tctggggtga 180
gtaccctgtg gccaggacta aaggggctnc accctcctgt ccatccctcg cagatcttga 240
gcaatgcccg gttatttctg gagaacctca tcaagtatgg catcctggtg gaccccatcc 300
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<210> 33
<211> 1766

<212> DNA
<213> Artificial Sequence

<220>
<223> Description of Artificial Sequence: Inferred cDNA sequence

<400> 33
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agctatctaa aaaaaaaaa aaaaaa 1766

<210> 34
<211> 409
<212> PRT
<213> Homo sapiens

<400> 34
Arg Arg Ser Leu Leu Asp Glu Leu Leu Glu Val Asp His Ile Arg Thr
1 5 10 15

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Ile Tyr His Met Phe Ile Ala Leu Leu Ile Leu Phe Ile Leu Ser Thr
            20                  25                  30

Leu Val Val Asp Tyr Ile Asp Glu Gly Arg Leu Val Leu Glu Phe Ser
        35                  40                  45

Leu Leu Ser Tyr Ala Phe Gly Lys Phe Pro Thr Val Val Trp Thr Trp
    50                  55                  60

Trp Ile Met Phe Leu Ser Thr Phe Ser Val Pro Tyr Phe Leu Phe Gln
65                  70                  75                  80

His Trp Arg Thr Gly Tyr Ser Lys Ser Ser His Pro Leu Ile Arg Ser
                85                  90                  95

Leu Phe His Gly Phe Leu Phe Met Ile Phe Gln Ile Gly Val Leu Gly
            100                 105                 110

Phe Gly Pro Thr Tyr Val Val Leu Ala Tyr Thr Leu Pro Pro Ala Ser
        115                 120                 125

Arg Phe Ile Ile Ile Phe Glu Gln Ile Arg Phe Val Met Lys Ala His
    130                 135                 140

Ser Phe Val Arg Glu Asn Val Pro Arg Val Leu Asn Ser Ala Lys Glu
145                 150                 155                 160

Lys Ser Ser Thr Val Pro Ile Pro Thr Val Asn Gln Tyr Leu Tyr Phe
                165                 170                 175

Leu Phe Ala Pro Thr Leu Ile Tyr Arg Asp Ser Tyr Pro Arg Asn Pro
            180                 185                 190

Thr Val Arg Trp Gly Tyr Val Ala Met Lys Phe Ala Gln Val Phe Gly
        195                 200                 205

Cys Phe Phe Tyr Val Tyr Tyr Ile Phe Glu Arg Leu Cys Ala Pro Leu
    210                 215                 220

Phe Arg Asn Ile Lys Gln Glu Pro Phe Ser Ala Arg Val Leu Val Leu
225                 230                 235                 240

Cys Val Phe Asn Ser Ile Leu Pro Gly Val Leu Ile Leu Phe Leu Thr
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Phe Phe Ala Phe Leu His Cys Trp Leu Asn Ala Phe Ala Glu Met Leu
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Arg Phe Gly Asp Arg Met Phe Tyr Lys Asp Trp Trp Asn Ser Thr Ser
        275                 280                 285

Tyr Ser Asn Tyr Tyr Arg Thr Trp Asn Val Val Val His Asp Trp Leu
    290                 295                 300

Tyr Tyr Tyr Ala Tyr Lys Asp Phe Leu Trp Phe Phe Ser Lys Arg Phe
305                 310                 315                 320

Lys Ser Ala Ala Met Leu Ala Val Phe Ala Val Ser Ala Val Val His
                325                 330                 335

Glu Tyr Ala Leu Ala Val Cys Leu Ser Phe Phe Tyr Pro Val Leu Phe
            340                 345                 350

Val Leu Phe Met Phe Phe Gly Met Ala Phe Asn Phe Ile Val Asn Asp
        355                 360                 365

Ser Arg Lys Lys Pro Ile Trp Asn Val Leu Met Trp Thr Ser Leu Phe
    370                 375                 380

Leu Gly Asn Gly Val Leu Leu Cys Phe Tyr Ser Gln Glu Trp Tyr Ala
385                 390                 395                 400

Arg Arg His Cys Pro Leu Lys Asn Pro
                405
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<210> 35
<211> 409
<212> PRT
<213> Mus musculus

<400> 35

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Arg Gln Ser Leu Leu Asp Glu Leu Phe Glu Val Asp His Ile Arg Thr
  1               5                  10                  15

Ile Tyr His Met Phe Ile Ala Leu Leu Ile Leu Phe Val Leu Ser Thr
             20                  25                  30

Ile Val Val Asp Tyr Ile Asp Glu Gly Arg Leu Val Leu Glu Phe Asn
         35                  40                  45

Leu Leu Ala Tyr Ala Phe Gly Lys Phe Pro Thr Val Ile Trp Thr Trp
     50                  55                  60

Trp Ala Met Phe Leu Ser Thr Leu Ser Ile Pro Tyr Phe Leu Phe Gln
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65 70 75 80

Pro Trp Ala His Gly Tyr Ser Lys Ser Ser His Pro Leu Ile Tyr Ser
85 90 95

Leu Val His Gly Leu Leu Phe Leu Val Phe Gln Leu Gly Val Leu Gly
100 105 110

Phe Val Pro Thr Tyr Val Val Leu Ala Tyr Thr Leu Pro Pro Ala Ser
115 120 125

Arg Phe Ile Leu Ile Leu Glu Gln Ile Arg Leu Ile Met Lys Ala His
130 135 140

Ser Phe Val Arg Glu Asn Ile Pro Arg Val Leu Asn Ala Ala Lys Glu
145 150 155 160

Lys Ser Ser Lys Asp Pro Leu Pro Thr Val Asn Gln Tyr Leu Tyr Phe
165 170 175

Leu Phe Ala Pro Thr Leu Ile Tyr Arg Asp Asn Tyr Pro Arg Thr Pro
180 185 190

Thr Val Arg Trp Gly Tyr Val Ala Met Gln Phe Leu Gln Val Phe Gly
195 200 205

Cys Leu Phe Tyr Val Tyr Tyr Ile Phe Glu Arg Leu Cys Ala Pro Leu
210 215 220

Phe Arg Asn Ile Lys Gln Glu Pro Phe Ser Ala Arg Val Leu Val Leu
225 230 235 240

Cys Val Phe Asn Ser Ile Leu Pro Gly Val Leu Ile Leu Phe Leu Ser
245 250 255

Phe Phe Ala Phe Leu His Cys Trp Leu Asn Ala Phe Ala Glu Met Leu
260 265 270

Arg Phe Gly Asp Arg Met Phe Tyr Lys Asp Trp Trp Asn Ser Thr Ser
275 280 285

Tyr Ser Asn Tyr Tyr Arg Thr Trp Asn Val Val Val His Asp Trp Leu
290 295 300

Tyr Tyr Tyr Val Tyr Lys Asp Leu Leu Trp Phe Phe Ser Lys Arg Phe
305 310 315 320

Lys Ser Ala Ala Met Leu Ala Val Phe Ala Leu Ser Ala Val Val His

325 330 335

Glu Tyr Ala Leu Ala Ile Cys Leu Ser Tyr Phe Tyr Pro Val Leu Phe
340 345 350

Val Leu Phe Met Phe Phe Gly Met Ala Phe Asn Phe Ile Val Asn Asp
355 360 365

Ser Arg Lys Arg Pro Ile Trp Asn Ile Met Val Trp Ala Ser Leu Phe
370 375 380

Leu Gly Tyr Gly Leu Ile Leu Cys Phe Tyr Ser Gln Glu Trp Tyr Ala
385 390 395 400

Arg Gln His Cys Pro Leu Lys Asn Pro
405

<210> 36
<211> 429
<212> PRT
<213> Saccharomyces cerevisiae

<400> 36
Asp Lys Ala Asp Ala Pro Pro Gly Glu Lys Leu Glu Ser Asn Phe Ser
1 5 10 15

Gly Ile Tyr Val Phe Ala Trp Met Phe Leu Gly Trp Ile Ala Ile Arg
20 25 30

Cys Cys Thr Asp Tyr Tyr Ala Ser Tyr Gly Ser Ala Trp Asn Lys Leu
35 40 45

Glu Ile Val Gln Tyr Met Thr Thr Asp Leu Phe Thr Ile Ala Met Leu
50 55 60

Asp Leu Ala Met Phe Leu Cys Thr Phe Phe Val Val Phe Val His Trp
65 70 75 80

Leu Val Lys Lys Arg Ile Ile Asn Trp Lys Trp Thr Gly Phe Val Ala
85 90 95

Val Ser Ile Phe Glu Leu Ala Phe Ile Pro Val Thr Phe Pro Ile Tyr
100 105 110

Val Tyr Tyr Phe Asp Phe Asn Trp Val Thr Arg Ile Phe Leu Phe Leu
115 120 125

```
His Ser Val Val Phe Val Met Lys Ser His Ser Phe Ala Phe Tyr Asn
    130                 135                 140

Gly Tyr Leu Trp Asp Ile Lys Gln Glu Leu Glu Tyr Ser Ser Lys Gln
145                 150                 155                 160

Leu Gln Lys Tyr Lys Glu Ser Leu Ser Pro Glu Thr Arg Glu Ile Leu
                165                 170                 175

Gln Lys Ser Cys Asp Phe Cys Leu Phe Glu Leu Asn Tyr Gln Thr Lys
            180                 185                 190

Asp Asn Asp Phe Pro Asn Asn Ile Ser Cys Ser Asn Phe Phe Met Phe
        195                 200                 205

Cys Leu Phe Pro Val Leu Val Tyr Gln Ile Asn Tyr Pro Arg Thr Ser
    210                 215                 220

Arg Ile Arg Trp Arg Tyr Val Leu Glu Lys Val Cys Ala Ile Ile Gly
225                 230                 235                 240

Thr Ile Phe Leu Met Met Val Thr Ala Gln Phe Phe Met His Pro Val
                245                 250                 255

Ala Met Arg Cys Ile Gln Phe His Asn Thr Pro Thr Phe Gly Gly Trp
            260                 265                 270

Ile Pro Ala Thr Gln Glu Trp Phe His Leu Leu Phe Asp Met Ile Pro
        275                 280                 285

Gly Phe Thr Val Leu Tyr Met Leu Thr Phe Tyr Met Ile Trp Asp Ala
    290                 295                 300

Leu Leu Asn Cys Val Ala Glu Leu Thr Arg Phe Ala Asp Arg Tyr Phe
305                 310                 315                 320

Tyr Gly Asp Trp Trp Asn Cys Val Ser Phe Glu Glu Phe Ser Arg Ile
                325                 330                 335

Trp Asn Val Pro Val His Lys Phe Leu Leu Arg His Val Tyr His Ser
            340                 345                 350

Ser Met Gly Ala Leu His Leu Ser Lys Ser Gln Ala Thr Leu Phe Thr
        355                 360                 365

Phe Phe Leu Ser Ala Val Phe His Glu Met Ala Met Phe Ala Ile Phe
    370                 375                 380
```

```
Arg Arg Val Arg Gly Tyr Leu Phe Met Phe Gln Leu Ser Gln Phe Val
385                 390                 395                 400

Trp Thr Ala Leu Ser Asn Thr Lys Phe Leu Arg Ala Arg Pro Gln Leu
                405                 410                 415

Ser Asn Val Val Phe Ser Phe Gly Val Cys Ser Gly Pro
            420                 425


<210> 37
<211> 432
<212> PRT
<213> Saccharomyces cerevisiae

<400> 37
Glu Thr Val Val Thr Val Glu Thr Thr Ile Ile Ser Ser Asn Phe Ser
  1               5                  10                  15

Gly Leu Tyr Val Ala Phe Trp Met Ala Ile Ala Phe Gly Ala Val Lys
            20                  25                  30

Ala Leu Ile Asp Tyr Tyr Tyr Gln His Asn Gly Ser Phe Lys Asp Ser
        35                  40                  45

Glu Ile Leu Lys Phe Met Thr Thr Asn Leu Phe Thr Val Ala Ser Val
    50                  55                  60

Asp Leu Leu Met Tyr Leu Ser Thr Tyr Phe Val Val Gly Ile Gln Tyr
65                  70                  75                  80

Leu Cys Lys Trp Gly Val Leu Lys Trp Gly Thr Thr Gly Trp Ile Phe
                85                  90                  95

Thr Ser Ile Tyr Glu Phe Leu Phe Val Ile Phe Tyr Met Tyr Leu Thr
            100                 105                 110

Glu Asn Ile Leu Lys Leu His Trp Leu Ser Lys Ile Phe Leu Phe Leu
        115                 120                 125

His Ser Leu Val Leu Leu Met Lys Met His Ser Phe Ala Phe Tyr Asn
    130                 135                 140

Gly Tyr Leu Trp Gly Ile Lys Glu Glu Leu Gln Phe Ser Lys Ser Ala
145                 150                 155                 160

Leu Ala Lys Tyr Lys Asp Ser Ile Asn Asp Pro Lys Val Ile Gly Ala
                165                 170                 175
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Leu Glu Lys Ser Cys Glu Phe Cys Ser Phe Glu Leu Ser Ser Gln Ser
180 185 190

Leu Ser Asp Gln Thr Gln Lys Phe Pro Asn Asn Ile Ser Ala Lys Ser
195 200 205

Phe Phe Trp Phe Thr Met Phe Pro Thr Leu Ile Tyr Gln Ile Glu Tyr
210 215 220

Pro Arg Thr Lys Glu Ile Arg Trp Ser Tyr Val Leu Glu Lys Ile Cys
225 230 235 240

Ala Ile Phe Gly Thr Ile Phe Leu Met Met Ile Asp Ala Gln Ile Leu
245 250 255

Met Tyr Pro Val Ala Met Arg Ala Leu Ala Val Arg Asn Ser Glu Trp
260 265 270

Thr Gly Ile Leu Asp Arg Leu Leu Lys Trp Val Gly Leu Leu Val Asp
275 280 285

Ile Val Pro Gly Phe Ile Val Met Tyr Ile Leu Asp Phe Tyr Leu Ile
290 295 300

Trp Asp Ala Ile Leu Asn Cys Val Ala Glu Leu Thr Arg Phe Gly Asp
305 310 315 320

Arg Tyr Phe Tyr Gly Asp Trp Trp Asn Cys Val Ser Trp Ala Asp Phe
325 330 335

Ser Arg Ile Trp Asn Ile Pro Val His Lys Phe Leu Leu Arg His Val
340 345 350

Tyr His Ser Ser Met Ser Ser Phe Lys Leu Asn Lys Ser Gln Ala Thr
355 360 365

Leu Met Thr Phe Phe Leu Ser Ser Val Val His Glu Leu Ala Met Tyr
370 375 380

Val Ile Phe Lys Lys Leu Arg Phe Tyr Leu Phe Phe Phe Gln Met Leu
385 390 395 400

Gln Met Pro Leu Val Ala Leu Thr Asn Thr Lys Phe Met Arg Asn Arg
405 410 415

Thr Ile Ile Gly Asn Val Ile Phe Trp Leu Gly Ile Cys Met Gly Pro
420 425 430

<210> 38
<211> 1942
<212> DNA
<213> Arabidopsis thaliana

<400> 38
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tccgctcttt ccctctccat tagattctgt ttcctctttc aatttcttct gcatgcttct 180
cgattctctc tgacgcctct tttctcccga cgctgtttcg tcaaacgctt ttcgaaatgg 240
cgattttgga ttctgctggc gttactacgg tgacggagaa cggtggcgga gagttcgtcg 300
atcttgatag gcttcgtcga cggaaatcga gatcggattc ttctaacgga cttcttctct 360
ctggttccga taataattct ccttcggatg atgttggagc tcccgccgac gttagggatc 420
ggattgattc cgttgttaac gatgacgctc agggaacagc caatttggcc ggagataata 480
acggtggtgg cgataataac ggtggtggaa gaggcggcgg agaaggaaga ggaaacgccg 540
atgctacgtt tacgtatcga ccgtcggttc cagctcatcg gagggcgaga gagagtccac 600
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ccggattcat gggatttata atagaacaat atataaatcc tattgtcagg aactcaaagc 1260
atcctttgaa aggcgatctt ctatatgcta ttgaaagagt gttgaagctt tcagttccaa 1320
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cagagcttct ctgcttcggg gatcgtgaat tctacaaaga ttggtggaat gcaaaaagtg 1440
tgggagatta ctggagaatg tggaatatgc ctgttcataa atggatggtt cgacatatat 1500
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acaactgttc aaaaaatgac tttcttcaaa catctatggc ctcgttggat ctccgttgat 1860
gttgtggtgg ttctgatgct aaaacgacaa atagtgttat aaccattgaa gaagaaaaga 1920
caattagagt tgttgtatcg ca 1942

<210> 39
<211> 520

<212> PRT
<213> Arabidopsis thaliana

<400> 39

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Met Ala Ile Leu Asp Ser Ala Gly Val Thr Thr Val Thr Glu Asn Gly
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Gly Gly Glu Phe Val Asp Leu Asp Arg Leu Arg Arg Arg Lys Ser Arg
             20                  25                  30

Ser Asp Ser Ser Asn Gly Leu Leu Leu Ser Gly Ser Asp Asn Asn Ser
         35                  40                  45

Pro Ser Asp Asp Val Gly Ala Pro Ala Asp Val Arg Asp Arg Ile Asp
     50                  55                  60

Ser Val Val Asn Asp Asp Ala Gln Gly Thr Ala Asn Leu Ala Gly Asp
 65                  70                  75                  80

Asn Asn Gly Gly Gly Asp Asn Asn Gly Gly Gly Arg Gly Gly Gly Glu
                 85                  90                  95

Gly Arg Gly Asn Ala Asp Ala Thr Phe Thr Tyr Arg Pro Ser Val Pro
            100                 105                 110

Ala His Arg Arg Ala Arg Glu Ser Pro Leu Ser Ser Asp Ala Ile Phe
        115                 120                 125

Lys Gln Ser His Ala Gly Leu Phe Asn Leu Cys Val Val Val Leu Ile
    130                 135                 140

Ala Val Asn Ser Arg Leu Ile Ile Glu Asn Leu Met Lys Tyr Gly Trp
145                 150                 155                 160

Leu Ile Arg Thr Asp Phe Trp Phe Ser Ser Arg Ser Leu Arg Asp Trp
                165                 170                 175

Pro Leu Phe Met Cys Cys Ile Ser Leu Ser Ile Phe Pro Leu Ala Ala
            180                 185                 190

Phe Thr Val Glu Lys Leu Val Leu Gln Lys Tyr Ile Ser Glu Pro Val
        195                 200                 205

Val Ile Phe Leu His Ile Ile Ile Thr Met Thr Glu Val Leu Tyr Pro
    210                 215                 220

Val Tyr Val Thr Leu Arg Cys Asp Ser Ala Phe Leu Ser Gly Val Thr
225                 230                 235                 240
```

Leu Met Leu Leu Thr Cys Ile Val Trp Leu Lys Leu Val Ser Tyr Ala
245 250 255

His Thr Ser Tyr Asp Ile Arg Ser Leu Ala Asn Ala Ala Asp Lys Ala
260 265 270

Asn Pro Glu Val Ser Tyr Tyr Val Ser Leu Lys Ser Leu Ala Tyr Phe
275 280 285

Met Val Ala Pro Thr Leu Cys Tyr Gln Pro Ser Tyr Pro Arg Ser Ala
290 295 300

Cys Ile Arg Lys Gly Trp Val Ala Arg Gln Phe Ala Lys Leu Val Ile
305 310 315 320

Phe Thr Gly Phe Met Gly Phe Ile Ile Glu Gln Tyr Ile Asn Pro Ile
325 330 335

Val Arg Asn Ser Lys His Pro Leu Lys Gly Asp Leu Leu Tyr Ala Ile
340 345 350

Glu Arg Val Leu Lys Leu Ser Val Pro Asn Leu Tyr Val Trp Leu Cys
355 360 365

Met Phe Tyr Cys Phe Phe His Leu Trp Leu Asn Ile Leu Ala Glu Leu
370 375 380

Leu Cys Phe Gly Asp Arg Glu Phe Tyr Lys Asp Trp Trp Asn Ala Lys
385 390 395 400

Ser Val Gly Asp Tyr Trp Arg Met Trp Asn Met Pro Val His Lys Trp
405 410 415

Met Val Arg His Ile Tyr Phe Pro Cys Leu Arg Ser Lys Ile Pro Lys
420 425 430

Thr Leu Ala Ile Ile Ile Ala Phe Leu Val Ser Ala Val Phe His Glu
435 440 445

Leu Cys Ile Ala Val Pro Cys Arg Leu Phe Lys Leu Trp Ala Phe Leu
450 455 460

Gly Ile Met Phe Gln Val Pro Leu Val Phe Ile Thr Asn Tyr Leu Gln
465 470 475 480

Glu Arg Phe Gly Ser Thr Val Gly Asn Met Ile Phe Trp Phe Ile Phe
485 490 495

Cys Ile Phe Gly Gln Pro Met Cys Val Leu Leu Tyr Tyr His Asp Leu
500 505 510

Met Asn Arg Lys Gly Ser Met Ser
515 520

<210> 40
<211> 29
<212> DNA
<213> Artificial Sequence

<220>
<223> Description of Artificial Sequence: Synthetic
oligonucleotide primer

<400> 40
tgcaaattga cgagcacacc aacccttc 29

<210> 41
<211> 28
<212> DNA
<213> Artificial Sequence

<220>
<223> Description of Artificial Sequence: Synthetic
oligonuclotide primer

<400> 41
aaggatgctt tgagttcctg acaatagg 28

<210> 42
<211> 1942
<212> DNA
<213> Arabidopsis thaliana

<400> 42
ctctcgtgaa tccttttttcc tttctttcttc ttcttctctt cagagaaaac tttgcttctc 60
tttctataag gaaccagaca cgaatcccat tcccaccgat ttcttagctt cttccttcaa 120
tccgctcttt ccctctccat tagattctgt ttcctctttc aatttcttct gcatgcttct 180
cgattctctc tgacgcctct tttctcccga cgctgtttcg tcaaacgctt ttcgaaatgg 240
cgattttgga ttctgctggc gttactacgg tgacggagaa cggtggcgga gagttcgtcg 300
atcttgatag gcttcgtcga cggaaatcga gatcggattc ttctaacgga cttcttctct 360
ctggttccga taataattct ccttcggatg atgttggagc tcccgccgac gttagggatc 420
ggattgattc cgttgttaac gatgacgctc agggaacagc caatttggcc ggagataata 480

acggtggtgg cgataataac ggtggtggaa gaggcggcgg agaaggaaga ggaaacgccg 540
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ttagctccga cgcaatcttc aaacagagcc atgccggatt attcaacctc tgtgtagtag 660
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atcctttgaa aggcgatctt ctatatgcta ttgaaagagt gttgaagctt tcagttccaa 1320
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tctctgcagt ctttcatgag ctatgcatcg cagttccttg tcgtctcttc aagctatggg 1620
cttttcttgg gattatgttt caggtgcctt tggtcttcat cacaaactat ctacaggaaa 1680
ggtttggctc aacggtgggg aacatgatct tctggttcat cttctgcatt ttcggacaac 1740
cgatgtgtgt gcttctttat taccacgacc tgatgaaccg aaaaggatcg atgtcatgaa 1800
acaactgttc aaaaaatgac tttcttcaaa catctatggc ctcgttggat ctccgttgat 1860
gttgtggtgg ttctgatgct aaaacgacaa atagtgttat aaccattgaa gaagaaaaga 1920
caattagagt tgttgtatcg ca 1942

<210> 43
<211> 234
<212> DNA
<213> Glycine max

<220>
<221> unsure
<222> (1)..(234)
<223> n=unknown

<400> 43
gtaagcttca agagcttagc atanttcctg gttgccccta ncattatgtt accagccaan 60
ctatcctcgc acaccttata ttcgaaaggg ttggctgttt cgccaacttg tcaactgata 120
atatttacag gagttatggg atttataata gaacaataca ttaatcccat tgtacaaaat 180
tcacagcatc ctctcaaggg aaaccttctt tacgccatcg agagagttct gaag 234

<210> 44
<211> 267
<212> DNA
<213> Glycine max

<400> 44
ctgcttttgt atctggtgtc acgttgatgc tattaacttg cattgtgtgg ttaaaattgg 60
tgtcatatgc acatacaaac tatgatatga gagcacttac tgtttcgaat gaaaagggag 120
aaacattacc caatactttg atatggagta tccgtacact gtgaccttca ggagtttggc 180
atacttcatg gttgctccta cattatgcta tcagacaagc tatcctcgca caccttcagt 240
tcgaaagggt tgggtgtttc gtcaact 267

<210> 45
<211> 275
<212> DNA
<213> Glycine max

<220>
<221> unsure
<222> (1)..(275)
<223> n=unknown

<400> 45
gtggaatgcc aaaactgttg aagattattg gaggatgtgg aatatgcctg ttcacaaatg 60
gatgatccgc cacctatatt ttccatgttt aaggcacggt ataccaaagg ccgttgctct 120
tttaattgcc ttcctgggttc tgctttattc catgagctgt gcatcgctgt tccttgccca 180
catattcaag tngtgggttt cngnggaatt nagtttcagg tnccttgggt ttcnaccnna 240
attnntnggc naaaaaattc cnngaacccc ggggg 275

<210> 46
<211> 257
<212> DNA
<213> Glycine max

<400> 46
aacggaattg agactccaga gaatatgcca aaatgtatta ataattgtca caacttggaa 60
ggcttttgga aaaactggca tgcttccttc aacaagtggc ttgtgaggta tatatacatt 120
cctcttgggg gatctaagaa aaagctacta aatgtgtggg ttgttttcac atttgttgca 180
atctggcatg atttagagtg gaagcttctt tcatgggcat ggttgacgtg tttattcttc 240
atccctgagt tggtttt 257

<210> 47
<211> 253
<212> DNA
<213> Zea mays

<400> 47
agaaaatgga acatgcctgt gcataaatgg attgttcgtc atatatattt tccttgcatg 60
cgaaatggta tatcaaagga agttgctgtt tttatatcgt tcttgtttct gctgtacttc 120

atgagttatg tgttgctgtt ccctgccaca tactcaagtt ctgggctttt tttaggaatc 180
atgcttcaga ttcccctcat catattgaca tcatacctca aaaataaatt cagtgacaca 240
atggttggca ata 253

<210> 48
<211> 254
<212> DNA
<213> Zea mays

<400> 48
tgaagtatgg cttattaata agatctggct tttggtttaa tgctacatca ttgcgagact 60
ggccactgct aatgtgttgc cttagtctac ccatatttcc ccttggtgca tttgcagtcg 120
aaaagttggc attcaacaat ctcattagtg atcctgctac tacctgtttt cacatccttt 180
ttacaacatt tgaaattgta tatccagtgc tcgtgattct taagtgtgat tctgcagttt 240
tatcaggctt tgtg 254

<210> 49
<211> 262
<212> DNA
<213> Zea mays

<400> 49
gaagtatggc ttattaataa gatctggctt ttggtttaat gctacatcat tgcgagactg 60
gccactgcta atgtgttgcc ttagtctacc catatttccc cttggtgcat ttgcagtcga 120
aaagttggca ttcaacaatc tcattagtga tcctgctact acctgttttc acatcctttt 180
tacaacattt gaaattgtat atccagtgct cgtgattctt aagtgtgatt ctgcagtttt 240
acaggctttg tgttgatgtt ta 262

<210> 50
<211> 325
<212> DNA
<213> Zea mays

<220>
<221> unsure
<222> (1)..(325)
<223> n=unknown

<400> 50
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aagatatctc catgctacgt tcaagcatgt aatggtgggc aacatgatan tttggntctn 180
cagtatagtc ggacagccga tgtnnnnnna tctatactac catgacgtca tgaacaggca 240
ggcccaggca agtagatagt ncggcagaga catgtacttc aacatcganc atcagnagca 300
nacngagcga gcggcangaa ncagc 325

<210> 51
<211> 519
<212> DNA
<213> Mortierrella alpina

<220>
<221> unsure
<222> (1)..(519)
<223> n=unknown

<400> 51
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ttanactnaa ttngaaaatn cnnncccaac ttnaggnact tnnagnnccc ccnacttgac 120
aacggagcac tatatttacc ccgtggtngt tcaacccagc catctcaccc ttgcgagcat 180
tggtgctgct cttgataccc ttcatgctta actatctcat gatcttttac atcattttcg 240
agtgcatctg caacgccttt gcggaactaa gttgctttgc ggatcgcaac ttttacgagg 300
attggtggaa ctgcgtcagc tttgatgagt gggcacgcaa atggaacaag cctgtgcaac 360
acttcttgct ccgccacgtg tacgactcga gcatccgagt ccttccactt gtccgaaatc 420
caatgccgcn aattgcaaac gttccttccc ggtcgtcaat gcgttcaacg aacctgggtg 480
aagaatgggt ggtgacaacg ttaaagtgcg cccggtatc 519

<210> 52
<211> 45
<212> DNA
<213> Artificial Sequence

<220>
<223> Description of Artificial Sequence:
Oligonucleotide primer

<400> 52
ggatccgcgg ccgcacaatg aaaaaaatat cttcacatta ttcgg 45

<210> 53
<211> 40
<212> DNA
<213> Artificial Sequence

<220>
<223> Description of Artificial Sequence:
Oligonucleotide primer

<400> 53
ggatcccctg caggtcattc attgacggca ttaacattgg 40

<210> 54
<211> 44
<212> DNA
<213> Artificial Sequence

<220>
<223> Description of Artificial Sequence: Synthetic
oligonucleotide primer

<400> 54
ggatccgcgg ccgcacaatg ggagcgaatt cgaaatcagt aacg 44

<210> 55
<211> 40
<212> DNA
<213> Artificial Sequence

<220>
<223> Description of Artificial Sequence: Synthetic
oligonucleotide primer

<400> 55
ggatcccctg caggttaata cccactttta tcaagctccc 40

<210> 56
<211> 41
<212> DNA
<213> Artificial Sequence

<220>
<223> Description of Artificial Sequence: Synthetic
oligonucleotide primer

<400> 56
ggatccgcgg ccgcacaatg tctctattac tggaagagat c 41

<210> 57
<211> 41
<212> DNA
<213> Artificial Sequence

<220>
<223> Description of Artificial Sequence: Synthetic

oligonucleotide primer

<400> 57
ggatcccctg caggttatgc atcaacagag acacttacag c 41

<210> 58
<211> 41
<212> DNA
<213> Artificial Sequence

<220>
<223> Description of Artificial Sequence: Synthetic
oligonucleotide primer

<400> 58
ggatccgcgg ccgcacaatg ggctggattc cgtgtccgtg c 41

<210> 59
<211> 38
<212> DNA
<213> Artificial Sequence

<220>
<223> Description of Artificial Sequence: Synthetic
oligonucleotide primer

<400> 59
ggatcccctg caggttaacc agaatcaact actttgtg 38

<210> 60
<211> 39
<212> DNA
<213> Artificial Sequence

<220>
<223> Description of Artificial Sequence:
Oligonucleotide primer

<400> 60
tcgacctgca ggaagcttag aaatggcgat tttggattc 39

<210> 61
<211> 36
<212> DNA

<213> Artificial Sequence

<220>
<223> Description of Artificial Sequence:
Oligonucleotide primer

<400> 61
ggatccgcgg ccgctcatga catcgatcct tttcgg 36

<210> 62
<211> 56
<212> DNA
<213> Artificial Sequence

<220>
<223> Description of Artificial Sequence: Annealed
oligonucleotide adapter

<400> 62
cgcgatttaa atggcgcgcc ctgcaggcgg ccgcctgcag ggcgcgccat ttaaat 56

<210> 63
<211> 32
<212> DNA
<213> Artificial Sequence

<220>
<223> Description of Artificial Sequence: Ligating
oligonucleotide

<400> 63
tcgaggatcc gcggccgcaa gcttcctgca gg 32

<210> 64
<211> 32
<212> DNA
<213> Artificial Sequence

<220>
<223> Description of Artificial Sequence: Ligating
oligonucleotide

<400> 64
tcgacctgca ggaagcttgc ggccgcggat cc 32

<210> 65
<211> 32
<212> DNA
<213> Artificial Sequence

<220>
<223> Description of Artificial Sequence: Ligating
oligonucleotide

<400> 65
tcgacctgca ggaagcttgc ggccgcggat cc 32

<210> 66
<211> 32
<212> DNA
<213> Artificial Sequence

<220>
<223> Description of Artificial Sequence: Ligating
oligonucleotide

<400> 66
tcgaggatcc gcggccgcaa gcttcctgca gg 32

<210> 67
<211> 36
<212> DNA
<213> Artificial Sequence

<220>
<223> Description of Artificial Sequence: Ligating
oligonucleotide

<400> 67
tcgaggatcc gcggccgcaa gcttcctgca ggagct 36

<210> 68
<211> 28
<212> DNA
<213> Artificial Sequence

<220>
<223> Description of Artificial Sequence: Ligating
oligonucleotide

<400> 68
cctgcaggaa gcttgcggcc gcggatcc 28

<210> 69
<211> 36
<212> DNA
<213> Artificial Sequence

<220>
<223> Description of Artificial Sequence: Ligating oligonucleotide

<400> 69
tcgacctgca ggaagcttgc ggccgcggat ccagct 36

<210> 70
<211> 28
<212> DNA
<213> Artificial Sequence

<220>
<223> Description of Artificial Sequence: Ligating oligonucleotide

<400> 70
ggatccgcgg ccgcaagctt cctgcagg 28

<210> 71
<211> 39
<212> DNA
<213> Artificial Sequence

<220>
<223> Description of Artificial Sequence: Ligating oligonucleotide

<400> 71
gatcacctgc aggaagcttg cggccgcgga tccaatgca 39

<210> 72
<211> 31
<212> DNA
<213> Artificial Sequence

<220>
<223> Description of Artificial Sequence: Ligating
oligonucleotide

<400> 72
ttggatccgc ggccgcaagc ttcctgcagg t 31

<210> 73
<211> 2013
<212> DNA
<213> Arabidopsis thaliana

<400> 73
atgcccctta ttcatcggaa aaagccgacg gagaaaccat cgacgccgcc atctgaagag 60
gtggtgcacg atgaggattc gcaaaagaaa ccacacgaat cttccaaatc ccaccataag 120
aaatcgaacg gaggagggaa gtggtcgtgc atcgattctt gttgttggtt cattgggtgt 180
gtgtgtgtaa cctggtggtt tcttctcttc ctttacaacg caatgcctgc gagcttccct 240
cagtatgtaa cggagcgaat cacgggtcct ttgcctgacc cgcccggtgt taagctcaaa 300
aaagaaggtc ttaaggcgaa acatcctgtt gtcttcattc ctgggattgt caccggtggg 360
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ggaacttttg gtgaagtcta caaaaggcct ctatgttggg tggaacacat gtcacttgac 480
aatgaaactg ggttggatcc agctggtatt agagttcgag ctgtatcagg actcgtggct 540
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aacacagagg tacgtgatca gactcttagc cgtatgaaaa gtaatataga gttgatggtt 720
tctaccaacg gtggaaaaaa agcagttata gttccgcatt ccatgggggt cttgtatttt 780
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aagacttata taagagaata caatcactct ccgccggcta acctgttgga agggcgcggg 1860
acgcagagtg gtgcccatgt tgatatcatg ggaaactttg ctttgatcga agatatcatg 1920
agggttgccg ccggaggtaa cgggtctgat ataggacatg accaggtcca ctctggcata 1980

tttgaatggt cggagcgtat tgacctgaag ctg 2013

<210> 74
<211> 671
<212> PRT
<213> Arabidopsis thaliana

<400> 74

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Met Pro Leu Ile His Arg Lys Lys Pro Thr Glu Lys Pro Ser Thr Pro
  1               5                  10                  15

Pro Ser Glu Glu Val Val His Asp Glu Asp Ser Gln Lys Lys Pro His
            20                  25                  30

Glu Ser Ser Lys Ser His His Lys Lys Ser Asn Gly Gly Gly Lys Trp
        35                  40                  45

Ser Cys Ile Asp Ser Cys Cys Trp Phe Ile Gly Cys Val Cys Val Thr
    50                  55                  60

Trp Trp Phe Leu Leu Phe Leu Tyr Asn Ala Met Pro Ala Ser Phe Pro
65                  70                  75                  80

Gln Tyr Val Thr Glu Arg Ile Thr Gly Pro Leu Pro Asp Pro Pro Gly
                85                  90                  95

Val Lys Leu Lys Lys Glu Gly Leu Lys Ala Lys His Pro Val Val Phe
            100                 105                 110

Ile Pro Gly Ile Val Thr Gly Gly Leu Glu Leu Trp Glu Gly Lys Gln
        115                 120                 125

Cys Ala Asp Gly Leu Phe Arg Lys Arg Leu Trp Gly Gly Thr Phe Gly
    130                 135                 140

Glu Val Tyr Lys Arg Pro Leu Cys Trp Val Glu His Met Ser Leu Asp
145                 150                 155                 160

Asn Glu Thr Gly Leu Asp Pro Ala Gly Ile Arg Val Arg Ala Val Ser
                165                 170                 175

Gly Leu Val Ala Ala Asp Tyr Phe Ala Pro Gly Tyr Phe Val Trp Ala
            180                 185                 190

Val Leu Ile Ala Asn Leu Ala His Ile Gly Tyr Glu Glu Lys Asn Met
        195                 200                 205
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Tyr Met Ala Ala Tyr Asp Trp Arg Leu Ser Phe Gln Asn Thr Glu Val
210 215 220

Arg Asp Gln Thr Leu Ser Arg Met Lys Ser Asn Ile Glu Leu Met Val
225 230 235 240

Ser Thr Asn Gly Gly Lys Lys Ala Val Ile Val Pro His Ser Met Gly
245 250 255

Val Leu Tyr Phe Leu His Phe Met Lys Trp Val Glu Ala Pro Ala Pro
260 265 270

Leu Gly Gly Gly Gly Gly Pro Asp Trp Cys Ala Lys Tyr Ile Lys Ala
275 280 285

Val Met Asn Ile Gly Gly Pro Phe Leu Gly Val Pro Lys Ala Val Ala
290 295 300

Gly Leu Phe Ser Ala Glu Ala Lys Asp Val Ala Val Ala Arg Ala Ile
305 310 315 320

Ala Pro Gly Phe Leu Asp Thr Asp Ile Phe Arg Leu Gln Thr Leu Gln
325 330 335

His Val Met Arg Met Thr Arg Thr Trp Asp Ser Thr Met Ser Met Leu
340 345 350

Pro Lys Gly Gly Asp Thr Ile Trp Gly Gly Leu Asp Trp Ser Pro Glu
355 360 365

Lys Gly His Thr Cys Cys Gly Lys Lys Gln Lys Asn Asn Glu Thr Cys
370 375 380

Gly Glu Ala Gly Glu Asn Gly Val Ser Lys Lys Ser Pro Val Asn Tyr
385 390 395 400

Gly Arg Met Ile Ser Phe Gly Lys Glu Val Ala Glu Ala Ala Pro Ser
405 410 415

Glu Ile Asn Asn Ile Asp Phe Arg Gly Ala Val Lys Gly Gln Ser Ile
420 425 430

Pro Asn His Thr Cys Arg Asp Val Trp Thr Glu Tyr His Asp Met Gly
435 440 445

Ile Ala Gly Ile Lys Ala Ile Ala Glu Tyr Lys Val Tyr Thr Ala Gly
450 455 460

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Glu Ala Ile Asp Leu Leu His Tyr Val Ala Pro Lys Met Met Ala Arg
465                 470                 475                 480

Gly Ala Ala His Phe Ser Tyr Gly Ile Ala Asp Asp Leu Asp Asp Thr
                485                 490                 495

Lys Tyr Gln Asp Pro Lys Tyr Trp Ser Asn Pro Leu Glu Thr Lys Leu
            500                 505                 510

Pro Asn Ala Pro Glu Met Glu Ile Tyr Ser Leu Tyr Gly Val Gly Ile
        515                 520                 525

Pro Thr Glu Arg Ala Tyr Val Tyr Lys Leu Asn Gln Ser Pro Asp Ser
    530                 535                 540

Cys Ile Pro Phe Gln Ile Phe Thr Ser Ala His Glu Glu Asp Glu Asp
545                 550                 555                 560

Ser Cys Leu Lys Ala Gly Val Tyr Asn Val Asp Gly Asp Glu Thr Val
                565                 570                 575

Pro Val Leu Ser Ala Gly Tyr Met Cys Ala Lys Ala Trp Arg Gly Lys
            580                 585                 590

Thr Arg Phe Asn Pro Ser Gly Ile Lys Thr Tyr Ile Arg Glu Tyr Asn
        595                 600                 605

His Ser Pro Pro Ala Asn Leu Leu Glu Gly Arg Gly Thr Gln Ser Gly
    610                 615                 620

Ala His Val Asp Ile Met Gly Asn Phe Ala Leu Ile Glu Asp Ile Met
625                 630                 635                 640

Arg Val Ala Ala Gly Gly Asn Gly Ser Asp Ile Gly His Asp Gln Val
                645                 650                 655

His Ser Gly Ile Phe Glu Trp Ser Glu Arg Ile Asp Leu Lys Leu
            660                 665                 670
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<210> 75
<211> 1986
<212> DNA
<213> Saccharomyces cerevisiae

<400> 75
atgggcacac tgtttcgaag aaatgtccag aaccaaaaga gtgattctga tgaaaacaat 60
aaaggggttt ctgttcataa caagcgagag agcagaaacc acattcatca tcaacaggga 120

ttaggccata agagaagaag gggtattagt ggcagtgcaa aaagaaatga gcgtggcaaa 180
gatttcgaca ggaaaagaga cgggaacggt agaaaacgtt ggagagattc cagaagactg 240
attttcattc ttggtgcatt cttaggtgta cttttgccgt ttagctttgg cgcttatcat 300
gttcataata gcgatagcga cttgtttgac aactttgtaa attttgattc acttaaagtg 360
tatttggatg attggaaaga tgttctccca caaggtataa gttcgtttat tgatgatatt 420
caggctggta actactccac atcttcttta gatgatctca gtgaaaattt tgccgttggt 480
aaacaactct tacgtgatta taatatcgag gccaaacatc ctgttgtaat ggttcctggt 540
gtcatttcta cgggaattga aagctgggga gttattggag acgatgagtg cgatagttct 600
gcgcattttc gtaaacggct gtggggaagt ttttacatgc tgagaacaat ggttatggat 660
aaagtttgtt ggttgaaaca tgtaatgtta gatcctgaaa caggtctgga cccaccgaac 720
tttacgctac gtgcagcaca gggcttcgaa tcaactgatt atttcatcgc agggtattgg 780
atttggaaca aagttttcca aaatctggga gtaattggct atgaacccaa taaaatgacg 840
agtgctgcgt atgattggag gcttgcatat ttagatctag aaagacgcga taggtacttt 900
acgaagctaa aggaacaaat cgaactgttt catcaattga gtggtgaaaa agtttgttta 960
attggacatt ctatgggttc tcagattatc ttttacttta tgaaatgggt cgaggctgaa 1020
ggccctcttt acggtaatgg tggtcgtggc tgggttaacg aacacataga ttcattcatt 1080
aatgcagcag ggacgcttct gggcgctcca aaggcagttc cagctctaat tagtggtgaa 1140
atgaaagata ccattcaatt aaatacgtta gccatgtatg gtttggaaaa gttcttctca 1200
agaattgaga gagtaaaaat gttacaaacg tggggtggta taccatcaat gctaccaaag 1260
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aatttgacaa tgaaagacgc cattaacatg acattatcga tatcacctga atggctccaa 1440
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aatgagctac accacaagca ctggtcgaat ccaatggaag taccacttcc agaagctccc 1560
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aaacaccagc cagatcgatt tgatatacgt ggtggagcaa aaagcgccga acacgtagac 1860
atcctcggca gcgcggagtt gaacgattac atcttgaaaa ttgcaagcgg taatggcgat 1920
ctcgtcgagc cacgccaatt gtctaatttg agccagtggg tttctcagat gcccttccca 1980
atgtaa 1986

<210> 76
<211> 661
<212> PRT
<213> Saccharomyces cerevisiae

<400> 76
Met Gly Thr Leu Phe Arg Arg Asn Val Gln Asn Gln Lys Ser Asp Ser
1 5 10 15

Asp Glu Asn Asn Lys Gly Gly Ser Val His Asn Lys Arg Glu Ser Arg
20 25 30

Asn His Ile His His Gln Gln Gly Leu Gly His Lys Arg Arg Arg Gly
35 40 45

```
Ile Ser Gly Ser Ala Lys Arg Asn Glu Arg Gly Lys Asp Phe Asp Arg
     50                  55                  60

Lys Arg Asp Gly Asn Gly Arg Lys Arg Trp Arg Asp Ser Arg Arg Leu
 65                  70                  75                  80

Ile Phe Ile Leu Gly Ala Phe Leu Gly Val Leu Leu Pro Phe Ser Phe
                 85                  90                  95

Gly Ala Tyr His Val His Asn Ser Asp Ser Asp Leu Phe Asp Asn Phe
            100                 105                 110

Val Asn Phe Asp Ser Leu Lys Val Tyr Leu Asp Asp Trp Lys Asp Val
        115                 120                 125

Leu Pro Gln Gly Ile Ser Ser Phe Ile Asp Asp Ile Gln Ala Gly Asn
    130                 135                 140

Tyr Ser Thr Ser Ser Leu Asp Asp Leu Ser Glu Asn Phe Ala Val Gly
145                 150                 155                 160

Lys Gln Leu Leu Arg Asp Tyr Asn Ile Glu Ala Lys His Pro Val Val
                165                 170                 175

Met Val Pro Gly Val Ile Ser Thr Gly Ile Glu Ser Trp Gly Val Ile
            180                 185                 190

Gly Asp Asp Glu Cys Asp Ser Ser Ala His Phe Arg Lys Arg Leu Trp
        195                 200                 205

Gly Ser Phe Tyr Met Leu Arg Thr Met Val Met Asp Lys Val Cys Trp
    210                 215                 220

Leu Lys His Val Met Leu Asp Pro Glu Thr Gly Leu Asp Pro Pro Asn
225                 230                 235                 240

Phe Thr Leu Arg Ala Ala Gln Gly Phe Glu Ser Thr Asp Tyr Phe Ile
                245                 250                 255

Ala Gly Tyr Trp Ile Trp Asn Lys Val Phe Gln Asn Leu Gly Val Ile
            260                 265                 270

Gly Tyr Glu Pro Asn Lys Met Thr Ser Ala Ala Tyr Asp Trp Arg Leu
        275                 280                 285

Ala Tyr Leu Asp Leu Glu Arg Arg Asp Arg Tyr Phe Thr Lys Leu Lys
    290                 295                 300
```

```
Glu Gln Ile Glu Leu Phe His Gln Leu Ser Gly Glu Lys Val Cys Leu
305                 310                 315                 320

Ile Gly His Ser Met Gly Ser Gln Ile Ile Phe Tyr Phe Met Lys Trp
                325                 330                 335

Val Glu Ala Glu Gly Pro Leu Tyr Gly Asn Gly Gly Arg Gly Trp Val
            340                 345                 350

Asn Glu His Ile Asp Ser Phe Ile Asn Ala Ala Gly Thr Leu Leu Gly
        355                 360                 365

Ala Pro Lys Ala Val Pro Ala Leu Ile Ser Gly Glu Met Lys Asp Thr
    370                 375                 380

Ile Gln Leu Asn Thr Leu Ala Met Tyr Gly Leu Glu Lys Phe Phe Ser
385                 390                 395                 400

Arg Ile Glu Arg Val Lys Met Leu Gln Thr Trp Gly Gly Ile Pro Ser
                405                 410                 415

Met Leu Pro Lys Gly Glu Glu Val Ile Trp Gly Asp Met Lys Ser Ser
            420                 425                 430

Ser Glu Asp Ala Leu Asn Asn Asn Thr Asp Thr Tyr Gly Asn Phe Ile
        435                 440                 445

Arg Phe Glu Arg Asn Thr Ser Asp Ala Phe Asn Lys Asn Leu Thr Met
    450                 455                 460

Lys Asp Ala Ile Asn Met Thr Leu Ser Ile Ser Pro Glu Trp Leu Gln
465                 470                 475                 480

Arg Arg Val His Glu Gln Tyr Ser Phe Gly Tyr Ser Lys Asn Glu Glu
                485                 490                 495

Glu Leu Arg Lys Asn Glu Leu His His Lys His Trp Ser Asn Pro Met
            500                 505                 510

Glu Val Pro Leu Pro Glu Ala Pro His Met Lys Ile Tyr Cys Ile Tyr
        515                 520                 525

Gly Val Asn Asn Pro Thr Glu Arg Ala Tyr Val Tyr Lys Glu Glu Asp
    530                 535                 540

Asp Ser Ser Ala Leu Asn Leu Thr Ile Asp Tyr Glu Ser Lys Gln Pro
545                 550                 555                 560
```

Val Phe Leu Thr Glu Gly Asp Gly Thr Val Pro Leu Val Ala His Ser
565 570 575

Met Cys His Lys Trp Ala Gln Gly Ala Ser Pro Tyr Asn Pro Ala Gly
580 585 590

Ile Asn Val Thr Ile Val Glu Met Lys His Gln Pro Asp Arg Phe Asp
595 600 605

Ile Arg Gly Gly Ala Lys Ser Ala Glu His Val Asp Ile Leu Gly Ser
610 615 620

Ala Glu Leu Asn Asp Tyr Ile Leu Lys Ile Ala Ser Gly Asn Gly Asp
625 630 635 640

Leu Val Glu Pro Arg Gln Leu Ser Asn Leu Ser Gln Trp Val Ser Gln
645 650 655

Met Pro Phe Pro Met
660

<210> 77
<211> 35
<212> DNA
<213> Artificial Sequence

<220>
<223> Description of Artificial Sequence: Synthetic oligonucleotide primer

<400> 77
ggatccgcgg ccgcacaatg ccccttattc atcgg 35

<210> 78
<211> 35
<212> DNA
<213> Artificial Sequence

<220>
<223> Description of Artificial Sequence: Synthetic oligonucleotide primer

<400> 78
ggatcccctg caggtcacag cttcaggtca atacg 35

<210> 79
<211> 37
<212> DNA
<213> Artificial Sequence

<220>
<223> Description of Artificial Sequence: Synthetic
oligonucleotide primer

<400> 79
ggatccgcgg ccgcacaatg ggcacactct ttcgaag 37

<210> 80
<211> 39
<212> DNA
<213> Artificial Sequence

<220>
<223> Description of Artificial Sequence: Synthetic
oligonucleotide primer

<400> 80
ggatcccctg caggttacat tgggcacact gtttcgaag 39